Immunosuppressive networks and checkpoints controlling antitumor immunity and their blockade in the development of cancer immunotherapeutics and vaccines

A Q Butt and K H G Mills

Oncogene (2014) 33, 4623–4631; doi:10.1038/onc.2013.432; published online 21 October 2013

Vaccines that promote protective adaptive immune responses have been successfully developed against a range of infectious diseases, and these are normally administered prior to exposure with the relevant virus or bacteria. Adaptive immunity also plays a critical role in the control of tumors. Immunotherapeutics and vaccines that promote effector T cell responses have the potential to eliminate tumors when used in a therapeutic setting. However, the induction of protective antitumor immunity is compromised by innate immunosuppressive mechanisms and regulatory cells that often dominate the tumor microenvironment. Recent studies have shown that blocking these suppressor cells and immune checkpoints to allow induction of antitumor immunity is a successful immunotherapeutic modality for the treatment of cancer. Furthermore, stimulation of innate and consequently adaptive immune responses with concomitant inhibition of immune suppression, especially that mediated by regulatory T (Treg) cells, is emerging as a promising approach to enhance the efficacy of therapeutic vaccines against cancer. This review describes the immunosuppressive mechanisms controlling antitumor immunity and the novel strategies being employed to design effective immunotherapeutics against tumors based on inhibition of suppressor cells or blockade of immune checkpoints to allow induction of more potent effector T cell responses. This review also discusses the potential of using a combination of adjuvants with inhibition of immune checkpoint or suppressor cells for therapeutic vaccines and the translation of pre-clinical studies to the nextgeneration vaccines against cancer in humans. Keywords: immunotherapy; vaccine; antitumor immunity; immune checkpoint; regulatory T cell; Toll-like receptor agonist

INTRODUCTION

It is well established that the immune system can restrain tumor growth. Adaptive immune responses, in particular IFN-y-secreting T cells, play a central role in tumor immune surveillance.\(^1\) However, tumor growth is also associated with immune escape, immune subversion and immune suppression. Immune oditing gives rise to tumor variants that can escape immune surveillance.\(^2\) Immune subversion strategies include downregulation of MHC class I or co-stimulatory molecules, such as CD80/86 on antigen presenting cells (APC) or enhancement of co-inhibitory inhibitory molecules such as cytotoxic T-lymphocyte antigen-4 (CTLA-4) or programmed death-1 (PD-1) on T cells.\(^3\) Furthermore, the development of cancer is associated with the induction or recruitment of regulatory cells and the production of molecules that suppress antitumor effector T cell responses.\(^4\) These immune suppression networks include regulatory T (Treg) cells, myeloid derived suppressor cells (MDSC) and type 2 (M2) macrophages, as well as the immunosuppressive cytokines, IL-10 and TGF-\(\textit{B}\).

The potential of using of immunotherapy, where the patient's own immune system is enhanced to attack tumors, is gaining momentum as a viable approach for the treatment of cancer. Immunotherapeutic approaches include strategies that directly enhance anti-tumor immunity or that block immune checkpoints or suppressor networks, thus allowing the development of effector

immune responses that eliminate the tumor (Figure 1). Immuneactivating approaches include non-specific stimulation of innate or adaptive immune responses against the tumor or specific stimulation of tumor-specific immune responses using vaccines that include tumor antigens. The aim is to selectively promote effective cytotoxic T-lymphocyte (CTL) and Th1 responses against the tumor. Prophylactic vaccines against infectious disease are among the most effective and least expensive interventions in modern medicine and function by generating protective adaptive immune responses with the help of adjuvants that activate innate immune responses. However, the application of these approaches against cancer is still in its infancy and is compromised by the fact that they must break tolerance to self antigens, they must work therapeutically and they must overcome the immunosuppressive environment of the growing tumor.

Dendritic cells (DCs) are key antigen APC, which play a vital role in activating and directing adaptive immune responses and thus DCs are being exploited in the development of vaccines against a range of cancers.⁶ The first licensed DC-based cancer vaccine Provenge⁷ induces antitumor responses *in vivo* by the adoptive transfer of DCs pulsed *in vitro* with tumor antigens and stimulated to mature with cytokines and immunomodulatory molecules. Nevertheless, this DC-based tumor vaccine confers only modest survival advantage and has limited success in mediating tumor regression.^{8,9}

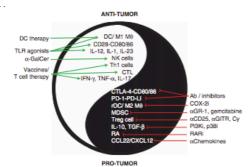


Figure 1. The pro- and anti-tumor arms of the immune response and the targets for the development of cancer immunotherapeutics and vaccines. The induction of antitumor immunity is dependent on activating DCs to express co-stimulatory molecule and innate inflammatory cytokine such as IL-12, IL-23 and IL-1 that promote Th1 and CTL responses. T cells secrete IFN-γ, TNF-α and IL-17, which have antitumor effects. However, these effector responses are suppressed by co-inhibitory molecules, such as PD-1/PD-I, CTL-4 and expansion of regulatory DCs (rDC), M2 macrophages and MDSC. These innate suppressor cells secrete IL-10 and TGF-β, which together with retinoic acid (RA), enhance induction of Treg cells that are recruited to the tumor under the influence of the chemokines, including CCL22 and CXCL12. Treg cells and innate regulatory cells suppress effector immune responses and thereby promote innate and adaptive immune responses, while cell-based therapies with DCs or T cells can also enhance antitumor immunity. Conversely, or in combination with these immune activating strategies, antibodies or inhibitors of the suppressive pathways and molecules can block immune checkpoints and suppressor cells and thereby enhance antitumor immunity. DC, dendritic cell; TLR, Coll-like receptor; αGalCer, alpha-galactosylceramide; MI Mθ, type I macrophage, NK, Natural killer; Th1, IFN-γ-secreting CD4 T cell; TLR, cytotoxic T lymphocyte, MDSC, myeloid-derived suppressor cell; Treg cell, regulatory T cell; RA, retinoic acid; Ab, antibody; i, inhibitor; Cy, cyclophosphoamide; Pi3Ki, Phosphatidylinositol 3-kinase inhibitor; p38i, p38 MAP kinase inhibitor; RARi, retinoic acid receptor-alpha inhibitors.

Toll-like receptor (TLR) agonists are potent activators of innate immune cells and also direct adaptive immunity and thus have been employed as adjuvants in a number of experimental cancer vaccines and have been tested in clinical trials. Although the TLR7 agonist, imiquimod is being used in humans for the treatment of superficial basal cell carcinoma, ¹⁰ clinical trials with other TLR agonists have not been so effective. One reason for the relatively modest success with TLR agonists-based immunotherapies and vaccines is that TLR agonists can promote regulatory as well as effector T cells and this is compounded in a therapeutic setting by the high prevalence of Treg cells infiltrating the tumor.

Depletion of Treg cells has been shown to promote antitumor responses in vivo in mouse tumor models. 11-13 Furthermore, Ipilimumab, a monoclonal neutralizing antibody that blocks CTLA-4 has shown good efficacy in melanoma patients, has been approved by the FDA. 12 Another therapeutic mononclonal antibody, Nivolumab (BMS-936558) that specifically blocks PD-1 has shown some efficacy against a range of human cancers. 14.15 However, persistent blockade of these inhibitory receptors has lead to the breakdown in immune self tolerance, thereby increasing susceptibility to autoimmune or auto-inflammatory side effects, including rash, colitis, hepatitis and endocrinopathies. 10 Alternatively, cancer vaccines that are rationally designed to specifically block tumor-associated immune checkpoints may

have reduced side effects. These could potentially involve combination therapy approaches, including inhibition of immunosuppressive cells, blockade of co-inhibitory molecules and the simultaneous activation of immune signaling pathways via TLR agonists to promote effector immune responses.

TARGETING TUMOR INFILTRATING IMMUNOSUPPRESSIVE CELLS

The induction of effective antitumor immune responses is hindered by the high number of infiltrating immunosuppressive cells in the tumor micro-environment, which in turn leads to poor effector immune responses. Therefore, targeting immunosuppressive cells, including Treg cells, MDSC and M2 macrophages has emerged as leading approach in the development of new immunotherapeutics and vaccines against cancer.

Regulatory T cells

Immunosuppressive cells, such as Treg cells, play a crucial role in maintaining the immune homeostasis, which depends on the balance between the immune responses that control infectious pathogens and tumors and the reciprocal immune responses that prevent inflammation and autoimmune diseases. If it is now well recognized that the natural Treg cells expressing FOXP3 and inducible antigen-specific Treg cells that secrete IL-10 and TGF-β play a protective role in immunity to infection by controlling pathogen-induced immunopathology and also prevent the development of autoimmune diseases. If However, in the tumor environment the frequency of Treg cells versus effector T cells is greater than in the general circulation, and their recruitment and activation is associated with tumor growth because of the local inhibition of the effector immune responses. Indeed, the effective immunosuppressive effects of Treg cells may in part explain the failure of many immunotherapeutic approaches to cancer. Inhibition of immunosuppressive cells using cyclophosphamide treatment has been shown to enhance antitumor immunity induced by vaccination in melanoma patients. Furthermore, systemic Treg cell depletion in melanoma patients induced regression of metastases. However, it is now accepted that more precise strategies are required to inhibit Treg cells to enhance effector cells that mediate antitumor immunity.

Treg cell depletion strategies have utilized monoclonal neutralizing antibodies or ligand-directed toxins targeted to cell surface receptors, such as CD25. Two anti-CD25 antibodies, daclizumab and basiliximab induce Treg cell death by cytokine deprivation (IL-2) and by triggering antibody-dependent cell-mediated cytotoxicity or complement-mediated cytotoxicity. The results of clinical trials have shown that daclizumab depletes Treg cells and thereby enhances CTL responses to tumor antigens induced by vaccination without any autoimmune side effects. ^{24,25}

In addition to Treg cell depletion strategies, other approaches have also been utilized to eliminate the immunosuppressive effects of Treg cells. For example, intratumoral injection with an agonistic antibody (DTA-1) to the glucocorticoid-induced tumor necrosis factor (TNF) receptor, which is constitutively expressed on Treg cells, invoked potent antitumor immunity and eradicated established tumors in mice. 26.27 Another strategy involves the inhibition of receptor activation of NF-κB (RANK) signaling using an anti-RANK ligand (RANKL) antibody denosumab, which blocks the engagement of RANKL on Treg cells to the RANK receptor on the tumor cells and thus blocks Treg cell-induced metastases of certain cancers. 4.28 Furthermore, targeting the Treg cell transcription factor FOXP3 by RNA interference, such as miR-31 (negative regulator of FOXP3), can modulate Treg cells functionality and abolishes their suppressor activity. 29

Furthermore, a peptide inhibitor of FOXP3 has been shown to impair Treg cell activity and enhance the efficacy of peptide vaccine against CT26 tumors in mice.³⁰ However, translating these approaches to clinical applications will be challenging, as they would have to be specifically delivered to tumor infiltrating Treg cells because FOXP3 is transiently expressed in activated effector T cells and in certain cancer cells.³¹

Despite the obvious benefits of increasing effector T cells in the tumor environment, depletion of Treg cells may have a high risk of developing systemic autoimmune diseases due to increased inflammatory responses. Thus, alternative approaches need to be devised that involve selective inhibition or depletion of immune suppressive cells in the tumor with optimal activation of antitumor T cell responses. One approach may be to target Treg cell migration into tumor. Studies in cancer patients have demonstrated significant trafficking of Tregs into tumors and have found an association between the frequency of intratumoral Treg cells and poor survival.³² Tumor cells and macrophages produce the chemokine C-C motif chemokine ligand 2 (CCL22), which promotes trafficking of Treg cells that express CCR4, the receptor for CCL22. Studies in mice showed that in vivo blockade of CCL22 reduced the trafficking of Treg cells into tumors.³³ CXCL12/CXCR4 signals have also been shown to regulate the Treg trafficking to the bone marrow in prostate cancer patients with bone metastasis.³⁴ Blocking the CXCL12/CXCR4 signals significantly reduces the Treg cell trafficking to bone marrow.^{32,34} Therefore, strategies that target chemokines to prevent T cell migration into tumor may be a more tailored and less risky approach than systemic depletion of Treg cells.

Myeloid-derived suppressor cells

MDSC are another class of immunosuppressive cells, which are a heterogeneous population of immature myeloid cells that accumulate under conditions of inflammation and in tumors and exert inhibitory function on immune responses. MDSCs are thought to promote the tumor growth by both enhancement of tumor angiogenesis and metastasis and also inhibition of antitumor effector T cell responses. Under normal physiological conditions, these cells are generated in bone marrow and differentiate into mature macrophages, DCs and granulocytes. However, in pathological conditions, there is a dramatic expansion of these Gr-1+CD11b+ cells, with the same phenotype and immunosuppressive activity in various tissues, and the differentia-tion into mature myeloid cells is blocked. 35,36 MDSCs regulate MDSCs regulate innate immune and T cell responses by depleting arginine, modulating cytokine production by macrophages, upregulating the production of immune-suppressive factors, such as nitric oxide and reactive oxygen species, $^{36-39}$ and by overexpressing anti inflammatory cytokines, such as TGF- β and IL-10. 40,41 MDSCs suppress proliferation and cytokine production by T cells and natural killer cells, as well as induce apoptosis of CD8⁺ T cells. ⁴² Interestingly, MDSCs have also been shown to indirectly

Interestingly, MDSCs have also been shown to indirectly suppress T cell activation by inducing other immunosuppressive cells such as Treg cells and M2 macrophages. A3-47 The MDSCs accumulation in the inflammatory pathogenic setting suggests that increased MDSC numbers play a protective role by providing an anti-inflammatory mechanism for the maintenance of homeostasis; however, in the tumor environment, the anti-inflammatory effect of MDSCs leads to attenuation of antitumor immunity. Thus, targeting the inhibitory functions of MDSCs present a promising approach to enhance therapeutic potential of antitumor vaccines and immunotherapies.

Anti-Gr-1 monoclonal antibodies have been widely used to deplete MDSCs *in vivo* in mice. However, the clinical translation of anti-Gr-1 antibodies to deplete MDSCs in humans is challenging, since they recognize both Ly6G and Ly6C (molecules that express the Gr-1 epitope on MDSCs), which could lead to non-selective

depletion of monocytes, T cells, natural killer cells and macrophages and might impair host immunity and lead to opportunistic infections. ⁴⁸ Recently, pre-clinical and clinical studies have shown that promoting the differentiation of immature suppressive MDSCs into mature, non-suppressive cells such as myeloid DCs, CD4⁺ and CD8⁺ T cells using all trans retinoic acid ^{49–51} and Vitamin D3^{52,53} can enhance the antitumor immune responses. A number of strategies have been used to deplete MDSCs, including the use of chemotherapeutic drug gemcitabine, ^{45,54} sunitinib, ^{55–57} 5-FU, ⁵⁸ docetaxel, ⁵⁹ retinoic acid ^{50,60} or the debulking of tumors. ^{61,62} Furthermore, multiple inhibitor molecules, such as CDDO-ME, ⁶³ PDE-5, ⁶³ COX-2 ^{64,65} and nitro aspirin ⁶⁶ have also been successfully shown to inhibit the suppressive function MDSC, restore T cell proliferation and decrease tumor growth *in vivo*. These MDSCs depletion and inhibitory approaches can improve immune surveillance and promote antitumor immune responses, thus targeting MDSCs may be a promising approach for enhancing the efficacy of cancer vaccines or immunotherapies.

M2 macrophages

Tumor-associated macrophages are a dominant population of immune cells present in the tumor microenvironment and are mostly characterized as alternatively-activated M2-like macrophages, which are anti-inflammatory, immunosuppressive and facilitate tumor progression, 67.68 unlike M1 macrophages, which are highly inflammatory, microbicidal and tumoricidal. M2 macrophages play a significant role in promoting tumor growth, angiogenesis, metastasis, matrix remodeling and facilitate immune evasion in various human and animal tumors. 68-70 M2 macrophages also provide chemotherapy resistance, 71,72 radiotherapy resistance, and promote tumor growth. Therefore, targeting M2 macrophages is now considered a promising approach for treatment of cancer.

A number of approaches have been employed to target M2

A number of approaches have been employed to target M2 macrophages in tumors. Firstly, macrophage recruitment into the tumor microenvironment can be suppressed by inhibiting chemoattractants, such as CCL2, macrophage colony-stimulating factor or C-C motif chemokine receptor 2 (CCR2) and macrophage colony-stimulating factor receptor (M-CSFR); this has reduced tumor growth and enhanced prognosis in multiple cancers. ^{74–80} Secondly, M2 macrophage survival can be suppressed using chemical drugs, such as bisphosphonates, dasatinib, ^{81–83} or bacterial infection with attenuated *Shigella flexnen* ⁸⁴ that selectively kill M2 macrophages and result in regression of tumor growth, angiogenesis and metastasis. Alternatively, M2 macrophages can be depleted using immunotoxin-conjugated monoclonal anti-FRβ antibody, which target membrane molecules of M2 macrophages. ^{85,86} Lastly, the tumor promoting activity of M2 macrophages can be blocked using inhibitors of STAT3 (for example: sunitinib, ⁸⁷ sorafenib, ⁸⁸ Wp 1066. ⁸⁹ corosolic acid. ⁹⁰ and oleanolic acid. ⁹¹), STAT6, ⁹² c-Myc, ⁹³ PI3K, ⁶⁴ KLF4, ⁹⁵ HIFs, ⁹⁶ Ets2. ⁹⁷ and Mtor.

BLOCKADE OF IMMUNE CHECKPOINTS

Among the most promising approaches to activate therapeutic antitumor immunity is the blockade of the immune checkpoints. Immune checkpoints are inhibitory pathways employed by the immune system to maintain self-tolerance and thereby prevent the development of autoimmunity.³ They also help to regulate the duration and amplitude of physiological immune responses against pathogens in order to minimize collateral tissue damage and prevent immunopathology during infection. There is increasing evidence that tumors exploit certain immune checkpoint pathways and thereby subvert antitumor immunity, especially T cell responses specific for tumor antigens. Fortunately, most of the immune checkpoint pathways are initiated by ligand-receptor

interactions, thus they can be readily blocked using neutralizing monoclonal antibodies and inhibitory peptides or modulated using recombinant forms of ligands or receptors. This can help to drive effector immune responses and can enhance the efficacy of tumor vaccines and immunotherapeutics.

T cell—APC interaction involves cell surface co-stimulatory and co-inhibitory molecules, 99 as well as membrane receptors that are involved in adhesion and migration. These include members of the immunoglobulin superfamily, such as CD80/CD86-CD28 molecules and the TNF receptor (TNFR) superfamily, such as CD40-CD40L, Antagonizing the interaction between co-inhibitory cell surface receptors with neutralizing antibodies or inhibitory peptides can block the tolerogenic pathways. For example, blockade of CTLA-4 pathway and programmed cell death protein 1 (PD-1)-PDL-1 (also known as B7 homolog 1; B7H1) pathway can enhance immune responses. DCs from ovarian carcinomas overexpress B7H-1 (PD-1) and blockade of B7-H1 reduced IL-10 and enhanced IFN-γ production by T cells, thereby reduced the growth of human ovarian carcinoma in non-obese diabetic-severe combined immunodeficient mice. 100 Furthermore, B7H4 expression in gastric cancer is associated with poor survival.⁽³⁾ Expression of B7-H4 by APC is enhanced by Treg cells through IL-10 production, ¹⁰² and blockade of B7-H4 enhances antitumor T cell responses.¹⁰³ Alternatively, agonists that induce signaling via co-stimulatory cell surface receptors, such as CD28, CD137, CD27 and CD40 have been used to enhance lymphocyte priming to promote antitumor immune responses. Ipilimumab, a CTLA-4-specific monoclonal antibody, developed by Bristol Myers Squibb, approved by the FDA in 2011 for the treatment of metastatic melanoma, was the first immunotherapeutic based on the blockade of immune checkpoints. However, the inhibition of CTLA-4 is associated with immune-related adverse events, ¹⁰⁵ including colitis/diarrhea, dermatitis. hepatitis and endocrinopathies, ¹⁰⁶ Another dermatitis, hepatitis and endocrinopathies. ¹⁰⁶ Another CTLA-4-specific monoclonal antibody, tremelimumab, is currently being evaluated for the treatment of several other cancers as a monotherapy, or as an adjuvant in a DC vaccine.

An antibody that targets the co-inhibitory molecule PD-1, BMS 936558, has been evaluated in clinical trials and has shown significant and durable responses in several types of refractory tumors. This antibody was designed to represent newer generation of immunomodulatory biologics that stimulate highly effective and long lasting host tumor immunity with controllable autoimmune toxicities. However, endocrine toxicities have been reported with this antibody. Other drugs that target PD-1, including MK3475, CT-011 and AMP-224 are currently in clinical trials for different cancers. Furthermore, Genentech, AstraZeneca and Roche have antibodies against PDL-1 under clinical evaluation for solid tumors, melanoma or renal cell cancer. AstraZeneca immune checkpoints inhibitors under evaluation include those that target lymphocyte activation gene 3, 110,111 2B4 (also known as CD244), B and T lymphocyte attenuator, 112 T cell membrane protein 3 113,114 and adenosine A2a receptor. 115

A number of immunotherapeutics based on multiple antibody and immunoglobulin fusion proteins targeting co-inhibitory molecules are in pre-clinical and clinical trials (for example, IMP321 against lymphocyte activation gene 3 is in phase III clinical trial in breast cancer). Alternatively, the direct activation of co-stimulatory molecule signaling pathways using agonists, such as the monoclonal antibody TGN1412 specific for CD28 has the capacity to stimulate naïve human T cells without the need for a T cell receptor signal. This has been evaluated in clinical trials. However, within 90 min of infusion of the antibody, severe systemic inflammatory responses were induced, characterized by a massive pro-inflammatory cytokine storm followed by multi-organ injury and lymphocyte depletion. Fortunately, all volunteers survived after receiving immunosupressive treatments and cardiopulmonary support, and this trial

cautioned against the global non-discriminatory stimulation of naı̈ve T cell activation. ¹¹⁶

TLR LIGANDS AS IMMUNOTHERAPEUTICS AND ADJUVANTS FOR CANCER VACCINES

A new approach to the development of tumor vaccines and immunotherapies has focused on enhancement of effector T cell responses by targeting innate immune cells and the receptors that mediate their activation. Pathogen recognition receptors, in particular TLRs, sense pathogen-associated molecular patterns and are the primary targets for activation of the innate immune cells and are fundamental in the development of effective adaptive immune responses. Innate immune cells not only act as APC for T cells but also provide signals necessary for activation. Ligand binding to the TLRs, such as lipopolysaccharide to TLR4, flagellin to TLR5, CpG to TLR9, activates downstream intracellular signaling cascades through the transcription factors NF-kB, interferon regulatory factor 3, mitogen-activated protein kinases leading to the production of pro-inflammatory cytokines and type I interferons, which in turn activate co-stimulatory molecules, which are necessary for the antigen-presenting function of DCs and macrophages to activate T cells. Therefore, TLR agonists are potent activators of innate and adaptive immune responses and thus can act as adjuvants to promote immune responses against tumor antigens.

Synthetic ligands for TLR4, TLR7/8 and TLR9 have been through clinical trials against cancer 117,118 and the TLR7 agonist, imiquimod (Aldara) has been licensed for clinical use for the treatment of superficial basal cell carcinoma. However, unsatisfactory results have been obtained from clinical trials with other TLR agonists, and one pharmaceutical company halted its clinical program for PF-3512676, a CpG oligodeoxynucleotide that activates TLR9. Terthermore, the TLR4 agonists OK-432 and OK-PSA have been evaluated as immunotherapeutics for uterine, cervical and non-small lung cancer, but did not show promising results in most patients. Tender 120,121

In addition to the use of TLR agonists as direct tumor immunotherapeutics, TLR agonists have also been used as adjuvants for co-administration with tumor antigens, peptides, recombinant proteins or killed tumor cells. The detoxified derivative of lipopolysaccharide, monophosphoryl lipid A has shown promise as an adjuvant for vaccines against multiple tumors. ¹²² Currently, a TLR3 agonist, hiltonol, is being evaluated in clinical trials as an adjuvant for NY-ESO-1 protein vaccine in patients with high-risk melanoma. ¹²³ A study of hiltonol administered with tumor lysate-pulsed DC vaccines is also ongoing in patients with malignant glioma. ¹²³ Immunization of melanoma patients with the melanoma antigen Melan-A/MART-1 formulated in incomplete Freund's adjuvant with CpG-ODN induced strong antigen-specific CD8 + T-cell responses. ¹²⁴

Studies in mice have demonstrated mixed results with TLR agonists as adjuvants for tumor vaccines. Prophylactic but not therapeutic immunization with CpG-ODN in a transgenic mouse model expressing SV40 T Ag prevented tumor growth. The Furthermore, immunization of Tg mice expressing the Her-2/neu gene product with a synthetic peptide specific for Her-2/neu in combination with CpG-ODN prevented tumor growth in mice. The Furthermore, the efficacy of a DC tumor vaccine was enhanced with the use of CpG as an adjuvant in the presence of a p38 mitogen-activated protein kinases signaling inhibitor. Finally, the therapeutic efficacy of adenovirus expressing human tyrosinase-related protein 2 was enhanced with a peritumoral injection of CpG and poly(I:C) (a synthetic ligand for TLR3). 128

It has also been demonstrated that TLR agonists have the potential to break tolerance to self-antigens by inhibiting the

function of immunosuppressive cells such as Tregs. $^{129-131}$ TLR induced IL-6 production by DCs blocked the suppressive function of CD4 + CD25 + Treg cells. 129 The TLR3 agonist Poly(I:C) has been shown to enhance tumor-suppressing M1 macrophages at the expense of tumor-supporting M2 macrophages and thus inhibit the production of anti-inflammatory cytokines IL-10 and TGF- β . 132,133 Furthermore, lipopolysaccharide and CpG can enhance activation of NF- κ B pathways important for the establishment of the M1 phenotype of macrophages and their production of the inflammatory cytokines IL-12, IFN- α / β and TNF- α . 138,134 It is now recognized that TLR agonists are effective adjuvants for cancer vaccines through their ability to promote the differentiation of IFN- γ -secreting Th1 cells. However, TLR2 agonists have the potential to inhibit effector T cell responses 135 and all TLR agonists stimulate IL-10 as well as IL-12 production from DC and thereby promote induction of IL-10 + Treg as well as Th1 cells. 136 This has lead to the idea of using combinations of TLR activation with inhibitors of Treg cell induction to promote more effective antitumor immunity. 137

COMBINATION STRATEGIES FOR NEXT-GENERATION CANCER IMMUNOTHERAPEUTICS AND VACCINES

Our increased understanding of the tumor microenvironment and the immune responses that mediate and regulate antitumor immunity has provided a foundation for the rational development of vaccines and immunotherapeutic approaches for the treatment of cancer. Whilst vaccines and immunotherapeutics that target one arm of the immune system have shown some promising results in generating antitumor responses, many of these are subverted by a failure to block the regulatory responses associated with the tumor. Conversely, strategies that globally suppress Treg cells increase the risks for the patient of developing autoimmune diseases. The design of combination vaccines or therapies that activate the inflammatory signaling pathways that target the tumor and specifically block inhibitory pathways in the immunosuppressive tumor microenvironment although challenging may be the way forward (Figure 2).

There have already been a number of studies in mice that have demonstrated the benefits of using combination approaches that

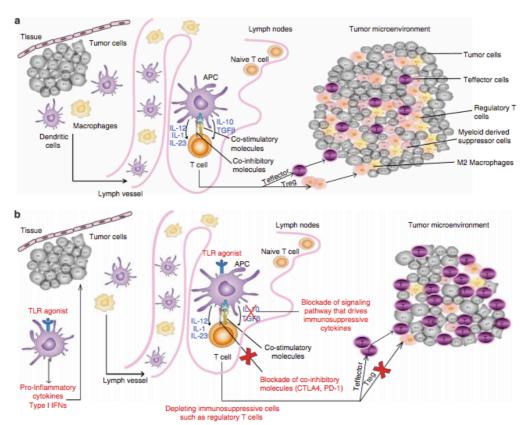


Figure 2. Combination immunotherapeutic/vaccine approaches against tumors. (a) In the initial stages of immune response against tumors, the local tissue resident innate immune cells, including macrophages and DCs recognizes the tumor antigens, are activated and migrate to regional lymph nodes via the lymphatic vessels, where they activate the naïve T cells by presenting the tumor antigens. Co-stimulatory and co-inhibitory molecules on APC interact with ligands on T cells, leading to the proliferation of effector and regulatory T cells respectively. A high percentage of immunosuppressive cells, including Treg cells, MDSCs, M2 macrophages, infiltrate the tumor microenvironment, which inhibit the effector immune responses against tumor. (b) Combination vaccine or immunotherapeutic approaches that target the immune response pathway are effective means of enhancing antitumor immunity. Strategies that combine innate cell activation with TLR agonists with depletion or inhibition of Tregs and MDSCs, or blockade of co-inhibitory molecules can enhance effector T cell responses against the tumor.

target multiple points in the immune system to enhance the antitumor immunity. For example, a study has demonstrated that the efficacy of DC vaccine against B16 melanoma in mice can be enhanced by the gene silencing of TGF- β 1, which reduced the tumor-associated Treg cells.¹³⁸ Furthermore, it has been shown that a single dose use of cyclophosphamide can reduce the numbers of Treg cells and enhance immune responses to the renal cell cancer vaccine IMA901, thus leading to prolonged survival of cancer patients after vaccination. 139 Also, a combination of the TLR agonist flagellin and a class I PI3 kinase inhibitor has been shown to block induction of Treg cells and promote effector T cells that mediate rejection of tumors in While PI3 kinase inhibitors are already in clinical trials based on their ability to arrest cell proliferation and induce tumor cell death, this study demonstrated that PI3 kinase inhibitors also attenuated TLR-induced IL-10 and TGF-B and thereby inhibited induction of Treg cells. Furthermore, gemcitabine has been shown to reduce the numbers of splenic MDSC in tumor-bearing mice and enhance antitumor immunity when used in combination with IFN-β.⁵⁴ In a HER-2/neu tumor model, therapeutic administration of anti-glucocorticoid-induced TNF receptor monoclonal antibody with a HER-2/neu vaccine enhanced vaccine efficacy and protected against pre-existing tumors.1

The benefits of combination approaches to inhibit immune checkpoints has recently been demonstrated with the report that the blockade of PD-1/PD-L1 pathway using anti-PD-L1 neutralizing antibodies or depletion of Treg cells alone failed to prevent recurrence of tumors, whereas the combination of PD-L1 blockade with Treg depletion effectively mediated disease regression. Furthermore, immune checkpoint blockade using anti-PD-L1 and anti-LAG-3 antibodies overcame the requirement to deplete tumor-specific Tregs. 141 Double immune checkpoint blockade with ipilimumab (anti-CTLA-4) and nivolumab/BMS-936558 (anti-PD-1) has recently been evaluated in a phase I clinical trial in patients with advance melanoma. 142 The results revealed that the combination treatment induced clinical activity in 65% of patients, with tumor regression of 80% or more in 53% of patients at the maximum doses associated with acceptable side effects. This combination may prove to be powerful for unleashing immune responses to melanoma, provided the incidences of autoimmune adverse effects are manageable.

Blockade of immune checkpoints or the inhibition of immunosuppressive cells are clearly promising approaches for the treatment of cancer. However, global blocking of the antiinflammatory arm of the immune system alone significantly enhances inflammatory responses, which can lead to the breaking of immune self-tolerance, thereby inducing autoimmune/auto inflammatory side effects including rash, colitis, hepatitis and endocrinopathies (summarized by Corsello *et al.* 2013). ¹⁴³ Generalized non-specific activation of the immune system as a result of blockade of immunosuppressive cells or checkpoints when combined with immune stimulators also has the potential to generate systemic inflammation. Thus a major challenge in the blockade of immune checkpoints alone or in combination with immune activators is to avoid exceeding the limits of safety associated with removing the natural breaks on the immune system that may lead to unacceptable toxicities. We believe that this can be achieved by employing strategies that specifically enhance tumor-specific immune responses, while transiently blocking immunosuppressive networks, not systemically but locally in the tumor or the draining lymph nodes. More specific inhibition of tumor associated immunosuppressive cytokines and cells and careful and controlled activation of immune system has the potential to generate antitumor immune responses without excessive systemic inflammation or autoimmune diseases. One approach to achieve this objective involves the use of TLR appnists as immunotherapeutics or adjuvants in tumor vaccines, in combination with transiently depleting immune suppressive

cells or blockade of immune checkpoints at the time of immunization in the vicinity of the tumor. For example, it has been shown that the efficacy of TLR agonists as therapeutics or as adjuvants in a DC vaccine were enhanced when Treg cells induction in the tumor microenvironment were attenuated through inhibition of p38 mitogen-activated protein kinases or Pi3K signaling. ^{127,136} Transient inhibition of Treg cells at the time of immunization generated potent antitumor immunity and immunological memory and is less likely to be associated with systemic autoimmunity. Therefore, combination approaches that specifically inhibit tumor-associated Treg and innate suppressor cells with concomitant enhancement of antitumor effector immune responses have considerable potential as safe and effective cancer immunotherapeutics and vaccines of the future.

CONFLICT OF INTEREST

KHG Mills is a co-founder and shareholder in Opsona Therapeutics and TriMod Therapeutics. TriMod Therapeutics is involved in the development of immunotherapeutics for cancer. Aisha Q. Butt declares no conflict of interest.

ACKNOWLEDGEMENTS

Kingston Mills's research group is supported by grants from Science Foundation

REFERENCES

- 1 Koebel CM, Vermi W, Swann JB, Zerafa N, Rodig SJ, Old LJ et al. Adaptive immunity maintains occult cancer in an equilibrium state. Nature 2007; 450: 903-907.
- 2 Smyth MJ, Dunn GP, Schreiber RD. Cancer immunosurveillance and immunoediting: the roles of immunity in suppressing tumor development and shaping tumor immunogenicity. Adv Immunol 2006; 90: 1–50.
- 3 Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 2012: 12: 252-264.
- 4 Byrne WL, Mills KH, Lederer JA, O'Sullivan GC. Targeting regulatory T cells in cancer. Cancer Res 2011; 71: 6915–6920.
- 5 Baxevanis CN, Perez SA, Papamichail M. Combinatorial treatments including vaccines, chemotherapy and monoclonal antibodies for cancer therapy. Cancer Immunol Immunother 2009; 58: 317–324.
- 6 Banchereau J, Palucka AK. Dendritic cells as therapeutic vaccines against cancer. Nat Rev Immunol 2005: 5: 296–306.
- 7 Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. N Engl J Med 2010: 363: 411–422.
- 8 Mellef CJ. Cancer immunotherapy by dendritic cells. Immunity 2008; 29: 372–383.
- Palucka K, Ueno H, Zurawski G, Fay J, Banchereau J. Building on dendritic cell subsets to improve cancer vaccines. Curr Opin Immunol 2010; 22: 258–263.
 Love WE, Bernhard JD, Bordeaux JS. Topical imiquimod or fluorouracil therapy
- Love WE, Bernhard JD, Bordeaux JS. Topical imiquimod or fluorouracil therapy for basal and squamous cell carcinoma: a systematic review. Arch Dermatol 2009; 145: 1431-1438.
- 11 Golgher D, Jones E, Powrie F, Elliott T, Gallimore A. Depletion of CD25 + regulatory cells uncovers immune responses to shared murine tumor rejection antigens. Eur J Immunol 2002; 32: 3267–3275.
- 12 Steltz J, Bruck J, Lenz J, Knop J, Tuting T. Depletion of CD25(+) CD4(+) T cells and treatment with tyrosinase-related protein 2-transduced dendritic cells enhance the interferon alpha-induced, CD8(+) T-cell-dependent immune defense of B16 melanoma. Cancer Res 2001; 61: 8643–8646.
- 13 Jamicki AG, Lysaght J, Todryk S, Mills KH. Suppression of antitumor immunity by IL-10 and TGF-beta-producing T cells infiltrating the growing tumor. influence of tumor environment on the induction of CD4+ and CD8+ regulatory T cells. J Immunol 2006; 177: 896–904.
- 14 Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 2012; 366: 2455–2465.
- 15 Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med 2012; 366: 2443–2454.
- 16 Corsello SM, Barnabei A, Marchetti P, De Vecchis L, Salvatori R, Torino F. Endocrine side effects induced by immune checkpoint inhibitors. J Clin Endocrinol Metab 2013; 98: 1361–1375.

- 17 Mills KH. Regulatory T cells: friend or foe in immunity to infection? Nat Rev Immunol 2004; 4: 841–855.
- 18 Bluestone JA, Abbas AK. Natural versus adaptive regulatory T cells. Nat Rev Immunol 2003; 3: 253–257.
- 19 Woo EY, Chu CS, Goletz TJ, Schlienger K, Yeh H, Coukos G et al. Regulatory CD4(+)CD25(+) T cells in tumors from patients with early-stage non-small cell lung cancer and late-stage ovarian cancer. Cancer research. 2001; 61: 4766–4772.
- 20 Mouglakakos D, Choudhury A, Lladser A, Kiessling R, Johansson CC. Regulatory T cells in cancer. Adv Cancer Res 2010: 107: 57–117.
- 21 Wang RF, Peng G, Wang HY. Regulatory T cells and Toll-like receptors in tumor immunity. Semin Immunol 2006; 18: 136–142.
- 22 Hoon DŚ, Foshag LJ, Nizze AS, Bohman R, Morton DL. Suppressor cell activity in a randomized trial of patients receiving active specific immunotherapy with melanoma cell vaccine and low dosages of cyclophosphamide. Cancer Res 1990; 50: 5358–5364.
- 23 Rasku MA, Clem AL, Telang S, Taft B, Gettings K, Gragg H et al. Transient T cell depletion causes regression of melanoma metastases. J Transi Med 2008; 6: 12.
- 24 Rech AJ, Vonderheide RH. Clinical use of anti-CD25 antibody daclizumab to enhance immune responses to tumor antigen vaccination by targeting regulatory T cells. Ann NY Acad Sci 2009; 1174: 99–106.
- 25 Morita R, Hirohashi Y, Sato N. Depletion of Tregs in vivo: a promising approach to enhance antitumor immunity without autoimmunity. *Immunotherapy* 2012; 4: 1103–1105.
- 26 Ko K, Yamazaki S, Nakamura K, Nishioka T, Hirota K, Yamaguchi T et al. Treatment of advanced tumors with agonistic anti-GITR mAb and its effects on tumorinfiltrating Foxp3+CD25+CD4+ regulatory T cells. J Exp Med 2005; 202: 885-891
- 27 Coe D, Begom S, Addey C, White M, Dyson J, Chai JG. Depletion of regulatory T cells by anti-GITR mab as a novel mechanism for cancer immunotherapy. Cancer immunol immunother 2010; 59: 1367–1377.
- 28 Tan W, Zhang W, Strasner A, Grivennikov S, Cheng JQ, Hoffman RM et al. Tumour-infiltrating regulatory T cells stimulate mammary cancer metastasis through RANKL-RANK signalling. Nature 2011; 470: 548–553.
- 29 Amendola M, Passerini L, Pucci F, Gentner B, Bacchetta R, Naldini L. Regulated and multiple miRNA and siRNA delivery into primary cells by a lentiviral platform. Mol Ther 2009; 17: 1039–1052.
- 30 Casares N, Rudilla F, Arribillaga L, Llopiz D, Riezu-Boj JI, Lozano T et al. A peptide inhibitor of FOXP3 impairs regulatory T cell activity and improves vaccine efficacy in mice. J Immunol 2010; 185: 5150–5159.
- 31 Karanikas V, Speletas M, Zamanakou M, Kalala F, Loules G, Kerenidi T et al. Foxp3 expression in human cancer cells. J Transl Med 2008: 6: 19.
- 32 Zou L, Barnett B, Safah H, Larussa VF, Evdemon-Hogan M, Mottram P et al. Bone marrow is a reservoir for CD4+CD25+ regulatory T cells that traffic through CXCL12/CXCR4 signals. Cancer Res 2004; 64: 8451–8455.
- 33 Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. Nat Med 2004: 10: 942–949.
- 34 Zhao E, Wang L, Dai J, Kryczek I, Wei S, Vatan L et al. Regulatory T cells in the bone marrow microenvironment in patients with prostate cancer. Oncoimmunology 2012; 1:152–161
- 35 Youn Ji, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. J Immunol 2008; 181: 5791–5802.
- 36 Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. Nat Rev Immunol 2009; 9: 162–174.
- 37 Sinha P, Clements VK, Fulton AM, Ostrand-Rosenberg S. Prostaglandin E2 promotes tumor progression by inducing myeloid-derived suppressor cells. Cancer Res 2007; 67: 4507–4513.
- 38 Ostrand-Rosenberg S, Sinha P. Myeloid-derived suppressor cells: linking inflammation and cancer. J Immunol 2009; 182: 4499–4506.
- 39 Peranzoni E, Zillo S, Marigo I, Dolcetti L, Zanovello P, Mandruzzato S et al. Myeloid-derived suppressor cell heterogeneity and subset definition. Curr Opin Immunol 2010; 22: 238–244.
- 40 Bunt SK, Clements VK, Hanson EM, Sinha P, Ostrand-Rosenberg S. Inflammation enhances myeloid-derived suppressor cell cross-talk by signaling through Tolllike receptor 4. J Leukocyte Biol 2009; 85: 996–1004.
- 41 Delano MJ, Scumpia PO, Weinstein JS, Coco D, Nagaraj S, Kelly-Scumpia KM et al. MyD88-dependent expansion of an immature GR-1(+)CD11b(+) population induces T cell suppression and Th2 polarization in sepsis. J Exp Med 2007; 204:
- 42 Bronte V, Wang M, Overwijk WW, Surman DR, Pericle F, Rosenberg SA et al. Apoptotic death of CD8+ T lymphocytes after immunization: induction of a suppressive population of Mac-1+/Gr-1+ cells. J Immunol 1998; 161: 5313-5320.
- 43 Hoechst B, Gamrekelashvili J, Manns MP, Greten TF, Korangy F. Plasticity of human Th17 cells and iTregs is orchestrated by different subsets of myeloid cells. Blood 2011; 117: 6532–6541.

- 44 Serafini P, Mgebroff S, Noonan K, Borrello I. Myeloid-derived suppressor cells promote cross-tolerance in B-cell lymphoma by expanding regulatory T cells. Cancer Res 2008: 68: 5439–5449.
- 45 Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S. Cross-talk between myeloid-derived suppressor cells and macrophages subverts tumor immunity toward a type 2 response. J Immunol 2007; 179: 977–983.
- 46 Huang B, Pan PY, Li Q, Sato AI, Levy DE, Bromberg J et al. Gr-1+CD115+ immature myeloid suppressor cells mediate the development of tumor-induced T regulatory cells and T-cell anergy in tumor-bearing host. Cancer Res 2006; 66: 1123–1131
- 47 Ostrand-Rosenberg S. Myeloid-derived suppressor cells: more mechanisms for inhibiting antitumor immunity. Cancer Immunol Immunother 2010; 59: 1593–1600.
- 48 Stewart TJ, Smyth MJ. Improving cancer immunotherapy by targeting tumorinduced immune suppression. Cancer Metastasis Rev 2011; 30: 125–140.
- 49 Gabrilovich DI, Velders MP, Sotomayor EM, Kast WM. Mechanism of immune dysfunction in cancer mediated by immature Gr-1+ myeloid cells. J Immunol 2001; 166: 5398–5406.
- 50 Mirza N, Fishman M, Fricke I, Dunn M, Neuger AM, Frost TJ et al. All-trans-retinoic acid improves differentiation of myeloid cells and immune response in cancer patients. Cancer Res 2006; 66: 9299–9307.
- 51 Nefedova Y, Fishman M, Sherman S, Wang X, Beg AA, Gabrilovich DI. Mechanism of all-trans retinoic acid effect on tumor-associated myeloid-derived suppressor cells. Cancer Res 2007; 67: 11021–11028.
- 52 Lathers DM, Clark JI, Achille NJ, Young MR. Phase 18 study to improve immune responses in head and neck cancer patients using scalating doses of 25hydroxyvitamin D3. Cancer Immunol Immunother 2004; 53: 422–430.
- 53 Ugel S, Delpozzo F, Desantis G, Papalini F, Simonato F, Sonda N et al. Therapeutic tameting of myeloid-derived suppressor cells. Curr Opin Pharmacol 2009; 9: 470–481.
- targeting of myeloid-derived suppressor cells. Curr Opin Pharmacol 2009; 9: 470–481.

 54 Suzuki E, Kapoor V, Jassar AS, Kaiser LR, Albelda SM. Gemcitabine selectively eliminates splenic Gr-1 + /CD11b + myeloid suppressor cells in tumor-bearing animals and enhances antitumor immune activity. Clin Cancer Res 2005; 11: 6713–6721.
- Ko JS, Rayman P, Ireland J, Swaidani S, Li G, Bunting KD et al. Direct and differential suppression of myeloid-derived suppressor cell subsets by sunitinib is compartmentally constrained. Cancer Res 2010; 70: 3526-3536.
 Ko JS, Zea AH, Rini Bi, Ireland JL, Elson P, Cohen P et al. Sunitinib mediates
- 56 Ko JS, Zea AH, Rini BI, Ireland JL, Elson P, Cohen P et al. Sunitinib mediates reversal of myeloid-derived suppressor cell accumulation in renal cell carcinoma patients. Clin Cancer Res 2009; 15: 2148–2157.
- 57 Ozao-Choy J, Ma G, Kao J, Wang GX, Meseck M, Sung M et al. The novel role of tyrosine kinase inhibitor in the reversal of immune suppression and modulation of tumor microenvironment for immune-based cancer therapies. Cancer Res 2009; 69: 2514–2522.
- 58 Vincent J, Mignot G, Chalmin F, Ladoire S, Bruchard M, Chevriaux A et al. 5-Fluorouracil selectively kills tumor-associated myeloid-derived suppressor cells resulting in enhanced T cell-dependent antitumor immunity. Cancer Res 2010; 70: 3053-3061
- 59 Kodumudi KN, Woan K, Gilvary DL, Sahakian E, Wei S, Djeu JY. A novel chemoinmunomodulating property of docetaxel: suppression of myeloid-derived suppressor cells in tumor bearers. Clin Cancer Res 2010: 16: 4583–4594.
- 60 Kusmartsev S, Cheng F, Yu B, Nefedova Y, Sotomayor E, Lush R et al. All-transretinoic acid eliminates immature myeloid cells from tumor-bearing mice and improves the effect of vaccination. Cancer Res 2003; 63: 4441–4449.
- Insproves the effect of vaccination. Cancer Nes 2005; 63: 4941-44499.
 61 Sinha P, Clements VK, Ostrand-Rosenberg S. Interleukin-13-regulated M2 macrophages in combination with myeloid suppressor cells block immune surveillance against metastasis. Cancer Res 2005; 65: 11743-11751.
- 62 Sinha P, Clements VK, Miller S, Ostrand-Rosenberg S. Tumor immunity: a balancing act between T cell activation, macrophage activation and tumor-induced immune suppression. Cancer Immunol Immunother 2005; 54: 1137–1142.
- 63 Nagaraj S, Youn JI, Weber H, Iclozan C, Lu L, Cotter MJ et al. Anti-inflammatory triterpenoid blocks immune suppressive function of MDSCs and improves immune response in cancer. Clin Cancer Res 2010; 16: 1812–1823.
- 64 Fujita M, Kohanbash G, Fellows-Mayle W, Hamilton RL, Komohara Y, Decker SA et al. COX-2 blockade suppresses gliomagenesis by inhibiting myeloid-derived suppressor cells. Cancer Res 2011: 71: 2664–2674.
- 65 Obermajer N, Muthuswamy R, Lesnock J, Edwards RP, Kalinski P. Positive feed-back between PGE2 and COX2 redirects the differentiation of human dendritic cells toward stable myeloid-derived suppressor cells. *Blood* 2011; 118: 5498–5505.
- 66 De Santo C, Serafini P, Marigo I, Dolcetti L, Bolla M, Del Soldato P et al. Nitroaspirin corrects immune dysfunction in tumor-bearing hosts and promotes tumor eradication by cancer vaccination. Proc Natl Acad Sci USA 2005; 102: 4185–4190.
- 67 Allavena P, Sica A, Solinas G, Porta C, Mantovani A. The inflammatory microenvironment in tumor progression: the role of tumor-associated macrophages. Crit Rev Oncol Hematol 2008; 66: 1–9.

- 68 Solinas G, Germano G, Mantovani A, Allavena P. Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation. J Leukoc Biol 2009; 86: 1065–1073.
- 69 Sica A, Bronte V. Altered macrophage differentiation and immune dysfunction in tumor development. J Clin Invest 2007; 117: 1155–1166.
- 70 Sica A, Larghi P, Mancino A, Rubino L, Porta C, Totaro MG et al. Macrophage polarization in tumour progression. Semin Cancer Biol 2008; 18: 349–355.
- 71 Fischer C, Jonckx B, Mazzone M, Zacchigna S, Loges S, Pattarini L et al. Anti-PIGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. Cell 2007; 131: 463–475.
- 72 Zhang W, Zhu XD, Sun HC, Xiong YQ, Zhuang PY, Xu HX et al. Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. Clin Cancer Res 2010; 16: 3420–3430.
- 73 Ahn GO, Tseng D, Liao CH, Dorie MJ, Czechowicz A, Brown JM. Inhibition of Mac-1 (CD11b/CD18) enhances tumor response to radiation by reducing myeloid cell recruitment. Proc Natl Acad Sci USA 2010; 107: 8363–8368.
- 74 Mizutani K, Sud S, McGregor NA, Martinovski G, Rice BT, Craig MJ et al. The chemokine CCL2 increases prostate tumor growth and bone metastasis through macrophage and osteoclast recruitment. Neoplasia 2009; 11: 1235–1242.
- 75 Qian BZ, Li J, Zhang H, Kitamura T, Zhang J, Campion LR et al. CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. Nature 2011; 475: 222-225.
- 76 Zhu X, Fujita M, Snyder LA, Okada H. Systemic delivery of neutralizing antibody targeting CCL2 for glioma therapy. J Neuropicol 2011; 104: 83–92.
- targeting CCL2 for glioma therapy. J Neurooncol 2011; 104: 83–92.
 77 Kubota Y, Takubo K, Shimizu T, Ohno H, Kishi K, Shibuya M et al. M-CSF inhibition selectively targets pathological angiogenesis and lymphangiogenesis. J Exp Med 2009; 206: 1089–1102.
- 78 Manthey CL, Johnson DL, Illig CR, Tuman RW, Zhou Z, Baker JF et al. JNJ-28312141, a novel orally active colony-stimulating factor-1 receptor/FMS-related receptor tyrosine kinase-3 receptor tyrosine kinase inhibitor with potential utility in solid tumors, bone metastases, and acute myeloid leukemia. Mol Cancer Ther 2009; 8: 3151–3161.
- 79 Paulus P, Stanley ER, Schafer R, Abraham D, Aharinejad S. Colony-stimulating factor-1 antibody reverses chemoresistance in human MCF-7 breast cancer xenografts. Cancer Res 2006; 66: 4349–4356.
- 80 Pyonteck SM, Gadea BB, Wang HW, Gocheva V, Hunter KE, Tang LH et al. Deficiency of the macrophage growth factor CSF-1 disrupts pancreatic neuroendocrine tumor development. Oncoare. 2012; 31: 1459–1467.
- 81 Hiraoka K, Zenmyo M, Watari K, Iguchi H, Fotovati A, Kimura YN et al. Inhibition of bone and muscle metastases of lung cancer cells by a decrease in the number of monocytes/macrophages. Cancer Sci 2008; 99: 1595–1602.
- 82 Miselis NR, Wu ZJ, Van Rooijen N, Kane AB. Targeting tumor-associated macrophages in an orthotopic murine model of diffuse malignant mesothelioma. Mol Cancer Ther 2008: 7: 788–799.
- 83 Zeisberger SM, Ödermatt B, Marty C, Zehnder-Fjallman AH, Ballmer-Hofer K, Schwendener RA. Clodronate-liposome-mediated depletion of tumour-associated macrophages: a new and highly effective antiangiogenic therapy approach. Br J Cancer 2006; 95: 272–281.
- 84 Suzuki T, Franchi L, Toma C, Ashida H, Ogawa M, Yoshikawa Y et al. Differential regulation of caspase-1 activation, pyroptosis, and autophagy via Ipaf and ASC in Shigella-Infected macrophages. PLoS Pathog 2007; 3: e111.
- 85 Nagai T, Tanaka M, Tsuneyoshi Y, Xu B, Michie SA, Hasui K et al. Targeting tumorassociated macrophages in an experimental glioma model with a recombinant immunot to folate receptor beta. Cancer Immunol Immunother 2009; 58: 1577-1586.
- 86 Pulg-Kroger A, Sierra-Filardi E, Dominguez-Soto A, Samaniego R, Corcuera MT, Gomez-Aguado F et al. Folate receptor beta is expressed by tumor-associated macrophages and constitutes a marker for M2 anti-inflammatory/regulatory macrophages. Cancer Res 2009; 69: 9395–9403.
- 87 Xin H, Zhang C, Herrmann A, Du Y, Figlin R, Yu H. Sunitinib inhibition of Stat3 induces renal cell carcinoma tumor cell apoptosis and reduces immunosuppressive cells. Cancer Res 2009; 69: 2506–2513.
- 88 Edwards JP, Emens LA. The multikinase inhibitor sorafenib reverses the suppression of IL-12 and enhancement of IL-10 by PGE(2) in murine macrophages. Int Immunopharmacol 2010; 10: 1220-1228.
- 89 Hussain SF, Kong LY, Jordan J, Conrad C, Madden T, Fokt I et al. A novel small molecule inhibitor of signal transducers and activators of transcription 3 reverses immune tolerance in malignant glioma patients. Cancer Res 2007; 67: 9630–9636.
- 90 Fujiwara Y, Komohara Y, Ikeda T, Takeya M. Corosolic acid inhibits glioblastoma cell proliferation by suppressing the activation of signal transducer and activator of transcription-3 and nuclear factor-kappa B in tumor cells and tumor-associated macrophages. Cancer Sci 2011; 102: 206–211.

- 91 Fujiwara Y, Komohara Y, Kudo R, Tsurushima K, Ohnishi K, Ikeda T et al. Oleanolic acid inhibits macrophage differentiation into the M2 phenotype and glioblastoma cell proliferation by suppressing the activation of STAT3. Oncol Rep 2011; 26: 1533–1537.
- 92 Sinha P, Clements VK, Ostrand-Rosenberg S. Reduction of myeloid-derived suppressor cells and induction of M1 macrophages facilitate the rejection of established metastatic disease. J Immunol 2005; 174: 636–645.
- 93 Pello OM, De Pizzol M, Mirolo M, Soucek L, Zammataro L, Amabile A et al. Role of c-MYC in alternative activation of human macrophages and tumor-associated macrophage biology. Blood 2012; 119: 411–421.
- 94 Weisser SB, McLarren KW, Voglmaier N, an Netten-Thomas CJ, Antov A, Flavell RA et al. Alternative activation of macrophages by IL-4 requires SHIP degradation. Eur. J Immunol 2011; 41: 1742–1753.
- 95 Liao X, Sharma N, Kapadia F, Zhou G, Lu Y, Hong H et al. Kruppel-like factor 4 regulates macrophage polarization. J Clin Invest 2011; 121: 2736–2749.
- 96 Doedens AL, Stockmann C, Rubinstein MP, Llao D, Zhang N, DeNardo DG et al. Macrophage expression of hypoxia-inducible factor-1 alpha suppresses T-cell function and promptes tumor progression. Caper Res 2010: 70: 7465–7475.
- function and promotes tumor progression. Cancer Res 2010; 70: 7465–7475.
 97 Zabuawala T, Taffany DA, Sharma SM, Merchant A, Adair B, Srinivasan R et al. An ets2-driven transcriptional program in tumor-associated macrophages promotes tumor metastasis. Cancer Res 2010; 70: 1323–1333.
- 98 Chen W, Ma T, Shen XN, Xia XF, Xu GD, Bai XL et al. Macrophage-induced tumor angiogenesis is regulated by the TSC2-mTOR pathway. Cancer Res 2012; 72: 1363–1372.
- 99 Zhu Y, Yao S, Chen L. Cell surface signaling molecules in the control of immune
- responses: a tide model. Immunity 2011; 34: 466–478.
 100 Curiel TJ, Wel S, Dong H, Alvarez X, Cheng P, Mottram P et al. Blockade of B7-H1 improves myeloid dendritic cell-mediated antitumor immunity. Nat Med 2003; 9: 562–567.
- 101 Jiang J, Zhu Y, Wu C, Shen Y, Wei W, Chen L et al. Tumor expression of 87-H4 predicts poor survival of patients suffering from gastric cancer. Cancer Immunol Immunother 2010: 59: 1707–1714.
- 102 Kryczek I, Zou L, Rodriguez P, Zhu G, Wei S, Mottram P et al. 87-H4 expression identifies a novel suppressive macrophage population in human ovarian carcinoma. J Exp Med 2006; 203: 871–881.
- 103 Dangaj D, Lanitis E, Zhao A, Joshi S, Cheng Y, Sandaltzopoulos R et al. Novel recombinant human b7-h4 antibodies overcome tumoral immune escape to potentiate T-cell antitumor responses. Cancer Res 2013; 73: 4820–4829.
- 104 Yao S, Zhu Y, Chen L. Advances in targeting cell surface signalling molecules for immune modulation. Nat Rev Drug Discov 2013; 12: 130–146.
- 105 Di Giacomo AM, Biagioli M, Maio M. The emerging toxicity profiles of anti-CTLA-4 antibodies across clinical indications. Semin Oncol 2010; 37: 499–507.
- 106 Fong L, Small EJ. Anti-cytotoxic T-lymphocyte antigen-4 antibody: the first in an emerging class of immunomodulatory antibodies for cancer treatment. J Clin Oncol 2008; 26: 5275–5283.
- 107 Ralph C, Elkord E, Burt DJ, O'Dwyer JF, Austin EB, Stern PL et al. Modulation of lymphocyte regulation for cancer therapy: a phase II trial of tremelimumab in advanced gastric and esophageal adenocarcinoma. Clin Cancer Res 2010; 16: 1662–1672.
- 108 Chung KY, Gore I, Fong L, Venook A, Beck SB, Dorazio P et al. Phase II study of the anti-cytotoxic T-lymphocyte-associated antigen 4 monoclonal antibody, tremelimumab, in patients with refractory metastatic colorectal cancer. J Clin Oncol 2010; 28: 3485–3490.
- 109 Mullard A. New checkpoint inhibitors ride the immunotherapy tsunami. Nat Rev Drug Discov 2013; 12: 489–492.
- 110 Woo SR, Turnis ME, Goldberg MV, Bankoti J, Selby M, Nirschi CJ et al. Immune inhibitory molecules LAG-3 and PD-1 synergistically regulate T-cell function to promote tumoral immune escape. Cancer Res 2012; 72: 917–927.
- 111 Goldberg MV, Drake CG. LAG-3 in Cancer Immunotherapy. Curr Top Microbiol Immunol 2011; 344: 269–278.
- 112 Lanier LL. Up on the tightrope: natural killer cell activation and inhibition. Nat Immunol 2008; 9: 495–502.
- 113 Sakulshi K, Apetoh L, Sullivan JM, Biazar BR, Kuchroo VK, Anderson AC. Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore anti-tumor immunity. J Exp Med 2010; 207: 2187–2194.
- 114 Fourcade J, Sun Z, Benallaoua M, Guillaume P, Luescher IF, Sander C et al. Upregulation of Tim-3 and PD-1 expression is associated with tumor antigenspecific CD8+ T cell dysfunction in melanoma patients. J Exp Med 2010; 207: 2175–2186.
- 115 Waickman AT, Alme A, Senaldi L, Zarek PE, Horton M, Powell JD. Enhancement of tumor immunotherapy by deletion of the A2A adenosine receptor. Cancer Immunol Immunother 2012; 61: 917–926.
- 116 Suntharalingam G, Perry MR, Ward S, Brett SJ, Castello-Cortes A, Brunner MD et al. Cytokine storm in a phase 1 trial of the anti-CD28 monoclonal antibody TGN1412. New Engl J Med 2006; 355: 1018–1028.

- 117 Kanzler H, Barrat FJ, Hessel EM, Coffman RL. Therapeutic targeting of innate immunity with Toll-like receptor agonists and antagonists. Nat Med 2007; 13: 552–559.
- 118 Krieg AM. Therapeutic potential of Toll-like receptor 9 activation. Nat Rev Drug Discov 2006; 5: 471–484.
- 119 Schmidt C. Clinical setbacks for toll-like receptor 9 agonists in cancer. Nat Biotechnol 2007; 25: 825–826.
- 120 Watanabe Y, Iwa T. Clinical value of immunotherapy with the streptococcal preparation OK-432 in non-small cell lung cancer. J Biol Response Mod 1987; 6: 169–180.
- 121 Kikkawa F, Kawai M, Oguchi H, Kojima M, Ishikawa H, Iwata M et al. Randomised study of immunotherapy with OK-432 in uterine cervical carcinoma. Eur J Cancer 1993: 29A: 1542–1546.
- 122 Cluff CW. Monophosphoryl lipid A (MPL) as an adjuvant for anti-cancer vaccines: clinical results. Adv Exp. Med Blol 2010: 667: 111–123.
- 123 Sharma S, Zhu L, Davoodi M, Harris-White M, Lee St JM, John M et al. TLR3 agonists and proinflammatory antitumor activities. Expert Opin Ther Targets 2013; 17: 481–483.
- 124 Speiser DE, Llenard D, Rufer N, Rubio-Godoy V, Rimoldi D, Lejeune F et al. Rapid and strong human CD8 + T cell responses to vaccination with peptide, IFA, and Cp6 oligodeoxynucleotide 7909. J Clin Invest 2005; 115: 739-746.
- 125 Garbi N, Arnold B, Gordon S, Hammerling GJ, Ganss R. CpG motifs as proinflammatory factors render autochthonous tumors permissive for infiltration and destruction. J Immunol 2004; 172: 5861–5869.
- 126 Nava-Parada P, Forni G, Knutson KL, Pease LR, Celis E. Peptide vaccine given with a Toll-like receptor agonist is effective for the treatment and prevention of spontaneous breast tumors. Cancer Res 2007; 67: 1326–1334.
- 127 Jarnicki AG, Conroy H, Brereton C, Donnelly G, Toomey D, Walsh K et al. Attenuating regulatory T cell induction by TLR agonists through inhibition of p38 MAPK signaling in dendritic cells enhances their efficacy as vaccine adjuvants and cancer immunotherapeutics. J Immunol 2008; 180: 3797–3806.
- 128 Tormo D, Ferrer A, Bosch P, Gaffal E, Basner-Tschakarjan E, Wenzel J et al. Therapeutic efficacy of antigen-specific vaccination and toll-like receptor stimulation against established transplanted and autochthonous melanoma in mice. Cancer Res 2006; 66: 5427–5435.
- 129 Pasare C, Medzhitov R. Toll pathway-dependent blockade of CD4+CD25+ T cell-mediated suppression by dendritic cells. Science 2003; 299: 1033–1036.
- 130 Peng G, Guo Z, Kiniwa Y, Voo KS, Peng W, Fu T et al. Toll-like receptor 8-mediated reversal of CD4+ regulatory T cell function. Science 2005; 309: 1380–1384.

- 131 Yang Y, Huang CT, Huang X, Pardoll DM. Persistent Toll-like receptor signals are required for reversal of regulatory T cell-mediated CD8 tolerance. Nat Immunol 2004; 5: 508–515.
- 132 Seya T, Shime H, Matsumoto M. TAMable tumor-associated macrophages in response to innate RNA sensing. Oncolmmunology 2012; 1: 1000-1001.
- 133 Shime H, Matsumoto M, Oshiumi H, Tanaka S, Nakane A, Iwakura Y et al. Toll-like receptor 3 signaling converts tumor-supporting myeloid cells to tumoricidal effectors. Proc Natl Acad Sci USA 2012; 109: 2066–2071.
- 134 Wilson HL, Dar A, Napper SK, Marianela Lopez A, Babluk LA, Mutwirl GK. Immune mechanisms and therapeutic potential of CpG oligodeoxynucleotides. Int Rev Immunol 2006; 25: 183–213.
- 135 Akdis CA, Kussebi F, Pulendran B, Akdis M, Lauener RP, Schmidt-Weber CB et al. Inhibition of T helper 2-type responses, IgE production and eosinophilia by synthetic lipopeptides. Eur J Immunol 2003; 33: 2717–2726.
- 136 Marshall NA, Galvin KC, Corcoran AM, Boon L, Higgs R, Mills KH. Immunotherapy with Pi3K inhibitor and Toll-like receptor agonist induces IFN-gamma + IL-17 + polyfunctional T cells that mediate rejection of murine tumors. Cancer Res 2012; 72: 581–591.
- 137 Conroy H, Marshall NA, Mills KH. TLR ligand suppression or enhancement of Treg cells? A double-edged sword in immunity to tumours. Oncogene 2008; 27: 168–180.
- 138 Conroy H, Galvin KC, Higgins SC, Mills KH. Gene silencing of TGF-beta1 enhances antitumor immunity induced with a dendritic cell vaccine by reducing tumorassociated regulatory T cells. Cancer immunol immunother 2012; 61: 425–431.
 139 Walter S, Weinschenk T, Reinhardt C, Singh-Jasuja H. Single-dose cyclopho-
- 39 Walter S, Weinschenk T, Reinhardt C, Singh-Jasuja H. Single-dose cyclophosphamide synergizes with immune responses to the renal cell cancer vaccine IMA901. Oncolimmunology 2013; 2: e22246.
- 140 Ko HJ, Kim YJ, Kim YS, Chang WS, Ko SY, Chang SY et al. A combination of chemoimmunotherapies can efficiently break self-tolerance and induce antitumor immunity in a tolerogenic murine tumor model. Cancer Res 2007; 67: 7477-7486.
- 141 Goding SR, Wilson KA, Xie Y, Harris KM, Baxi A, Akpinarli A et al. Restoring Immune Function of Tumor-Specific CD4 + T Cells during Recurrence of Melanoma. J Immunol 2013; 190: 4899–4909.
- 142 Wolchok JD, Kluger H, Callahan MK, Postow MA, Rizvi NA, Lesokhin AM et al. Nivolumab plus ipilimumab in Advanced Melanoma. New Engl J Med 2013; 369: 122-133.
- 143 Corsello SM, Barnabel A, Marchetti P, De Vecchis L, Salvatori R, Torino F. Endocrine side effects induced by immune checkpoint inhibitors. J Clin Endocr Metab 2013; 98: 1361–1375.