

Interannual variation in nitrous oxide emissions from perennial ryegrass/white clover grassland used for dairy production

WILLIAM BURCHILL^{1,2}, DEJUN LI³, GARY J. LANIGAN⁴, MICHEAL WILLIAMS² and JAMES HUMPHREYS¹

¹Animal & Grassland Research and Innovation Centre, Teagasc, Moorepark, Fermoy, Co. Cork, Ireland, ²Department of Botany, School of Natural Sciences, Trinity College Dublin, College Green, Dublin 2, Ireland, ³Huanjiang Observation and Research Station for Karst Ecosystems, Key Laboratory of Agro-ecological Processes in Subtropical Region, Institute of Subtropical Agriculture, Chinese Academy of Sciences, Changsha, Hunan 410125, China, ⁴Johnstown Castle Environment Research Centre, Teagasc, Johnstown Castle, Co. Wexford, Ireland

Abstract

Nitrous oxide (N₂O) emissions are subject to intra- and interannual variation due to changes in weather and management. This creates significant uncertainties when quantifying estimates of annual N₂O emissions from grazed grasslands. Despite these uncertainties, the majority of studies are short-term in nature (<1 year) and as a consequence, there is a lack of data on interannual variation in N₂O emissions. The objectives of this study were to (i) quantify annual N₂O emissions and (ii) assess the causes of interannual variation in emissions from grazed perennial ryegrass/white clover grassland. Nitrous oxide emissions were measured from fertilized and grazed perennial ryegrass/white clover grassland (WC) and from perennial ryegrass plots that were not grazed and did not receive N input (GB), over 4 years from 2008 to 2012 in Ireland (52°51'N, 08°21'W). The annual N₂O-N emissions (kg ha⁻¹; mean ± SE) ranged from 4.4 ± 0.2 to 34.4 ± 5.5 from WC and from 1.7 ± 0.8 to 6.3 ± 1.2 from GB. Interannual variation in N₂O emissions was attributed to differences in annual rainfall, monthly (December) soil temperatures and variation in N input. Such substantial interannual variation in N₂O emissions highlights the need for long-term studies of emissions from managed pastoral systems.

Keywords: freeze-thaw cycles, grazing, interannual variation, nitrous oxide, rainfall, regulating factors, white clover grassland

Received 25 July 2013 and accepted 27 February 2014

Introduction

Nitrous oxide (N₂O) is a potent greenhouse gas (GHG) and agriculture has been identified as a major source of N₂O accounting for 58% of global emissions, with direct emissions of N₂O from grazed grassland systems comprising between 16% and 33% of this total (de Klein *et al.*, 2008).

Nitrous oxide is primarily produced in soil by microbial processes such as nitrification and denitrification. Other processes such as nitrifier denitrification, codenitrification and dissimilatory reduction of nitrate to ammonium (DNRA) are also known to produce N₂O (Stevens & Laughlin, 1998). Both nitrification and denitrification are regulated by soil nitrate (NO₃⁻), ammonium (NH₄⁺), organic carbon (C), pH, temperature, moisture content and oxygen supply (Firestone & Davidson, 1989; Liu *et al.*, 2007; Cuhel *et al.*, 2010). Climatic conditions can directly influence these factors

and, hence, N₂O emissions (Cantarel *et al.*, 2012). For example, rainfall has a direct impact on soil moisture and oxygen status and can consequently affect N₂O production (Luo *et al.*, 1999). Changes in these controlling factors over time create considerable temporal variation in emissions of N₂O on a seasonal and annual basis, making it difficult to acquire representative estimates of annual N₂O emissions.

Considerable interannual variation in N₂O emissions has been observed in previous studies of temperate grassland (2 years) (Hyde *et al.*, 2006; Flechard *et al.*, 2007; Rafique *et al.*, 2011), native semi-arid grasslands (Du *et al.*, 2006), arable rotations (Cai *et al.*, 2013) and differing soil types (Webster & Dowdell, 1982). This variation can be driven by differences in (i) management such as stocking densities, N fertilization rates and drainage and (ii) climatic factors such as total precipitation and temperature (Du *et al.*, 2006; Flechard *et al.*, 2007; Cai *et al.*, 2013; Rees *et al.*, 2013). Despite this, the majority of N₂O studies are focused over relatively short periods (<1 year) and are insufficient to provide robust estimates of annual N₂O emissions or

Correspondence: William Burchill, tel. + 00353861926452 or 003532542685, fax 003532542340, e-mail: william.burchill@teagasc.ie

assess the variation that can occur over longer time periods.

This study measured N₂O emissions from fertilized and grazed perennial ryegrass/white clover grassland and nongrazed perennial ryegrass grassland receiving no N input over 4 years. The objectives were to (i) quantify annual N₂O emissions and (ii) access the causes of interannual variation in emissions from grazed perennial ryegrass/white clover grassland.

Materials and methods

Site description

This study was carried out at the Teagasc Solohead Research Farm (52°51'N, 08°21'W; 95 m above sea level). The predominant soils are poorly drained Gleys (90%) and Grey Brown Podzolics (10%) with a clay loam texture (29% clay, 36% silt and 34% sand) and low permeability. The soils are seasonally wet, waterlogged or flooded due to impeded drainage and a shallow water table depth (0–2.2 m below ground level) at the site. Mean soil organic carbon, total N content, bulk density and pH at 0–10 cm depth are 5.12%, 0.54%, 0.87 g cm⁻³ and 6.2, respectively. The permanent grassland at the site is used predominately for grazing and occasionally harvested for silage. The swards range from 20 to 30 years in age. This region has a temperate moist climate allowing for pasture-based dairy production with a grazing season extending to 9 or 10 months each year. Mean annual rainfall at the site (1997–2008) was 1031 mm. Mean minimum monthly soil temperature at 10 cm depth ranged from 4.3 to 8.5 °C during winter (November–January) and maximum monthly temperatures ranged from 13.9 to 18.2 °C in summer (May–July).

Experiment design and treatments

The experiment was a randomized block design with two treatments and three replicates. The treatments were (i) grazed perennial ryegrass/white clover-based pastures (WC) and (ii) background perennial ryegrass plots (GB). The WC treatment formed part of a perennial ryegrass (*Lolium perenne* L.)/white clover (*Trifolium repens* L.)-based system of dairy production at the Solohead Research Farm which consisted of six paddocks, with an annual stocking density of 2.35 cows ha⁻¹. Three of these paddocks were devoted to the WC treatment. The WC paddocks were rotationally grazed by spring calving Holstein-Friesian cows during the main grazing season (March–November) with surplus herbage occasionally removed as baled silage. Paddocks ranged in size from 1.6 to 2.1 ha with a total area of 5.4 ha and were dominated by perennial ryegrass with an annual average white clover content of herbage dry matter (DM) of 24% during the experimental period. The grazing rotation length on WC varied from 21 days in late spring/early summer to 42 days in autumn. Postgrazing sward height, maintained at 4 cm, was measured using a rising plate meter (Grasstec, Charleville, Ireland) and used to determine when cows were moved to the next paddock.

Fertilizer N and slurry applications to WC are presented in Table 1. Fertilizer and slurry N were applied in a number of applications between early spring and early summer. The additional N input to WC in the form of N deposited by grazing cows was calculated based on the stocking density, residence time and N excretion per cow per day for each grazing event (Table 1). Annual N excretion per cow was estimated as the difference between the cow's annual intake of N in feeds and the N output in milk and calves or in live-weight change of the cow as described in detail by Humphreys *et al.* (2008). Annual input of N for the WC paddocks due to biological N fixation was estimated to be (mean ± SE) 78.6 ± 13.2 kg N ha⁻¹ in 2008/09, 112.4 ± 8.8 kg N ha⁻¹ in 2009/10, 102.6 ± 10.7 kg N ha⁻¹ in 2010/11 and 65.4 ± 5.6 kg N ha⁻¹ in 2011/12 based on white clover contents and herbage yields using methodology described by Humphreys *et al.* (2008).

The GB treatment was laid down to measure background soil emissions of N₂O in the absence of N input. The area of each GB plot was 3 × 11 m and was dominated by perennial ryegrass and contained no white clover. The GB plots received no fertilizer N or cattle slurry and were not grazed. Herbage on GB was harvested by cutting during the growing season to a height of 4 cm, with clippings removed.

Both of these treatments were imposed from January 2008 and measurements began in October 2008. Therefore, each annual measurement spanned two calendar years and ran from 1st October 2008 to 30th September 2009 (2008/09), 1st

Table 1 Applications of fertilizer N and cattle slurry, and annual N deposited by grazing animals to perennial ryegrass/white clover pastures (WC) in 2008/09, 2009/10, 2010/11 and 2011/12

Date	Application	N input (kg ha ⁻¹)
2008/09		
9th February 2009	Cattle slurry	93.4
25th March 2009	Urea	28.8
8th May 2009	CAN*	34
	Excreta N†	129
2009/10		
10th February 2010	Cattle slurry	76.9
14th April 2010	Urea	57.5
	Excreta N	136
2010/11		
21st January 2011	Cattle slurry	102.9
24th February 2011	Urea	28.8
22nd March 2011	Urea	28.8
13th April 2011	Urea	28.8
	Excreta N	203
2011/12		
23rd January 2012	Cattle slurry	30.9
15th February 2012	Urea	28.8
13th March 2012	Urea	28.8
2nd May 2012	Urea	28.8
	Excreta N	171

*Calcium ammonium nitrate.

†Annual N deposited by grazing livestock.

October 2009 to 30th September 2010 (2009/10), 1st November 2010 to 31st October 2011 (2010/11) and 1st November 2011 to 31st October 2012 (2011/12).

Soil and weather measurements

Daily weather data including rainfall (mm), air temperature (°C) and soil temperature at 10 cm depth (°C) were recorded at the experimental site as described by Fitzgerald & Fitzgerald (2004). Effective rainfall (rainfall minus evapo-transpiration) was determined using a water-mass balance model described by Schulte *et al.* (2005). This model is based on a modified version of the Penman–Monteith evapo-transpiration model. Input weather data included daily maximum and minimum air temperatures, rainfall, wind speed and solar radiation.

At each gas sampling, volumetric soil water content (VWC) was recorded at 0–5 cm depth using a HH2 moisture meter in conjunction with a Theta Probe sensor (Delta-T Devices, Cambridge, UK). Soil water filled pore space (WFPS) at 0–5 cm depth was calculated according to Ri *et al.* (2003).

N₂O flux measurements

A closed static chamber technique (Hutchinson & Moiser, 1981) was used to measure N₂O fluxes. The chamber consisted of two parts: a permanent collar and a chamber, both of which were constructed from polyvinylchloride (PVC) pipe. The headspace height of the chamber was 40 cm with an internal diameter of 29.5 cm, resulting in a cover area of 0.0683 m² and a headspace volume of 0.0273 m³. A 10 cm long plastic tube (internal diameter of 0.5 cm) was permanently inserted into the top of each chamber. One port of a three-way valve (Discofix) was fitted to the top of this tube on the outside of the chamber. A second port of the valve was connected to a needle for extraction of gas samples from the chamber. The third port of the valve could be left open when placing the chamber on the collar to prevent the build-up of air pressure within the chamber. Once the chamber was in place, this port was closed to create an air tight seal between the chamber and collar. The collars were permanently inserted into the soil to a depth of 12 cm and extended 4 cm above the soil surface following installation. The collars on WC were exposed to grazing livestock throughout the experiment. Five chambers and collars were deployed in each paddock of WC and one in each of the smaller GB plots. The N₂O sampling was conducted (between 9:30 am and 1:00 pm) on a weekly basis with increased frequency (three times *per week*) following N applications on WC. Likewise, N₂O sampling was conducted (between 9:30 am and 1:00 pm) on a weekly basis on GB with increased frequency (three times *per week*) following N applications on WC in 2008/09 and 2009/10. Sampling frequency on GB was lower in 2010/11 and 2011/12 and plots were sampled on five occasions in 2010/11 and on thirteen occasions in 2011/12.

On each sampling occasion, the chamber was placed on the collar. Samples were taken using a gas-tight syringe and the three-way valve fitted to the top of the chamber. Gas samples were taken immediately after chamber closure (T_0) and again

after 15 (T_{15}) and 30 (T_{30}) minutes. All gas samples were transferred to 7 ml screw-cap exetainer with Teflon-coated butyl septa (Perbio Science, Cambridge, UK). Air temperature inside the chamber and soil temperature at 5 cm depth beside the chamber was recorded using a thermo-sensor (Sensor-Tech Ltd., Castlebellingham, Ireland).

Gas samples were analysed using a gas chromatograph (GC) (Varian GC 450; Netherlands) fitted with an electron capture detector (ECD) and automatic sampler (Combi-PAL auto sampler; CTC Analytics, Zwingen, Switzerland). Column and detector temperatures were 60 and 300 °C, respectively. The N₂O concentration (parts per million, ppm) of each sample was calculated using calibration curves. These curves were determined using N₂O standards gases with concentrations of 0.2, 0.5, 1, 5, 10, 20, 50 and 100 ppm.

Herbage yield and N uptake in herbage

Herbage yield was measured in each WC paddock before each grazing (or silage harvest) and concurrently on each GB plot. A 10 × 1.2 m strip was harvested to 4 cm above ground level using a rotary mower (Etesia UK Ltd, Warick, UK). The fresh herbage was weighed and a sub-sample of 100 g of the fresh herbage was dried for 24 h in an oven with forced-air circulation at 100 °C to determine DM content. A second sub-sample was stored at –20 °C before being freeze dried and milled through a 0.2 mm sieve prior to N content analysis. The N concentration in the herbage DM was determined using a Leco N analyser (LECO FP-2000; Leco Coroportiaon, St. Joseph, MI, USA). Uptake of N in herbage DM (kg N ha^{–1}) was calculated using the N concentration in the herbage DM (g kg^{–1}) and the DM herbage yield (kg DM ha^{–1}). Nitrogen uptake in herbage DM for the GB plots was predominantly from the mineralization of soil organic matter N (MSON) and was therefore used as an indication of the difference in MSON between years but not as a direct measure of MSON. This assumes that there was little difference in the input of N to GB through dry deposition of NH₃ between years.

Calculations

Pre-experimental testing showed a linear increase in N₂O concentration in the chambers over a 60 minute test period. Therefore, nitrous oxide flux calculations (g N ha^{–1} day^{–1}) were calculated using the linear change in N₂O concentrations in the chamber between T_0 , T_{15} and T_{30} . Daily N₂O fluxes from each WC paddock were calculated using the arithmetic means of five chambers. Cumulative N₂O emissions were calculated by linear interpolation between sampling occasions. Global N₂O emission factors (ϵ_{EF}) of all applied N were calculated for WC using:

$$\epsilon_{\text{EF}}(\%) = \frac{(\text{Annual N}_2\text{O} - \text{N})_{\text{WC}} \text{ minus } (\text{Annual N}_2\text{O} - \text{N})_{\text{GB}}}{\text{Total N applied}}$$

where total N applied is the sum of fertilizer N, N in cattle slurry and N deposited during grazing on WC. Total N

applied was adjusted to account for losses of NH_3 and NO_x assumed to be 10% from fertilizer N and 20% from N in cattle slurry and N deposited during grazing (IPCC, 2006). Biologically fixed N (BFN) was not included in the G_{EF} calculations in this study as IPCC guidelines do not include BFN as a source of N_2O (IPCC, 2006).

Annual soil N balances (SNB) for WC were calculated to determine available soil N for N_2O emissions in each year using:

$$\text{SNB} = \text{N input}_{\text{WC}} \text{ minus } (\text{N uptake in herbage}_{\text{WC}} \text{ minus } \text{N uptake in herbage}_{\text{GB}})$$

where N input is the sum of fertilizer N, slurry N, N deposited during grazing, BFN and rainfall N deposition. Fertilizer N, slurry N and N deposited during grazing were adjusted for NH_3 volatilization as above. Rainfall N deposition was calculated by multiplying annual rainfall by rainfall total N concentration as described by (Necpálová *et al.*, 2013). The N outputs from the SNB were the N removed in herbage on WC minus the background N supplied for herbage production i.e. N uptake on GB. The N removed in herbage on WC in the SNB was corrected for N uptake on GB as background supply of N is an important contributor to herbage N uptake at this site (O'Connell, 2005).

Statistical analysis

All data were statically analysed using SAS 9.3 (SAS 2011, Institute Inc., Cary, NC, USA). Data were checked for normality by using residual normality and variance analysis. All N_2O data required a natural log transformation [$y = \log(x + 1)$]. The value of one was added before log transforming the N_2O data, which was sufficient to prevent the generation of negative transformed values. Substantially larger values of the constant added to the data may affect inference. Annual cumulative N_2O emissions and N uptake in herbage were analysed by ANOVA. The variables included in the model were treatment, year and the treatment by year interaction. Year was included as a repeated measure in the ANOVA. Post hoc

Holm–Tukey and Turkey–Kramer multiple comparisons test were carried out to determine differences between treatments within and between years.

Pearson correlation analysis was used to test for relationships between transformed annual N_2O emissions and N input, weather and soil variables using data from each plot in each year ($n = 12$ for each treatment). Single and multiple linear regression (stepwise) analysis were carried out to create simple explanatory models using the same variables to account of variation in annual N_2O emissions. A statistical probability of $P < 0.05$ was considered significant for all statistical tests.

Results

Soil and weather conditions

Annual rainfall was 1183, 1037, 919 and 1394 mm in 2008/09, 2009/10, 2010/11 and 2011/12, respectively. Monthly rainfall was higher than evapo-transpiration for much of the study particularly in 2008/09 and 2011/12 (Fig. 1). Soil WFPS ranged between 30% and 100% and remained above 70% for the majority of the study. The only major exception to this was the second half of 2009/10 and the grazing season of 2010/11 (Fig. 2a). Minimum and maximum mean monthly soil temperatures at 5 cm depth ranged from 3.9 to 18.3 °C, 1.9 to 18.4 °C, 1.5 to 17.1 °C and 5.4 to 17.7 °C for each of the 4 years. There was an exceptional period of cold temperatures in December of 2010/11 (Fig. 2b). The mean monthly air temperature in December 2010 (0.6 °C) was much lower than typical December temperatures at this site (1950–2012 mean excluding 2010 $\pm$ SD; 6.0 ± 1.2 °C). Furthermore, the mean monthly air temperature in December 2010 was the lowest recorded between 1950 and 2012.

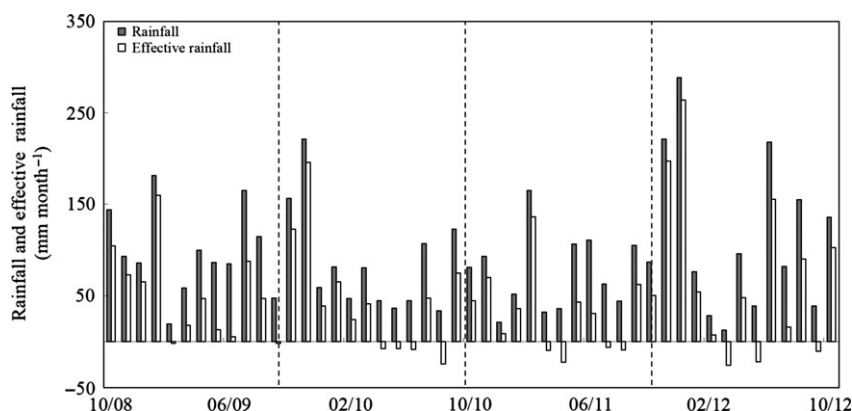


Fig. 1 Monthly rainfall and effective rainfall (rainfall minus evapotranspiration) (mm month^{-1}) at Solohead Research farm during the study period (mm/yr). Dashed vertical lines separate measurement years.

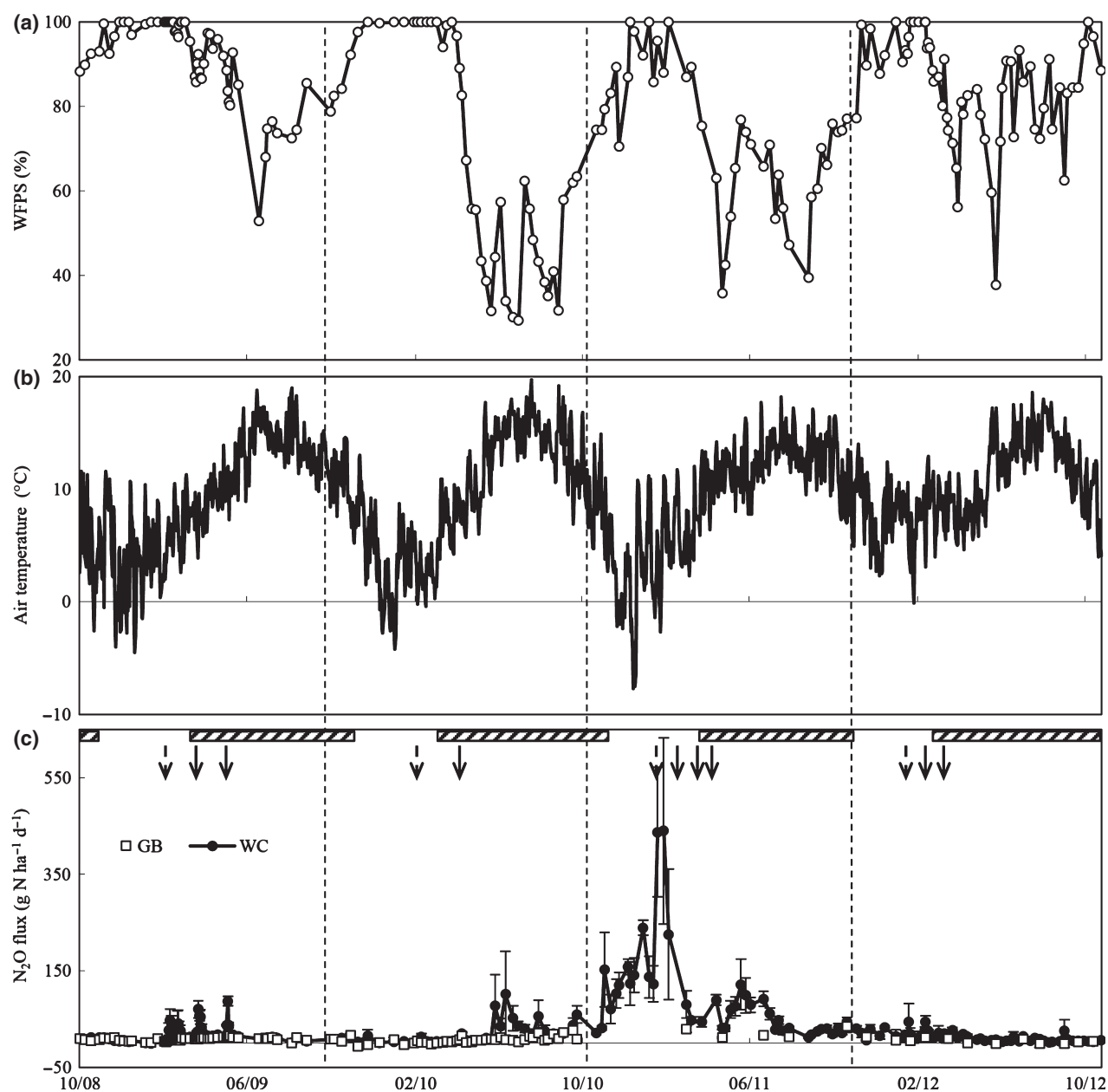


Fig. 2 Water filled pore space (%) at 0–5 cm depth (a), daily air temperature (°C) (b) and daily N₂O fluxes (mean $\pm$ SE) from unfertilized perennial ryegrass plots (GB) and grazed and fertilized perennial ryegrass/white clover pastures (WC) (c) during the study (mm/yy). Dashed vertical lines separate measurement years. Dashed arrows indicate dates of cattle slurry application; full arrows indicate dates of fertilizer N application. The vertical edges of the shaded boxes represent the beginning and end of each grazing season.

Temporal trends in daily N₂O fluxes

Nitrous oxide fluxes from WC were highly episodic with daily mean values displaying large variation throughout the study and there were clear differences in the rates of daily N₂O fluxes both intra- and interannually (Fig. 2c). Mean daily N₂O-N emissions (g ha⁻¹) from WC ranged from -0.1 to 87.3 in 2008/09, 0.8 to

101.7 in 2009/10, 12.3 to 439.8 in 2010/11 and 1.7 to 44.0 in 2011/12. Highest daily N₂O fluxes from WC were generally found following N applications and during the main grazing season. Fluxes during winter were relatively low except for the winter of 2010/11, where peak fluxes reached 239.3 g N₂O-N ha⁻¹ d⁻¹ on the 4 January 2011. Cumulative N₂O-N emissions from WC were 9.2 kg ha⁻¹ between 20 November 2010 and 20 January

2011 (period between the end of grazing season 2010 to first application of cattle slurry in 2011: Fig. 2c) and equated to an 8 to 39-fold increase relative to the cumulative emissions during the same period of each of the other 3 years of the study.

The range in background emissions of N_2O (from GB plots) was lower than WC, particularly following N applications to WC and during the grazing seasons (Fig. 2c). Mean daily N_2O -N emissions ($g\ ha^{-1}$) from GB ranged from 0.5 to 14.7 in 2008/09, -2.1 to 31.5 in 2009/10, 2.0 to 35.1 in 2010/11 and -0.9 to 12.8 in 2011/12. The daily fluxes on GB were generally higher during the spring to autumn period compared with the winter.

Annual N_2O emissions

The annual N_2O emissions ($kg\ N_2O$ -N ha^{-1} ; mean $\pm$ SE) from WC ranged from 4.4 ± 0.2 in 2008/09 to 34.4 ± 5.5 in 2010/11 and from GB ranged from 1.7 ± 0.8 in 2011/12 to 6.3 ± 1.2 in 2010/11 (Fig. 3). Emissions of N_2O were affected by treatment ($P < 0.01$), year ($P < 0.001$) and an interaction ($P < 0.05$) between treatment and year. There was no difference in annual N_2O emissions between WC and GB in 2008/09, while N_2O emissions were higher ($P < 0.01$) from WC in each of the other years. The exceptionally high annual N_2O emissions from WC in 2010/11 were higher ($P < 0.001$) than emissions from WC in any of the other years, which were not different from each other.

On GB, N_2O emissions were higher ($P < 0.05$) in 2010/11 than in 2011/12. Otherwise, there were no differences in annual emissions from GB between years.

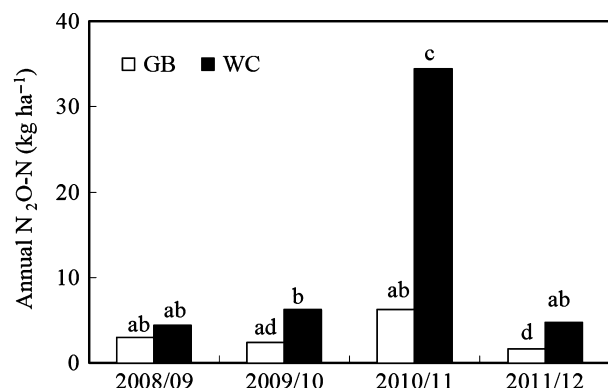


Fig. 3 Annual N_2O emissions from unfertilized perennial ryegrass plots (GB) and grazed and fertilized perennial ryegrass/white clover pastures (WC) in 2008/09, 2009/10, 2010/11 and 2011/12. Statistically significant differences ($P < 0.05$) between year $\times$ treatments are indicated by different letters. Emissions on GB in 2010/11 may be underestimated as no measurements were taken during the winter of 2010, which had elevated emissions on WC.

The annual G_{EF} (mean $\pm$ SE) calculated for WC were 0.5 ± 0.2 in 2008/09, 1.6 ± 0.5 in 2009/10, 7.7 ± 1.6 in 2010/11 and 1.2 ± 0.3 in 2011/12.

Uptake of N in herbage

Uptake of N in herbage was affected by treatment ($P < 0.05$), year ($P < 0.001$) and an interaction ($P < 0.001$) between treatment and year (Fig. 4). Uptake of N was higher ($P < 0.01$) on WC than GB in 2009/10, 2010/11 and 2011/12 but not different in 2008/09.

On GB, uptake of N in 2010/11 was higher ($P < 0.05$) than in 2009/10 and 2011/12, while there was no difference in uptake between the other years (Fig. 4).

Factors affecting annual N_2O emissions

There were significant correlations ($P < 0.05$) between weather, soil variables and N input with annual N_2O emissions (Table 2). There was no significant correlation between annual soil N balances and N_2O emissions from WC.

The best regression model for annual N_2O emissions from WC included mean soil temperature in December and annual rainfall (Table 3). The inclusion of annual soil temperature instead of annual rainfall also produced a robust model. Mean soil temperature in December accounted for the largest proportion of variation (highest semi-partial R^2) in both of these models and hence variation in annual N_2O emissions from WC.

Single regression models including either effective rainfall or soil temperature in December were the best model to account for variation in annual N_2O emissions from GB (Table 3).

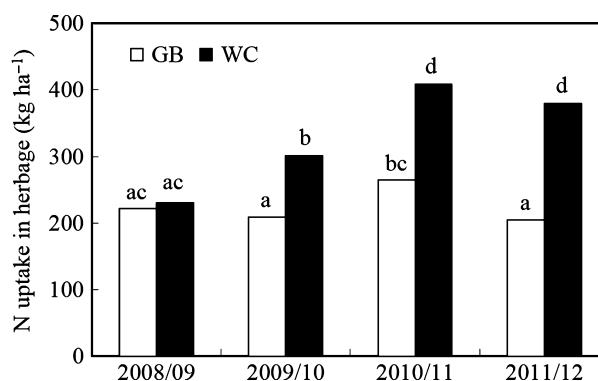


Fig. 4 Annual N uptake in herbage dry matter on unfertilized perennial ryegrass plots (GB) and grazed and fertilized perennial ryegrass/white clover pastures (WC) in 2008/09, 2009/10, 2010/11 and 2011/12. Statistically significant differences ($P < 0.05$) between year $\times$ treatments are indicated by different letters.

Table 2 Pearson correlation coefficients (*r*) between annual N₂O emissions (*n* = 12) on unfertilized perennial ryegrass plots (GB) and grazed and fertilized perennial ryegrass/white clover pastures (WC) with annual and monthly explanatory variables

	N input§	Soil N balance††	N uptake WC	N uptake GB	Rainfall	Effective rainfall	Mean WFPS‡‡	Temp December§§	Annual Temp†††	N ₂ O WC
N ₂ O WC‡	0.91***	0.29NS	0.65*	0.52†	−0.74**	−0.75**	−0.71**	−0.91***	−0.65*	1
N ₂ O GB‡	–	–	–	0.51†	−0.69*	−0.74**	−0.46NS	−0.74**	−0.71**	0.68*

‡N₂O data were transformed using the natural log transformation: $\ln(N_2O+1)$.

§N input = Fertilizer N + slurry N + excreta N + biologically fixed N + rainfall N deposition.

††Soil N balance = N input minus (N uptake WC minus N uptake GB).

‡‡WFPS = Mean annual soil water filled pore space (0–5 cm).

§§Temp December = Mean monthly soil temperature in December (0–5 cm).

†††Annual temp = Mean annual soil temperature (0–5 cm).

†*P* < 0.1, **P* < 0.05, ***P* < 0.01, ****P* < 0.001, NS = not significant.

Table 3 Multiple and single linear regressions models accounting for variation in annual N₂O emissions from unfertilized perennial ryegrass plots (GB) and grazed and fertilized perennial ryegrass/white clover pastures (WC) using annual and monthly explanatory variables

Treatment*	Variable†	Estimate	SE	Semi-partial <i>R</i> ²	<i>P</i> value	Model <i>R</i> ²
WC	Temp Dec	−1.011	0.115	0.83	<0.001	0.95
	Rainfall	0.005	0.001	0.12	<0.001	
WC	Temp Dec	−0.831	0.099	0.83	<0.001	0.94
	Annual temp	1.875	0.487	0.11	<0.01	
GB	E. rain	−0.002	0.001	–	<0.01	0.55
	Temp Dec	−0.255	0.073	–	<0.01	

*N₂O data were transformed using the natural log transformation: $\ln(N_2O+1)$.

†Temp Dec = Mean monthly soil temperature in December (0–10 cm), Rainfall = annual rainfall, Annual temp = Mean annual soil temperature (0–5 cm), N Input = Fertilizer N + slurry N + excreta N + biologically fixed N + rainfall N deposition, E. rain = annual effective rainfall, SE = standard error.

Discussion

Annual N₂O emissions

The mean annual N₂O-N emission of 12.5 kg ha^{−1} recorded between 2008 and 2012 for the WC pastures was very high compared to the annual average of 1.77 kg ha^{−1} from multiple managed grassland sites across Europe (Flechard *et al.*, 2007). The high level of N₂O emissions in this study were, however, similar to other Irish studies on fertilized grazed perennial ryegrass pastures receiving annual N input of between 53 and 390 kg ha^{−1} (Hyde *et al.*, 2006; Rafique *et al.*, 2011). The emissions in this study were also within the range of emissions (0.85 to 51.3 kg ha^{−1}) found in fertilized grassland in the UK (Cardenas *et al.*, 2010; Rees *et al.*, 2013).

The annual background soil N₂O-N emissions from GB were also high compared with the range of −0.5 to 1.2 kg ha^{−1} at multiple grassland sites across Europe (Flechard *et al.*, 2007) and with emissions (circa. 1 kg ha^{−1}) from sandy loam-based pastures in Ireland

(Abdalla *et al.*, 2009). However, emissions within the range of those reported in this study (with the exception of the high rates recorded for 2010/11) have been previously reported for gleysol-based grasslands in Ireland (−1.6 to 4.66 kg ha^{−1}) (Hyde *et al.*, 2006; Rafique *et al.*, 2011).

Global N₂O emission factors

The *G*_{EF} for all applied N in this study are somewhat different to IPCC EF which calculate EF individually for fertilizer N, manure N and excreta N. Similar calculations have been carried out on other N₂O studies on grasslands, with *G*_{EF} ranging from 0.7% to 5.1% (Hyde *et al.*, 2006; Rafique *et al.*, 2011). The reliability of the *G*_{EF} estimates in 2010/11 and 2011/12 is questionable due to the lack of N₂O sampling on GB, particularly over the winter of 2010/11. More intensive gas sampling on GB would have improved the accuracy of these annual *G*_{EF} estimates and emphasizes the need for regular measurement of background emissions for accurate emission factor (EF) calculation. Nevertheless,

the robust $\delta^{15}\text{N}$ estimates produced in 2008/09 and 2009/10 (0.5–1.6%) were relatively low compared with the other studies outlined above.

Interannual variation in N_2O emissions

Few published studies have reported interannual variation in N_2O emissions from managed temperate grasslands. There was up to 7.8-fold difference in annual N_2O emissions over the 4 years in this study. This is significantly higher than other studies on managed grassland sites in Ireland, Wales and the Netherlands where 1.1 to 3-fold differences have been reported over 2 years (Velthof *et al.*, 1996; Hyde *et al.*, 2006; Cardenas *et al.*, 2010; Rafique *et al.*, 2011). Furthermore, background emissions from the GB plots in this study also displayed up to 3.7-fold differences between years whilst previous studies reported 1.1 to 1.9-fold differences between years (Hyde *et al.*, 2006; Rafique *et al.*, 2011).

Highest emissions from both the WC and GB treatments were measured in 2010/11, which contributed to the large variation between years in this study. During 2010/11, there were exceptionally low temperatures in winter, lower annual rainfall and the highest N inputs.

Influence of N input on N_2O emissions

Input of N had a large impact on annual N_2O emissions in this study, with significantly higher emissions of N_2O from WC compared to GB in 3 years of the study (Fig. 3). Input of N also explains some of the increase in N_2O emissions in 2010/11 relative to the other years. However, the higher N input to WC in 2010/11 (29 to 39% higher relative to other years) was relatively small compared with the large increase in annual N_2O -N emissions (444 to 682% relative to other years; Fig 5a).

Input of N, background supply of N from MSON and the rate of N removal in herbage are all crucial factors driving the availability of soil N for N_2O emissions. Annual soil N balance for WC in 2010/11 was similar to 2008/09 and 2009/10 and indicates that despite the higher N additions in this year, the increase in N uptake in herbage on WC removed much of this additional N from the soil (Fig. 5b).

All the points above indicate that alternations in N management had a significant but small contribution to the higher N_2O emissions during 2010/11 relative to the effects of weather and soil conditions in this study.

Influence of low soil temperatures in December

The low temperatures recorded in December 2010 were atypical of the generally mild winter conditions

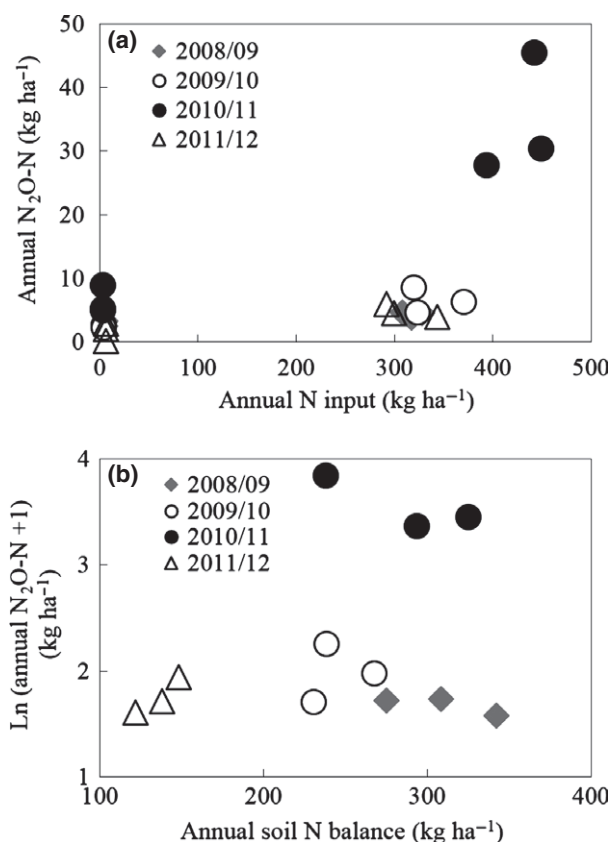


Fig. 5 Relationship between annual N_2O emissions and N input (a) and natural log transformed annual N_2O emissions and soil N balances (Pearson correlation $r = 0.29$, $P > 0.05$) (b) using data from unfertilized perennial ryegrass plots (GB) and grazed and fertilized perennial ryegrass/white clover pastures (WC). Annual N input is the sum of fertilizer N, slurry N, N deposited during grazing, biological N fixation and rainfall N deposition. Annual N input was adjusted to account for losses of NH_3 and NO_x assumed to be 10% from fertilizer N and 20% from N in cattle slurry and N deposited during grazing (IPCC, 2006).

recorded at this site. Daily mean air temperatures were also low during periods of the winter of 2008/09 and 2009/10 (Fig. 2b) but maximum daily air temperatures generally remained above zero. In contrast, maximum daily air temperature dropped below zero (minimum of -4.7°C) for an extended period of 6 days between 20 and 25 December 2010.

Emissions of N_2O -N during winter 2010 were up to 39 times higher than in other years of this study. The relatively cold temperatures in December 2010 are likely to have led to soil freezing and thawing, which have previously been found to stimulate large N_2O emissions from both grassland and arable soils (Kaiser *et al.*, 1998; Kammann *et al.*, 1998; Lampe *et al.*, 2006). Increased N_2O emissions can be attributed to the release of easily degradable N and C through the lysis of micro-organisms and disrupted soil aggregates and

the reduction in N₂O reductase activity (Christensen & Tiedje, 1990; Christensen & Christensen, 1991; van Bochove *et al.*, 2000; Holtan-Hartwig *et al.*, 2002; Müller *et al.*, 2002; Mørkved *et al.*, 2006). Increased emissions during thawing can be explained by the physical release of accumulated N₂O which was produced by active micro-organisms in unfrozen soil water (Teepe *et al.*, 2001) or by a reduction in the diffusion resistance to gases that occurs during thawing (de Bruijn *et al.*, 2009).

Influence of rainfall on N₂O emissions

In this study, N₂O emissions were lower under high rainfall and high WFPS (Table 2). These relationships are quite different to most other studies which generally find N₂O emissions to increase following rainfall events and with increasing WFPS (Dobbie & Smith, 2003; Liu *et al.*, 2007). Taking into account, the heavy textured soils and impeded drainage at this site, periods of high rainfall led to a shift from wet to completely saturated soil conditions in this study, whereas in many other studies it is likely that periods of high rainfall lead to a shift from dry to wet soil conditions. Saturated soil conditions (anaerobic) create more favourable conditions for complete denitrification of nitrate to dinitrogen gas, rather than nitrification and partial denitrification which would be favoured under lower WFPS (i.e. anoxic soil conditions) which were experienced in 2010/11 in this study (Monaghan & Barraclough, 1993; Rudaz *et al.*, 1999).

The relatively low rainfall in 2010/11 allowed the soils at this site to dry out more than normal, which stimulated a transition from completely saturated anaerobic to wet partially aerobic (anoxic) soil conditions (Fig. 2a). Drying soil conditions is likely to have favoured net mineralization of soil organic N (Danevčič *et al.*, 2010; Goldberg *et al.*, 2010), which is reflected in the increase in N uptake in herbage DM on GB in 2010/11 (Fig. 3), and which provided additional substrate for nitrifying and denitrifying bacteria.

Nitrification has been found to be a major contributor to N₂O under WFPS ranging from 40 to 60% (Stevens *et al.*, 1997; Bateman & Baggs, 2005). The reduction of WFPS to between 30% and 80% during these periods in the present study is likely to have allowed both nitrification and partial denitrification to contribute to N₂O production simultaneously or lead to coupled nitrification–denitrification (Wrage *et al.*, 2001). This may explain some of the increase in N₂O emissions in 2010/11 relative to other years. The results confirm that N₂O emissions are very much increased under these anoxic soil conditions. Management decisions which support, or lead to, such soil conditions should be avoided if

N₂O emissions from grasslands, such as those in this study, are to be mitigated.

Implications of the study

The results indicate that annual variation in N₂O emissions from WC were regulated by weather and soil variables and N input. The dominance of weather-driven variables in regulating interannual variation has implications in the context of future climate scenarios where increased weather volatility, including temperature extremes and patterns of rainfall are predicted (Solomon *et al.*, 2007; Miraglia *et al.*, 2009).

Many studies have stressed the errors and uncertainty associated with N₂O chamber measurements (Smith & Dobbie, 2001; Rochette & Eriksen-Hamel, 2007; Kroon *et al.*, 2008). The results of this study suggest that short-term studies may lead to bias and the underestimation or likewise the overestimation of annual N₂O emissions. At least 2–3 years of measurement may be required to gain a representative estimate of annual N₂O emissions from an individual site or production system which contrasts with the recommendations of Velthof *et al.* (1996). The significant contribution of the winter of 2010/11 to annual N₂O emissions also highlights the need for frequent N₂O measurements over winter periods even in temperate-grazed grasslands as disregarding this period could lead to underestimations of annual N₂O emission estimates. The large variation in emissions that occurred in this study also highlights the need for regular measurement of background emissions to provide robust data for EF calculation.

Acknowledgements

This work was funded by the EU Framework Programme 7 project Legume Futures. The author thanks Kevin McNamara, internship students and the staff of Solohead Research Farm for assistance with field sampling and Dr Jim Grant for assistance with statistical analysis of the data.

References

- Abdalla M, Jones M, Smith P, Williams M (2009) Nitrous oxide fluxes and denitrification sensitivity to temperature in Irish pasture soils. *Soil Use and Management*, **25**, 376–388.
- Bateman EJ, Baggs EM (2005) Contributions of nitrification and denitrification to N₂O emissions from soils at different water-filled pore space. *Biology and Fertility of Soils*, **41**, 379–388.
- van Bochove E, Prévost D, Pelletier F (2000) Effects of freeze-thaw and soil structure on nitrous oxide produced in a clay soil. *Soil Science Society of America Journal*, **64**, 1638–1643.
- de Bruijn AMG, Butterbach-Bahl K, Blagodatsky S, Grote R (2009) Model evaluation of different mechanisms driving freeze-thaw N₂O emissions. *Agriculture Ecosystems & Environment*, **133**, 196–207.

- Cai YJ, Ding WX, Luo JF (2013) Nitrous oxide emissions from Chinese maize-wheat rotation systems: a 3-year field measurement. *Atmospheric Environment*, **65**, 112–122.
- Cantarel AAM, Bloor JMG, Pommier T, Guillaumaud N, Moirot C, Soussana J-F, Poly F (2012) Four years of experimental climate change modifies the microbial drivers of N₂O fluxes in an upland grassland ecosystem. *Global Change Biology*, **18**, 2520–2531.
- Cardenas LM, Thorman R, Ashlee N *et al.* (2010) Quantifying annual N₂O emission fluxes from grazed grassland under a range of inorganic fertiliser nitrogen inputs. *Agriculture, Ecosystems & Environment*, **136**, 218–226.
- Christensen S, Christensen BT (1991) Organic matter available for denitrification in different soil fractions: effect of freeze/thaw cycles and straw disposal. *Journal of Soil Science*, **42**, 637–647.
- Christensen S, Tiedje JM (1990) Brief and vigorous N₂O production by soil at spring thaw. *Journal of Soil Science*, **41**, 1–4.
- Cuhel J, Simek M, Laughlin RJ, Bru D, Cheneby D, Watson CJ, Philippot L (2010) Insights into the effect of soil pH on N₂O and N₂ emissions and denitrifier community size and activity. *Applied and Environmental Microbiology*, **76**, 1870–1878.
- Danevčič T, Mandić-Mulec I, Stres B, Stopar D, Hacin J (2010) Emissions of CO₂, CH₄ and N₂O from Southern European peatlands. *Soil Biology and Biochemistry*, **42**, 1437–1446.
- Dobbie KE, Smith KA (2003) Nitrous oxide emission factors for agricultural soils in Great Britain: the impact of soil water-filled pore space and other controlling variables. *Global Change Biology*, **9**, 204–218.
- Du R, Lu D, Wang G (2006) Diurnal, seasonal, and inter-annual variations of N₂O fluxes from native semi-arid grassland soils of inner Mongolia. *Soil Biology and Biochemistry*, **38**, 3474–3482.
- Firestone MK, Davidson EA (1989) Microbiological basis of NO and N₂O production and consumption in soil. In: *Exchange of Trace Gases Between Terrestrial Ecosystems and The Atmosphere* (eds Andreae MO, Schimel DS), pp. 7–21. John Wiley & Sons, Chichester, UK.
- Fitzgerald D, Fitzgerald D (2004) Meteorological measurements. In: *Climate, Weather and Irish Agriculture* (eds Keane T, Collins JF), pp. 9–26. Met Éireann, Dublin, Ireland.
- Flechar CR, Ambus P, Skiba U *et al.* (2007) Effects of climate and management intensity on nitrous oxide emissions in grassland systems across Europe. *Agriculture, Ecosystems & Environment*, **121**, 135–152.
- Goldberg SD, Knorr K-H, Blodau C, Lischied G, Gebauer G (2010) Impact of altering the water table height of an acidic fen on N₂O and NO fluxes and soil concentrations. *Global Change Biology*, **16**, 220–233.
- Holtan-Hartwig L, Dörsch P, Bakken LR (2002) Low temperature control of soil denitrifying communities: kinetics of N₂O production and reduction. *Soil Biology and Biochemistry*, **34**, 1797–1806.
- Humphreys J, O'Connell K, Casey IA (2008) Nitrogen flows and balances in four grassland-based systems of dairy production on a clay-loam soil in a moist temperate climate. *Grass and Forage Science*, **63**, 467–480.
- Hutchinson GL, Moirer AR (1981) Improved soil cover method for field measurement of nitrous oxide fluxes. *Soil Science Society of America Journal*, **45**, 311–316.
- Hyde BP, Hawkins MJ, Fanning AF, Noonan D, Ryan M, O' Toole P, Carton OT (2006) Nitrous oxide emissions from a fertilized and grazed grassland in the South East of Ireland. *Nutrient Cycling in Agroecosystems*, **75**, 187–200.
- IPCC (2006) 2006 IPCC guidelines for national greenhouse gas inventories. In: *Agriculture, Forestry and Other Land Use* (eds De Klein C, Novoa RSA, Ogle S, Smith KA, Rochette P, Wirth TC, McConkey BG, Mosier A, Rypdal K), pp. 5–53. IGES, Japan.
- Kaiser EA, Kohrs K, Kücke M, Schnug E, Heinemeyer O, Munch JC (1998) Nitrous oxide release from arable soil: importance of N-fertilization, crops and temporal variation. *Soil Biology and Biochemistry*, **30**, 1553–1563.
- Kammann C, Grünhage L, Miillerb C, Grünhage Ludger, Christoph M, Jacobi S, Hans-Jürgen J (1998) Seasonal variability and mitigation options for N₂O emissions from differently managed grasslands. *Environmental Pollution*, **102**, 179–186.
- de Klein CAM, Pinares-Patino C, Waghorn GC (2008) Greenhouse gas emissions. In: *Environmental Impacts of Pasture-based Farming* (ed. McDowell RW), pp. 1–32. CAB International, Oxfordshire, UK.
- Kroon P, Hensen A, van den Bulk W, Jongejan P, Vermeulen A (2008) The importance of reducing the systematic error due to non-linearity in N₂O flux measurements by static chambers. *Nutrient Cycling in Agroecosystems*, **82**, 175–186.
- Lampe C, Dittert K, Sattelmacher B, Wachendorf M, Loges R, Taube F (2006) Sources and rates of nitrous oxide emissions from grazed grassland after application of ¹⁵N-labelled mineral fertilizer and slurry. *Soil Biology and Biochemistry*, **38**, 2602–2613.
- Liu XJ, Mosier AR, Halvorson AD, Reule CA, Zhang FS (2007) Dinitrogen and N₂O emissions in arable soils: effect of tillage, N source and soil moisture. *Soil Biology and Biochemistry*, **39**, 2362–2370.
- Luo J, Tillman RW, Ball PR (1999) Factors regulating denitrification in a soil under pasture. *Soil Biology and Biochemistry*, **31**, 913–927.
- Miraglia M, Marvin HJP, Kleter GA *et al.* (2009) Climate change and food safety: an emerging issue with special focus on Europe. *Food and Chemical Toxicology*, **47**, 1009–1021.
- Monaghan RM, Barraclough D (1993) Nitrous-oxide and dinitrogen emissions from urine-affected soil under controlled conditions. *Plant and Soil*, **151**, 127–138.
- Mørkved PT, Dörsch P, Henriksen TM, Bakken LR (2006) N₂O emissions and product ratios of nitrification and denitrification as affected by freezing and thawing. *Soil Biology and Biochemistry*, **38**, 3411–3420.
- Müller C, Martin M, Stevens RJ, Laughlin RJ, Kammann C, Ottow JCG, Jäger HJ (2002) Processes leading to N₂O emissions in grassland soil during freezing and thawing. *Soil Biology and Biochemistry*, **34**, 1325–1331.
- Necpálová M, Phelan P, Casey IA, Humphreys J (2013) Soil surface N balances and soil N content in a clay-loam soil under Irish dairy production systems. *Nutrient Cycling in Agroecosystems*, **96**, 47–65.
- O' Connell K (2005) Environmentally sustainable fertiliser nitrogen management practices for pasture production. PhD. Thesis, Queen's University of Belfast, Belfast.
- Rafique R, Hennessy D, Kiely G (2011) Nitrous oxide emission from grazed grassland under different management systems. *Ecosystems*, **14**, 563–582.
- Rees RM, Augustin J, Alberti G *et al.* (2013) Nitrous oxide emissions from European agriculture - an analysis of variability and drivers of emissions from field experiments. *Biogeosciences*, **10**, 2671–2682.
- Ri X, Wang Y, Zhang X, Ji B, Wang M (2003) A comparison between measured and modeled N₂O emissions from Inner Mongolian semi-arid grassland. *Plant and Soil*, **255**, 513–528.
- Rochette P, Eriksen-Hamel NS (2007) Chamber measurements of soil nitrous oxide flux: are absolute values reliable? *Soil Science Society of America Journal*, **72**, 331–342.
- Rudaz AO, Wälti E, Kyburz G, Lehmann P, Fuhrer J (1999) Temporal variation in N₂O and N₂ fluxes from a permanent pasture in Switzerland in relation to management, soil water content and soil temperature. *Agriculture, Ecosystems & Environment*, **73**, 83–91.
- Schulte RPO, Diamond J, Finkle K, Holden NM, Brereton AJ (2005) Predicting the soil moisture conditions of Irish grasslands. *Irish Journal of Agricultural and Food Research*, **44**, 95–110.
- Smith KA, Dobbie KE (2001) The impact of sampling frequency and sampling times on chamber-based measurements of N₂O emissions from fertilized soils. *Global Change Biology*, **7**, 933–945.
- Solomon S, Qin D, Manning M *et al.* (2007) Technical summary. In: *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* (eds Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL), pp. 66–79. Cambridge University Press, Cambridge.
- Stevens RJ, Laughlin RJ (1998) Measurement of nitrous oxide and di-nitrogen from agricultural soils. *Nutrient Cycling in Agroecosystems*, **52**, 131–139.
- Stevens RJ, Laughlin RJ, Burns LC, Arah JRM, Hood RC (1997) Measuring the contributions of nitrification and denitrification to the flux of nitrous oxide from soil. *Soil Biology and Biochemistry*, **29**, 139–151.
- Teepe R, Brumme R, Beese F (2001) Nitrous oxide emissions from soil during freezing and thawing periods. *Soil Biology and Biochemistry*, **33**, 1269–1275.
- Velthof GL, Brader AB, Oenema O (1996) Seasonal variations in nitrous oxide losses from managed grasslands in The Netherlands. *Plant and Soil*, **181**, 263–274.
- Webster CP, Dowdell RJ (1982) Nitrous oxide emission from permanent grass swards. *Journal of the Science of Food and Agriculture*, **33**, 227–230.
- Wrage N, Velthof GL, van Beusichem ML, Oenema O (2001) Role of nitrifier denitrification in the production of nitrous oxide. *Soil Biology and Biochemistry*, **33**, 1723–1732.