Sequence and analysis of the citrulline biosynthetic operon argC-F from Bacillus subtilis

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Department of Genetics, Trinity College, Dublin 2, Ireland The citrulline biosynthetic operon argC-F located at 100° on the Bacillus subtilis chromosome contains seven open reading frames which encode all the enzymes required for the biosynthesis of citrulline. The operon is transcribed as a single transcription unit. The second cistron of the operon is homologous to ArgJ (ornithine acetyltransferase) from Bacillus stearothermophilus and Neisseria gonorrhoeae, suggesting that the acetylation of glutamate and the deacetylation of acetylornithine are carried out by a single enzyme in a cyclical pathway. The argF gene is an orthologue of argF from Pseudomonas aeruginosa and a paralogue of arcB from P. aeruginosa and argF/argI from Escherichia coli.

Keywords: Bacillus subtilis, citrulline biosynthetic operon, argC-F

The citrulline biosynthetic operon argC-F is located at approximately 100° on the Bacillus subtilis chromosome between the loci metD and trpS. A schematic diagram of the operon is presented in Fig. 1. The operon contains seven open reading frames (ORFs), argC, argI, argB, argD, car.A, carB and argF, encoding all the enzymes required for the biosynthesis of citrulline. These were identified, with the exception of argC and argF, by their homology with genes from Escherichia coli, Neisseria gonorrhoeae and Bacillus stearothermophilus. This operon is transcribed as a single transcription unit. There are three overlapping ORFs (argB/argD, carA/carB and carB/argF), suggesting that translation of these ORFs is coupled. The sequences of argC and argF were reported previously (Smith et al., 1990; Mountain et al., 1990), and our sequences concur with those reported.

It has been reported that DNA located between the argC and argB cistrons of this B. subtilis operon complements both argA and argE mutants of E. coli (Mountain et al., 1984, 1986). There is, however, only one ORF between the argC and argB cistrons in our sequence. The putative product of this ORF is 64% identical at the amino acid level to ArgJ from B. stearothermophilus and 37% identical to the ArgJ protein of Neisseria gonorrhoeae (Fig. 2, Table 1; Martin & Mulks, 1992; Sakanyan et al., 1993). The ArgJ protein is an ornithine acetyltransferase, an enzyme which uses acetylornithine (an intermediate in the citrul-

The EMBL accession number for the nucleotide sequence reported in this paper is ${\bf Z26919}$.

line biosynthetic pathway) as acetyl donor in the first reaction in the pathway, the acetylation of glutamate. Thus the acetylation of glutamate and the deacetylation of acetylornithine reactions in the biosynthesis of citrulline in *B. subtilis* appear to be carried out in a manner similar to that observed in *B. stearothermophilus* (Sakanyan *et al.*, 1992, 1993) and *N. gonorrhoeae* (Martin & Mulks, 1992), and differently from that found in *E. coli*, where two separate enzymes, ArgA and ArgE, carry out the two reactions (Cunin *et al.*, 1986).

The percentage amino acid identities between the ORFs from this operon and homologous proteins from B. subtilis and other organisms are shown in Table 1. The percentage amino acid identities with homologues from B. stearothermophilus range from 51 % (ArgB) to 64 % (ArgJ), and with homologues from E. coli from 33% (ArgB) to 42 % (CarB). B. subtilis contains two carbamoyl phosphate synthetases, one involved in the biosynthesis of arginine (CarA, CarB, this work) and a second involved in pyrimidine biosynthesis (PyrAA, PyrAB, Quinn et al., 1991). These homologues are 48% (CarA, PyrAA) and 53% (CarB, PyrAB) identical (Table 1). The subunits of the two B. subtilis carbamoyl phosphate synthetases are more similar to each other than either is to the E. coli enzyme, suggesting that the gene duplication event in B. subtilis occurred after the divergence of Gram-negative and Gram-positive bacteria.

The *Pseudomonas aeruginosa* genome contains two genes encoding ArgF-like proteins, *argF* (Itoh *et al.*, 1988) and *arcB* (Bauer *et al.*, 1987), which appear to reflect an ancient

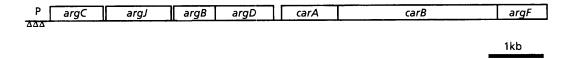


Fig. 1. Schematic representation of the citrulline biosynthetic operon from *B. subtilis*. Each ORF is represented by a box. A gap between boxes indicates that the ORFs do not overlap; a single vertical line indicates that these ORFs do overlap. The operon promoter is represented by P and the putative arginine boxes recognized by the arginine repressor are indicated by triangles (\triangle).

R.	SUBTIL	M-IOLSEDOIVKVT-GDVSSPKGFOAKGVHCGLRYSKKDLGVIISETPAVSAAVYTOSHF
	STEARO	MTITKOTGOVTAVADGTVVTPEGFOAAGVNAGLRYSKNDLGVILCDVPASAAAVYTOSHF
	GONORR	MAVNLTEKTAEOLPDIDGIALYTAOAGVKKPGHTDLTLIAVAAGSTVGAVFTTNRF
14.	GONORR	MAVNLIERIAEQLEDIDGIALIIAQAGVRPGHIDLILIAVAAGSIVGAVFIINRF
		* . ** * ** *
_	attomate	ALL DELUMANAT HUARMI HALLEDIAL TANA AMARAAT MALUMMARAH AAT AT DRIVING
	SUBTIL	QAAPIKVTQDSLKHGPTLKAVIVNSAIANACTGEQGLKDAYTMRESFASQLGIEPELVAV
	STEARO	QAAPLKVTQASLAVEQKLQAVIVNRPCANACTGAQGLKDAYEMRELCAKQFGLALHHVAV
N.	GONORR	CAAPVHIAKSHLFDEDGVRALVINTGNANAGTGAQGRIDALAVCAAAARQIGCKPNQVMP
		*** ** *** ** ** . * *.* . *
В.	SUBTIL	SSTGVIGEHLDMEKIHAGIELLKETPAGSGDFEEAILTTDTVIKQTCYELAIGGK-TV
В.	STEARO	ASTGVIGEYLPMEKIRAGIKQLVPGVTMADAEAFQTAILTTDTVMKRACYQTTIDGK-TV
N.	GONORR	FSTGVILEPLPADKIIAALPKMQPAFWNEAARAIMTTDTVPKAASREGKVGDQHTV
		· **** * * · ** * · · · · **.*** * · · · ·
В.	SUBTIL	TIGGARKGSGMIHPNMATMLGFVTTDAAIEEKALQKALRETTDVSFNQITVDGETSTNDM
В.	STEARO	TVGGAAKGSGMIHPNMATMLAFITTDANVSSPVLHAALRSITDVSFNQITVDGDTSTNDM
N.	GONORR	RATGIAKGSGMIHPNMATMLGFIATDAKVSQPVLQLMTQEIADETFNTITVDGDTSTNDS
		* **************** *
В.	SUBTIL	VLVMANACAENECLTE-DHPDWPVFKKALLLTCEDLAKEIARDGEGATKLIEAQVQGAKN
В.	STEARO	VVVMASGLAGNDELTP-DHPDWENFYEALRKTCEDLAKQIAKDGEGATKLIEVRVRGAKT
N.	GONORR	FVIIATGKNSOSEIDNIADPRYAOLKELLCSLALELAOAIVRDGEGATKFITVRVENAKT
		*
В.	SUBTIL	NLDANVIAKKIVGSNLVKTAVYGTDANWGRIIGAIGHSA-AQVTAEEVEVYLGGQCLFK-
B.	STEARO	DEEAKKIAKOIVGSNLVKTAVYGADANWGRIIGAIGYSD-AEVNPDNVDVAIGPMVMLK-
N.	GONORR	CDEAROAAYAAARSPLVKTAFFASDPNLGKRLAAIGYADVADLDTDLVEMYLDDILVAEH
		.* * * ******. * *
В.	SUBTIL	NNEPQPFSESIAKEYLEGDEITIVIKMAEGDGNGRAWGCDLTYDYIKINASYRT
	STEARO	GSEPOPFSEEEAAAYLOOETVVIEVDLHIGDGVGVAWGCDLTYDYVKINASYRT
	GONORR	GGRAASYTEAOGOAVMSKDEITVRIKLHRGOAAATVYTCDLSHGYVSINADYRS
	- 521-52121	* * ** * ** **

Fig. 2. Alignment of the amino acid sequences of the ArgJ proteins from B. subtilis, (B. subtil) [this work] and N. gonorrhoeae (N. gonorr) [Martin & Mulks, 1992] and B. stearothermophilus (B. stearo) [Sakanyan et al., 1993] using the CLUSTAL V package (Higgins et al., 1992). Exact amino acid matches between all three proteins are indicated by an asterisk (*); conservative amino acid substitutions are indicated by a full stop (.).

Table 1. Percentage identity between the proteins encoded by the citrulline biosynthetic operon argC–F of B. subtilis and homologous proteins from other bacteria

B. subtilis	(F _{op})*	Percentage identity with proteins from:					
ergC–F orotein		Gram-negative bacteria†		B. subtilis		B. stearo- thermophilus	
ArgC	(0.22)	ArgC ^{Ee}	37	_		ArgC‡	59
ArgJ	(0.30)	Arg J ^{Ng}	37	_		ArgJ	64
ArgB	(0.25)	${ m Arg}{ m B}^{ m Ec}$	33	_		ArgB	51
ArgD	(0.30)	$ArgD^{Ee}$	38	_		ArgD‡	62
CarA	(0.23)	CarA Ec	35	PyrAA	48	_	
CarB	(0.25)	CarB ^{Ee}	42	PyrAB	53	-	
ArgF	(0.26)	ArgF ^{Pa}	47	_		_	
		ArcB ^{Pa}	42				
		${ m Arg} { m F}^{ m Ng}$	42				
		$ArgF^{Ec}$	40				
		ArgI ^{Ec}	40				

^{*}Frequency of optimal codon usage.

[†] Ec, Ng and Pa superscripts refer to proteins identified in E. coli, N. gonorrhoeae and P. aeruginosa respectively.

[‡] Comparisons calculated using partial amino acid sequences.

duplication event (the encoded proteins exhibit only 40% identity). The B. subtilis argF product is more similar to the P. aeruginosa argF product (47% identity) than it is to the arcB product (42% identity). Thus B. subtilis argF and P. aeruginosa argF appear to be orthologous genes, while the P. aeruginosa arcB gene is paralogous. The E. coli argF and argI genes (whose products share 87% identity) represent a comparatively recent duplication in the E. coli lineage (Van Vliet et al., 1984) and both appear to be orthologues of P. aeruginosa arcB (with which they share 58% identity) rather than to argF (40% identity). Thus it appears that the B. subtilis argF and E. coli argF/argI are paralogues.

The extent of codon usage bias in the *B. subtilis* genes was estimated by the frequency of optimal codons (F_{op} , Table 1) as defined by Sharp *et al.* (1990). The values obtained are typical of moderately/lowly expressed genes in *B. subtilis*; the *E. coli* homologues have similarly moderately biased codon usage (data not shown).

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