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Efficient Synthesis of Glycoporphyrins via Microwave Mediated 'Click' Reactions

Oliver B. Locos,[a] Claudia C. Heindl,[a] Ariadna Corral,[a] Mathias O. Senge,[b] and Eoin M. Scanlan,*[a]

Keywords: Glycoporphyrin / Click / Microwave/ Carbohydrate

The Cu(I)-catalysed Huisgen cycloaddition 'click' reaction has been applied to the synthesis of a range of triazole linked glycoporphyrins. The 'click' reaction under microwave heating conditions has been shown to provide a general and robust methodology for the synthesis of mono-, di-, tri- and tetra-modified glycoporphyrins. A sequential 'double-click' process was employed to access a new class of bis-modified 5,10-diglycoporphyrins displaying heterogeneous carbohydrates.

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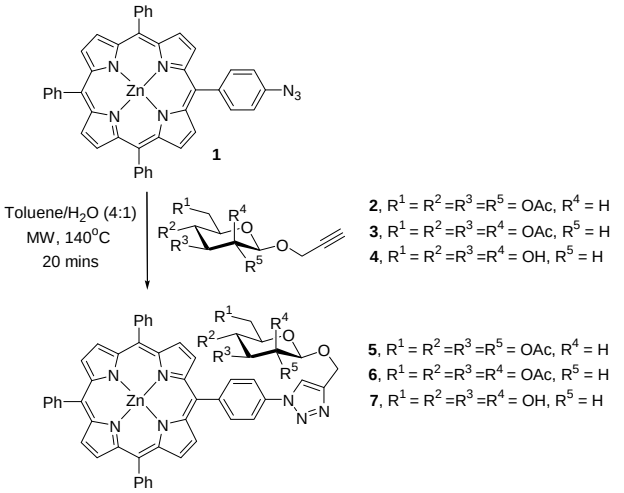
Introduction

Glycoporphyrins offer fascinating prospects for medicinal chemistry and glycobiology.[1a-1d] Carbohydrates not only improve the solubility of porphyrins in an aqueous environment but also offer improved targeting of porphyrin therapeutics.[2] For photodynamic therapy (PDT),[3] carbohydrates can bind to tumour associated lectins displayed on the surface of cancer cells therefore offering the potential for enhanced efficiency and improved selectivity in cancer treatment. Since lectin-carbohydrate interactions are relatively weak it would be advantageous to have access to a PDT photosensitiser with a defined cluster displaying more than one carbohydrate unit. Even more advantageous would be a defined system that displays more than one '*type*' of carbohydrate. The synthesis of selectively modified, heterogeneous glycoporphyrins has not previously been reported. To date, the synthesis of glycoporphyrins has relied predominantly on the condensation reaction between a glycosylated aldehyde and a glycosylated aldehyde and a pyrrole.[4] This reaction is often low yielding and is unsuitable for systems where small quantities of a carbohydrate are available. A more general strategy involves the introduction of carbohydrates at specific sites predefined by a regioselective chemical modification.[5] A number of methodologies for the functionalisation of porphyrins with carbohydrates have been investigated including the use of Sonogashira [6] and olefin metathesis [7] cross coupling.

Results and Discussion

For this study we chose to investigate if the Cu(I)-catalysed 1,3-dipolar 'click' reaction [8] could be employed as a robust methodology to access a highly defined, multifunctionalised glycoporphyrin system.

Click chemistry has previously been applied to the functionalisation of porphyrins,[9] but to date there is only one literature example describing a click reaction involving a mono-carbohydrate modification of a chlorin[10]. 5,10,15,20-Tetraphenylporphyrin was chosen as a common starting material for all modifications. This symmetrical porphyrin can be readily accessed in good yield via a condensation reaction of benzaldehyde and pyrrole. Porphyrin **(1)**, displaying a single azide was prepared according to the literature procedure.[11] The coupling reaction with commercially available β- propargyl glucose **(2)** (Scheme 1) was investigated under both conventional and microwave mediated heating. The use of MW heating conditions reduced the reaction time from 3 days to 20 mins. A range of reaction conditions were screened. The results of this initial study are outlined in Table 1.



Scheme 1. Synthesis of mono-glycoporphyrin by microwave mediated 'click' reaction.

It was found that copper chloride in toluene/water (4:1) at 120°C for 20 mins furnished the desired mono-glycoporphyrin (**5**) in excellent yield (93%). THF/water (4:1) also gave a very good yield of 81% at a lower temperature but longer times (40 mins) were required for complete consumption of the starting material. It was found that zinc porphyrin was required for 'click' conditions in order to avoid complex formation with the copper catalyst.

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Table 1. Optimisation of Conditions for Cu catalysed click reaction.

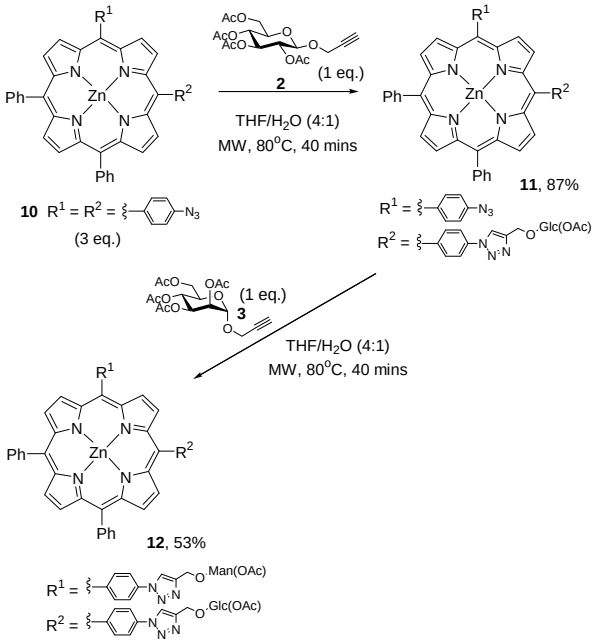
Entry	Sugar	Catalyst[a]	Solvent[b]	Temp[c]	Yield(%)[d]
1	**2**	CuCl	Toluene	90-140	47
2	**2**	CuCl	Toluene	140	85
3	**2**	CuCl	Toluene/H_2O	140	93
4	**2**	CuBr	Toluene/H_2O	140	78
5	**2**	CuI	Toluene/H_2O	140	44
6	**2**	$CuSO_4$. Na^+ ascorbate	Toluene/H_2O	140	89
7	**2**	CuCl	DMSO/H_2O	140	67
8	**2**	CuCl	DMSO/H_2O	160	63
9	**2**	CuCl	DMSO/H_2O	160	52
10	**2**	CuCl	THF/H_2O	80	81
11	**2**	CuCl	DMF/H_2O	140	74

[a] Catalyst loading was 30% [b] Where two solvents were used ratio was (4:1) [c] Temperatures were obtained under microwave heating for 20 mins [d] isolated yield.

The reaction was repeated with α-propargyl mannose, prepared according to the literature procedure,[12] and the desired mannosylated glycoporphyrin (**6**) was isolated in 91% yield. A fully deprotected mannosyl residue (**4**) was also introduced via the 'click' reaction to furnish (**7**) in good yield 61%. This reaction highlights the potential for the introduction of unprotected oligosaccharides isolated from natural sources using this methodology. It also means that deprotection reactions, which can be low yielding for oligosaccharides can be carried out prior to functionalisation of the porphyrin. The fact that the glycoporphyrin was easily visible on the chromatography column facilitated the rapid and simple purification of the glycoconjugate displaying the fully deprotected sugar.

The next step was to investigate if the methodology could be applied to diazidoporphyrins. Nitration of 5,10,15,20-tetraphenylporphyrin furnished the 5,10 and 5,15 regioisomers in a 2:1 ratio and offers an alterantive entry to the 5,10-disubstitution pattern in porphyrins.[13] These compounds were easily separated by column chromatography. They were both subsequently converted to the corresponding azidophenylporphyrins after reduction of the nitro groups were achieved. The click reaction using two equivalents of the propargyl glucose worked efficently and the 5,10-diglycoporphyrin (**8**) was isolated in 80% yield and the 5,15- compound (**9**) in 75% yield. This ability to control the relative position of carbohydrate side chains on the porphyrin may have very important consequences for lectin binding.

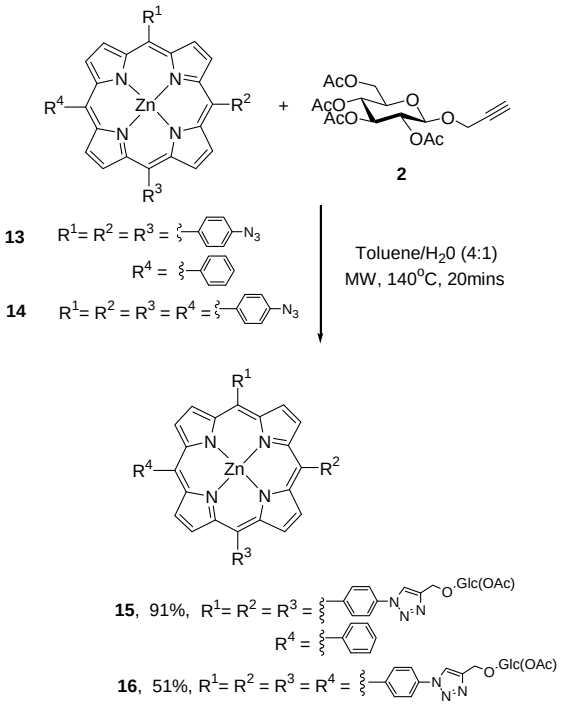
Efficient, regioselective synthesis of bis-modified glycoporphyrins has not previously been described. It was determined that a sequential 'double click' reaction where one carbohydrate moiety was introduced followed by addition of a second onto the 'latent' azide could be achieved. Reaction of propargyl glucoside (**2**) with three equivalents of diazaporphyrin (**10**) furnished the glycoporphyrin (**11**) as the sole product isolated in very good yield 87%. The propargyl mannoside (**3**) was then introduced via a second sequential click reaction to furnish the bis-modified glycoporphyrin (**10**) in 53% yield.



Scheme 2. Synthesis of 5,10-bis-modified heterogeneous glycoporphyrin via stepwise 'double click' reaction.

In order to avoid reduction of the 'latent' azide and allow the stepwise process to occur, the amount of copper catalyst had to be carefully controlled. It was found that 30% catalyst loading was optimum for each step of the 'double click' reaction. The excess porphyrin required for the first click reaction was recovered almost quantitatively by colum chromatography making this an efficient process. The scope of this reaction is not just limited to carbohydrates and studies are currently underway to prepare mixed porphyrins functionalised with carbohydrates, lipids and amino acids.

Following the successful preparation of the bis-modified product, the tri- (**13**) and tetra-azido (**14**) porphyrins were synthesised. The conditions employed for the mono- and dimodification were repeated, using three and four equivalents of the propargyl glycoside, respectively. The tri- (**15**) and tetraglycosylated porphyrins (**16**) were both isolated in good to moderate yields, respectively. This demonstrated that the same general conditions could be used to prepare the mono-, di-, tri- and tetraglycoporphyrins. This suggests that the methodology could even be applied to the synthesis of dendritic glycoporphyrins displaying a large number of carbohydrate side chains. It should be noted that while the conditions for (**15**) and (**16**) are unoptimised, the yield of (**16**) is considerably lower, most likely due to solubility issues with the azaporphyrin (**14**).



Scheme 3: Synthesis of tri- and tetra- substituted glycoporphyrins.

Conclusions

In conclusion the 'click' reaction under microwave heating conditions represents an efficient and robust methodology for the synthesis of defined glycoporphyrins amenable for further elaborations. General conditions were determined for the introduction of homogeneous carbohydrates and conditions for the stepwise bis-modification with heterogeneous carbohydrates were also determined. The methodology has been optimised to allow functionalistion of the tetrapyrrole core with both protected and fully deprotected carbohydrates. Studies are currently underway to explore additional, orthogonal modification strategies including cross metathesis and organo metallic coupling reactions for the construction of highly diverse glycoporphyrins and analogs. Carbohydrate sequences suitable for binding tumour associated lectins will be introduced using this methodolgy. Biological assays to study the therapeutic applications of these systems are also under investigation.

Supporting Information (see footnote on the first page of this article): General experimental procedures and

Acknowledgments

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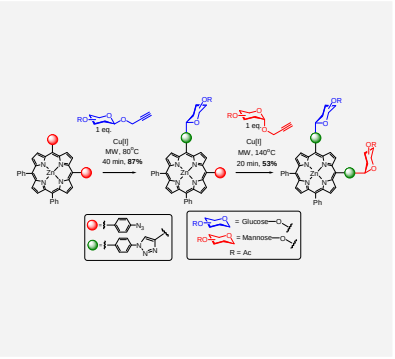
Entry for the Table of Contents

Layout 1:

Carbohydrate Chemistry

A novel, two-step, sequential ‘double-click’ process allows the preparation of heterogeneous glycoporphyrins in good yields. Microwave heating was used to accelerate reaction times and improve yields for the Cu catalysed cycloaddition reaction. The latent azide was determined to be stable under these conditions.



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Efficient Synthesis of Glycoporphyrins via Microwave Mediated Click reactions

Keywords: Glycoporphyrin, Click, Microwave, Carbohydrate.

Layout 2:

((Key Topic))

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Keywords: ((Keyword 1 / Keyword 2 / Keyword 3 / Keyword 4 / Keyword 5))

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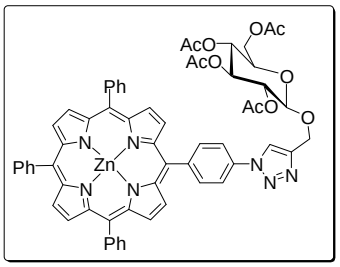
General Information

Proton Nuclear Magnetic Resonance spectra were recorded on a 600 or 400 MHz spectrometer in $CDCl_3$ ($CDCl_3$/d5-pyridine or neat d5-pyridine for zinc(II) porphyrins) referenced relative to residual $CHCl_3$ (δ = 7.26 ppm). Chemical shifts are reported in ppm and coupling constants in Hertz. Carbon NMR spectra were recorded on the same instrument (150 or 100 MHz) with total proton decoupling. Flash chromatography was carried out using silica gel, particle size 0.04-0.063 mm. TLC analysis was performed on precoated 60F254 slides, and visualised by UV irradiation, $KMnO_4$ or Molybdenum staining. Mass spectrometry analysis was performed with a Q-Tof Premier Waters Maldi-quadrupole time-of-flight (Q-Tof) mass spectrometer equipped with Z-spray electrospray ionization (ESI) and matrix assisted laser desorption ionisation (MALDI) sources. All azidophenylporphyrins (mono, 5,10-bis, 5,15-bis tris- and tetra) were synthesised according to the general procedure below.

General Procedure for the azidation of aminophenyl porphyrins: To a stirred solution of the mono, di, tri or tetra aminophenyl porphyrin (100 mg) in TFA (1 mL) at 0 ^oC a solution of $NaNO_2$ in H_2O was added dropwise (2 eq. of $NaNO_2$ for each amine). The mixture was stirred at 0 ^oC for 10 mins. After this time a solution of NaN_3 in H_2O (4 eq. of NaN_3 for each amine) was added dropwise. The reaction mixture was stirred at 0 ^oC for 45 min and monitored by TLC (CH_2Cl_2 /*n*-hexane). After full conversion, as determined by TLC the solution was diluted with H_2O and extracted with CH_2Cl_2. The organic layer was washed with aqueous $NaHCO_3$ until the organic layer turned from green to purple. The organic layer was dried over $MgSO_4$, filtered and concentrated to furnish the azidoporphyrin.

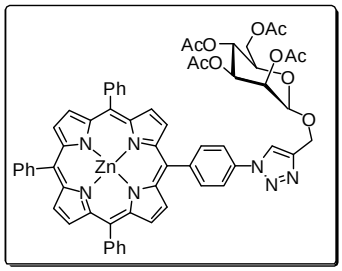
General Procedure for Metallation of Azidophenyl Porphyrin: To a stirred solution of the azidoporphyrin (100 mg) in chloroform (30 mL) at rt was added a solution of $Zn(OAc)_2$ (100 mg, 3.6 eq) in MeOH (1 mL). The mixture was stirred at rt for 1 h during which time the colour of the reaction mixture changed from light purple to dark purple. The reaction was monitored by TLC (CH_2Cl_2 /*n*-hexane). After full conversion, as determined by TLC, the reaction mixture was evaporated to dryness and the resulting purple solid was redissolved in CH_2Cl_2 and filtered through a plug of silica using CH_2Cl_2 as eluent. The solvent was evaporated to furnish the zinc azidoporphyrin as a shiny dark purple solid.

5-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-10,15,20-triphenylporphyrinatozinc(II) (5)



18 mg of porphyrin **1** (0.025 mmol), 1 mg CuCl (0.007 mmol), 12 mg of propargyl glucose **2** (0.030 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140°C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:4). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:10) and the desired glycoporphyrin **5** was isolated as a bright purple shiny solid (25 mg, 93%). ^{1}H NMR (600 MHz, $CDCl_3$): 8.93 (2H, d, *J* = 4.6 Hz, 2,8-β-H), 8.91 (4H, s, 12,13,17,18-β-H), 8.88 (2H, d, *J* = 4.6 Hz, 3,7-β-H), 8.37 (2H, d, *J* = 8.3 Hz, o-Ph-H), 8.23 (6H, m, o-Ph-H), 8.23 (1H, 5-triazole-H), 8.08 (2H, d, *J* = 8.3, m-Ph-H), 7.76 (9H, m, m,p-Ph-H), 5.30 (1H, t, *J* = 9.4, H-3), 5.19 (1H, t, *J* = 9.7, H-4), 5.15 (1H, dd, *J* = 7.9, 7.8 Hz, H-2), 5.13 (1H, d, *J* = 12.6 Hz, OC*H*H), 5.03 (1H, d, *J* = 12.6 Hz, OCH*H*), 4.82 (1H, d, *J* = 7.9 Hz, H-1), 4.35 (1H, dd, *J* = 4.7, *J* = 12.4 Hz, H-6), 4.22 (1H, dd, *J* = 2.0, 12.4 Hz, H-6'), 3.80 (1H, ddd, *J* = 9.7, 4.6, 2.0 Hz, H-5), 2.17 (3H, s, CH_3), 2.12 (3H, s, CH_3), 2.09 (3H, s, CH_3), 2.07 (3H, s, CH_3); ^{13}C NMR (150 MHz, $CDCl_3$): 171.1, 170.5, 170.0, 169.3 (C=O), 150.0, 149.9, 149.4, 148.2, 148.0, 147.8, 147.6, 145.0, 144.4, 143.2, 135.9, 135.7 (Ar-C) 135.4, 134.4 (o-Ph-C), 131.8, 131.6, 131.6, 130.8 (β-C), 127.02, 126.1 (m-Ph-C), 121.1 (5-triazole-C), 120.9, 120.7, 118.2, 118.0 (m-Ph-C), 99.9 (C-1), 72.7 (C-3), 71.9 (C-5), 71.2 (C-2), 68.2 (C-4), 62.9 (CH_2), 61.7 (C-6), 20.7, 20.6, 20.6, 20.4 (4 x CH_3); IR: 2923 (m, C-H stretch), 2853 (m, C-H stretch), 1744 (s, C=O stretch), MS(MADLI-TOF): Calculated for $C_{61}H_{49}N_7O_{10}Zn$ M^+ 1103.2832; Found 1103.2832.

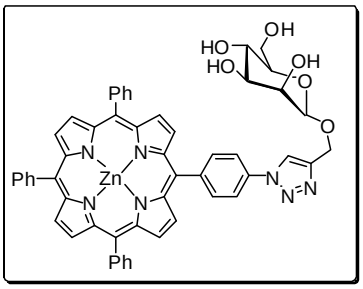
5-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-mannopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-10,15,20-triphenylporphyrinatozinc(II) (6)



20 mg of porphyrin **1** (0.028 mmol), 1 mg CuCl (0.007 mmol), 12 mg of propargyl mannose **3** (0.030 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140°C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:4). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:10) and the desired glycoporphyrin was isolated as a bright purple shiny solid (26 mg, 86%). ^{1}H NMR (600 MHz,

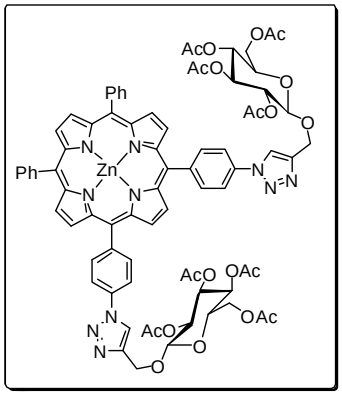
$CDCl_3$): 9.20 (2H, d, *J* = 4.5 Hz, 2,8-β-H), 8.99 (4H, s, 12,13,17,18-β-H), 8.97 (2H, d, *J* = 4.5 Hz, 3,7-β-H), 8.42 (2H, d, *J* = 7.7 Hz, o-Ph-H), 8.34 (1H, 5-triazole-H), 8.26 (6H, m, o-Ph-H), 8.16 (2H, d, *J* = 6.9, m-Ph-H), 7.79 (9H, m, m,p-Ph-H), 5.46-5.33 (3H, m, H-2, H-3, H-4), 5.11 (1H, s, H-1), 5.00 (1H, d, *J* = 12.0 Hz, OC*H*H), 4.83 (1H, d, *J* = 12.0, OCH*H*), 4.40 (1H, dd, *J* = 12.2, 5.0 Hz, H-6), 4.30 (1H, m, H-6'), 4.20 (1H, m, H-5), 2.22 (3H, s, CH_3), 2.20 (3H, s, CH_3), 2.10 (3H, s, CH_3), 2.04 (3H, s, CH_3); ^{13}C NMR (150 MHz, $CDCl_3$): 170.8, 170.1, 170.0, 169.7 (C=O), 150.5, 150.4, 150.3, 149.8, 144.6, 144.0, 142.7, 136.3, 135.4, 134.5, 132.4, 132.2, 132.2, 131.4, 127.6, 126.6, 121.6, 121.4 (5-triazole-C), 118.6, 97.1 (C-1), 69.6 (C-2), 69.1 (C-3 or C-4), 68.9 (C-5), 66.1 (C-3 or C-4), 62.5 (C-6), 61.2 (OCH_2), 20.9, 20.8, 20.7, 20.7 (4 x CH_3); IR: 2923 (m, C-H stretch), 2855 (m, C-H stretch), 1743 (s, C=O stretch), MS (ESI -TOF): Calculated for $C_{61}H_{50}N_7O_{10}Zn$ $(M+H)^+$ 1104.2911 ; Found 1104.2925.

5-((4-β-D-mannopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-10,15,20-triphenylporphyrinatozinc(II) (7)



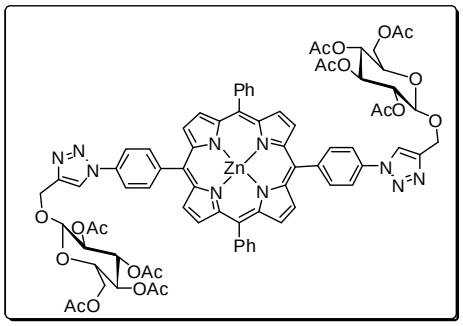
20 mg of porphyrin **1** (0.028 mmol), 1 mg CuCl (0.007 mmol), 6.2 mg of deprotected propargyl mannose **4** (0.028 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140^{o}C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:4). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:10) and the desired glycoporphyrin was isolated as a bright purple shiny solid (16 mg, 61%). ^{1}H NMR (600 MHz, pyridine-d_5): 9.20 (3H, q, *J* = 4.4 Hz, β-H), 9.15 (3H, s, β-H), 9.02 (2H, s, β-H), 8.47 (2H, d, *J* = 8.2 Hz, o-Ph-H), 8.41 (6H, m, o-Ph-H), 8.39 (1H, 5-triazole-H), 8.33 (2H, d, *J* = 8.2, m-Ph-H), 7.78 (9H, m, m,p-Ph-H), 5.74 (1H, s, H-1), 5.40 (1H, d, *J* = 5.4 Hz, OC*H*H), 5.20 (1H, d, *J* = 5.3 Hz, OCH*H*), 4.74-4.66 (4H, m, H-2, H-3, H-4, H-6), 4.58 (1H, m, H-5), 4.49 (1H, m, H-6'); ^{13}C NMR (150 MHz, pyridine-d_5): 150.4, 150.4, 150.1, 149.1, 145.8, 143.7, 143.6, 143.4, 143.4, 136.5, 135.6, 135.5, 134.8, 134.7, 132.3, 132.1, 132.0, 131.7, 127.4, 126.6, 126.6, 122.3, 121.3, 121.3, 121.2, 119.2, 118.1, 100.9 (C-1), 75.6 (C-5), 72.8, 71.8, 68.9, (C-2, C-3, C-4), 64.1 (C-6), 61.6 (OCH_2); MS (MALDI-TOF): Calculated for $C_{53}H_{41}N_7O_6Zn$ M^+ 935.2410; Found 935.2446.

5,10-Bis-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-15,20-diphenylporphyrinatozinc(II) (8)



12 mg of porphyrin **10** (0.015 mmol), 1 mg CuCl (0.007 mmol), 12 mg of propargyl glucose **2** (0.030 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140°C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:2). The solvent was removed in vacuo and the residue was purified by column chromatography on 20 mL silica using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:2) and the desired glycoporphyrin was isolated as a bright purple shiny solid (21 mg, 87%). ^{1}H NMR ($CDCl_3$): 8.93 (2H, d, *J* = 4.6 Hz, 2,13-β-H), 8.92 (2H, s, 17,18-β-H) 8.90 (2H, s, 5,6-β-H), 8.89 (2H, d, *J* = 4.6, 3,12-β-H) 8.39 (4H, d, *J* = 8.1, o-Ph-H), 8.31 (2H, s, 5-triazole-H), 8.22 (4H, d, *J* = 7.6, o-Ph-H), 8.13 (2H, d, *J* = 8.1, m-Ph-H), 7.77 (6H, m, m,p-Ph-H), 5.31 (2H, t, *J* = 9.4, H-3), 5.19 (2H, t, *J* = 9.7, H-4), 5.18 (2H, d, *J* =12.6, OCH_2), 5.13 (2H, dd, *J* = 8.3, *J* = 7.9, H-2), 4.86 (2H, d, *J* = 12.6, CH_2), 4.82 (1H, d, *J* = 7.9, H-1), 4.35 (2H, dd, *J* = 4.8, *J* = 12.5, H-6), 4.22 (1H, dd, *J* = 2.3, *J* = 12.5, H-6'), 3.85 (1H, ddd, *J* = 9.5, 4.8, 2.3, H-5), 2.16 (3H, s, CH_3), 2.11 (3H, s, CH_3), 2.08 (3H, s, CH_3), 2.05 (3H, s, CH_3); ^{13}C NMR (150 MHz, $CDCl_3$): 170.6, 170.2, 169.5, 169.4 (C=O), 149.6, 149.3, 149.0, 135.7, 135.6, 135.5, 135.2, 134.6, 131.6, 127.2, 126.4, 123.4, 123.2, 122.9, 121.4, 118.4, 100.1 (C-1), 72.8, 72.0, 71.3, 68.4, 63.1, 63.1, 61.9, 30.9, 29.7, 20.8, 20.6; MS(MALDI-TOF): IR: 2922 (m, C-H stretch), 2853 (m, C-H stretch), 1744 (s, C=O stretch), Calculated for $C_{78}H_{70}N_{10}O_{20}Zn$ M^+ 1530.4059; Found 1530.4037.

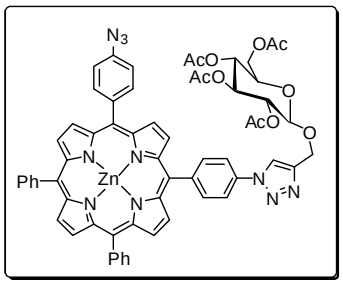
5,15-Bis((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-10,20-diphenylporphyrinatozinc(II) (9)



12 mg of porphyrin **8a** (0.015 mmol), 0.5 mg CuCl (0.004 mmol), 12 mg of propargyl glucose **2** (0.030 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140°C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:4). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:2) and the desired glycoporphyrin was isolated as a bright purple shiny solid (18 mg, 75%). ^{1}H NMR (400 MHz, $CDCl_3$):

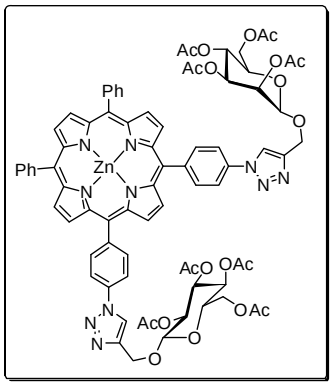
8.94 (4H, d, *J* = 4.6, 3,7,13,17-β-H), 8.89 (4H, d, *J* = 4.6, 2,8,12,18-β-H) 8.38 (4H, d, *J* = 8.3, o-Ph-H), 8.31 (2H, s, 5-triazole-H), 8.23 (4H, dd, *J* =7.3, *J* =1.8, o-Ph-H), 8.12 (4H, d, *J* = 8.3, m-Ph-H), 7.77 (6H, m, m,p-Ph-H), 5.30 (2H, t, *J* = 9.4, H-3), 5.19 (2H, t, *J* = 9.4, H-4), 5.18 (2H, d, *J* = 12.5, OC*H*H), 5.15 (2H, dd, *J* = 9.4, 7.9, H-2), 5.05 (2H, d, *J* = 12.5, OCH*H*), 4.86 (1H, d, *J* = 7.9, H-1), 4.37 (2H, dd, *J* = 4.4, *J* = 12.3, H-6), 4.22 (1H, dd, *J* = 2.3, *J* = 12.3, H-6'), 3.85 (1H, ddd, *J* = 9.6, 4.4, 2.3, H-5), 2.16 (3H, s, CH_3), 2.11 (3H, s, CH_3), 2.08 (3H, s, CH_3), 2.05 (3H, s, CH_3); ^{13}C NMR (100 MHz, $CDCl_3$): 170.5, 170.1, 169.3, 169.3 (C=O), 150.1, 149.6, 149.1, 148.9, 148.7, 145.1, 144.3, 143.1, 136.1, 135.7 (o-Ph-C), 135.6, 135.4, 135.3, 134.4 (o-Ph-C), 132.0 (β-C), 131.1 (β-C), 127.2 (m-Ph-C), 126.2 (p-Ph-C), 123.2, 123.0, 122.9, 121.2 (5-triazole-C), 121.0, 118.6, 118.3 (m-Ph-C), 100.0 (C-1), 72.7 (C-3), 72.0 (C-5), 71.2 (C-2), 68.3 (C-4), 63.0 (CH_2), 61.8 (C-6), 20.7, 20.6 (2 x CH_3); IR: 2925 (m, C-H stretch), 2851 (m, C-H stretch), 1741 (s, C=O stretch), MS(MALDI-TOF): Calculated for $C_{78}H_{70}N_{10}O_{20}NaZn$ $(M+Na)^+$ 1553.3957; Found 1553.3954.

5-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-10-(4-azidophenyl)-15,20-diphenylporphyrinatozinc(II) (11)



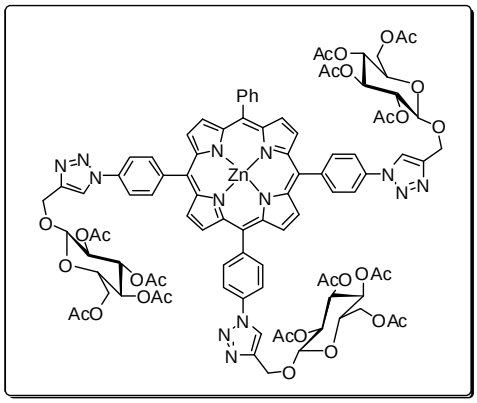
30 mg of porphyrin **8** (0.039 mmol), 0.5 mg CuCl (0.004 mmol), 5 mg of propargyl glucose **2** (0.013 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL THF/H_2O (4:1). The tube was then sealed and heated to 80°C in a microwave reactor for 20 mins. TLC analyses after the 40 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:4). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:10) and the desired glycoporphyrin was isolated as a bright purple shiny solid (25 mg, 80%). ^{1}H NMR (600 MHz, $CDCl_3$): 8.90 (8H, br m, β-H), 8.39 (2H, d, *J* = 6.7, o-Ph-H), 8.30 (1H, s, 5-triazole-H), 8.22 (6H, br m, o-Ph-H), 8.12 (2H, d, *J* = 6.7, m-Ph-H), 7.76 (6H, m, m,p-Ph-H), 7.73 (2H, d, *J* = 6.7, p-Ph-H), 5.31 (1H, br t, *J* = 9.2, H-3), 5.19 (1H, br t, *J* = 9.2, H-4), 5.17 (1H, br d, *J* = 12.3, OC*H*H), 5.15 (1H, br t, *J* = 9.0, H-2), 5.06 (1H, br d, *J* = 12.3, OCH*H*), 4.86 (1H, br d, *J* = 7.6, H-1), 4.37 (1H, dd, *J* = 3.4, *J* = 12.0, H-6), 4.26 (1H, br dd, *J* = 1.8, *J* = 12.0, H-6'), 3.85 (1H, br ddd, *J* = 9.2, 3.4, 1.8, H-5), 2.17 (3H, s, CH_3), 2.12 (3H, s, CH_3), 2.09 (3H, s, CH_3), 2.06 (3H, s, CH_3); ^{13}C NMR (150 MHz, DEPT); 135.7 (o-Ph-C), 135.5 (o-Ph-C), 134.4 (o-Ph-C), 131.9 (β-C), 131.0 (β-C), 127.3 (m-Ph-C), 126.3 (p-Ph-C), 121.1 (5-triazole-C), 118.5 (m-Ph-C), 117.1 (p-Ph-C), 100.2 (C-1), 72.8 (C-3), 72.2 (C-5), 71.3 (C-2), 68.3 (C-4), 63.0 (CH_2), 61.9 (C-6), 20.7 (CH_3), 20.5 (CH_3); IR: 1745 (s, C=O stretch), MS(MALDI-TOF): Calculated for $C_{61}H_{49}N_{10}O_{10}Zn$ $(M+H)^+$ 1145.2925; Found 1145.2925.

5-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-10-((4-β-D-mannopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)15,20-diphenylporphyrinatozinc(II) (12)



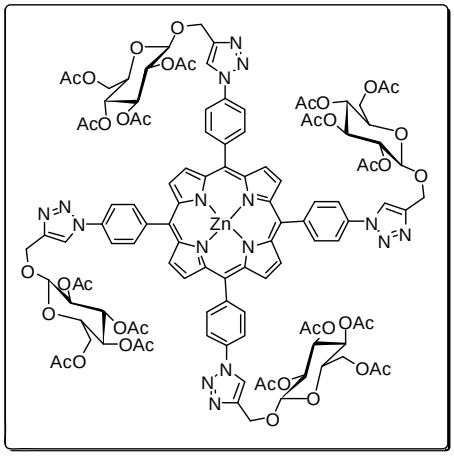
18 mg of porphyrin **9** (0.016 mmol), 0.5 mg CuCl (0.005 mmol), 6.1 mg of propargyl mannose **3** (0.016 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140^{o}C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both, EtOAc/CH_2Cl_2 (1:4) and EtOAc/CH_2Cl_2 (2:3). The solvent was removed in vacuo and the residue was purified by column chromatography using EtOAc/CH_2Cl_2 (1:4) as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (2:3) and the desired glycoporphyrin was isolated as a bright purple shiny solid (8 mg, 33%). ^{1}H NMR (600 MHz, $CDCl_3$): 9.02 (2H, d, *J* = 4.3 Hz, β-H), 9.00 (4H, s, β-H), 8.96 (2H, d, *J* = 4.3 Hz, β-H), 8.42 (4H, d, *J* = 9.4 Hz, o-Ph-H), 8.29 (1H, 5-triazole-H), 8.26 (4H, d, *J* = 6.5 Hz, o-Ph-H), 8.17 (1H, 5-triazole-H), 8.07 (4H, d, *J* = 7.3 Hz, m-Ph-H), 7.79 (6H, m, m,p-Ph-H), 5.44 (1H, m, H-4-Man), 5.38 (1H, m, H-3-Man), 5.35 (1H, m, H-2-Man), 5.27 (1H, t, *J* = 9.4 Hz, H-3-Glc), 5.15 (1H, t, *J* = 9.7 Hz, H-4-Glc), 5.08 (2H, m, H-2-Glc, H-1-Man), 4.87 (2H, m, 2 x OC*H*H), 4.72 (3H, m, H-1-Glc, 2 x OCH*H*), 4.37 (1H, m, H-6), 4.33 (1H, m, H-6), 4.21 (1H, m, H-6), 4.15 (2H, m, H-6', H-5-Man), 3.80 (1H, m, H-5-Glc), 2.21, 2.18, 2.13, 2.09, 2.07, 2.07, 2.04, 2.03 (8 x CH_3); ^{13}C NMR (150 MHz, $CDCl_3$); 170.7, 170.6, 170.2, 170.1, 169.9, 169.7, 169.4, 169.4 (C=O), 150.5, 150.4, 150.0, 149.8, 145.0, 144.4, 143.9, 142.7, 136.3, 135.5, 134.5, 132.6, 132.3, 131.7, 131.5, 127.6, 126.6, 121.8 (Ar-C), 119.0, 118.8, 118.6 (m-Ph-C), 100.2 (C-1-Glc), 97.0 (C-1-Man), 72.7 (C-3-Glc), 72.0 (C-5-Glc), 71.3 (C-2-Glc), 69.5 (C-2-Man), 69.0 (C-4-Man), 68.9 (C-5-Man), 68.3 (C-4-Glc), 66.1 (C-3-Man), 62.8 (CH_2), 62.5, 61.9 (C-6-Man, C-6-Glc), 61.0 (CH_2), 20.9, 20.8, 20.7, 20.7, 20.7, 20.7, 20.6, 20.6 (8 x CH_3); IR: 2922 (m, C-H stretch), 2853 (m, C-H stretch), 1741 (s, C=O stretch); MS (MALDI-TOF): Calculated for $C_{78}H_{70}N_{10}O_{20}Zn$ M^+ 1530.4059; Found 1530.4080.

5,10,15-Tri-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-20-phenylporphyrinatozinc(II) (15)



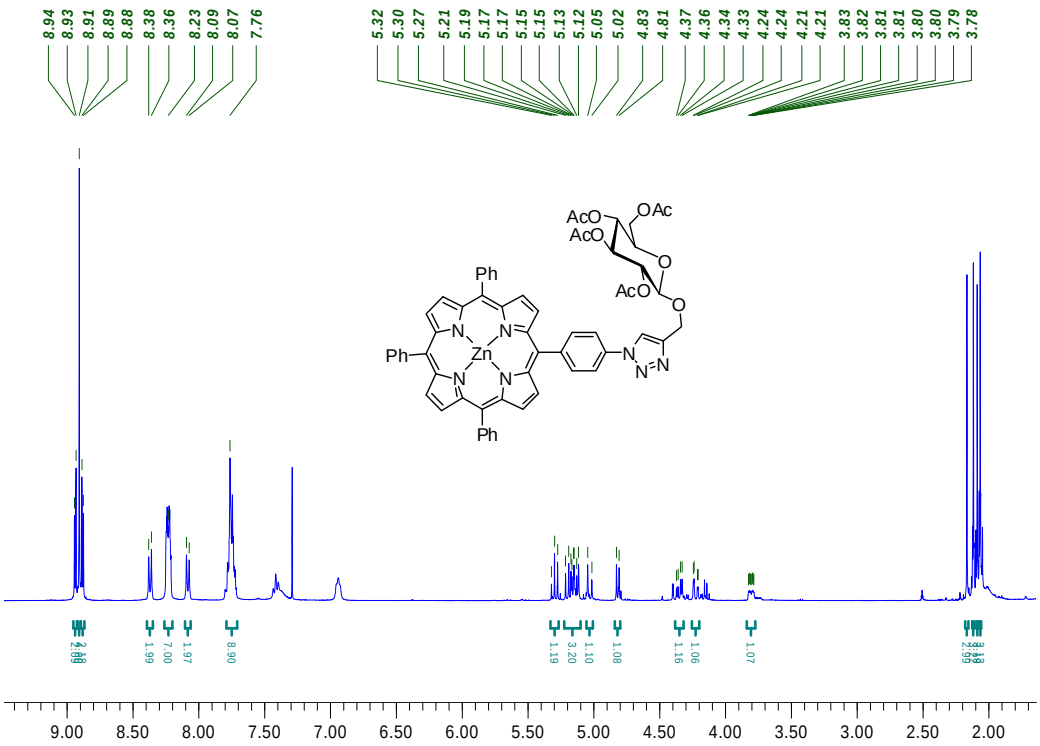
12 mg of porphyrin **11** (0.015 mmol), 0.5 mg CuCl (0.004 mmol), 36 mg of glucose **2** (0.090 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140°C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (1:2). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (1:1) and the desired glycoporphyrin was isolated as a bright purple shiny solid (27 mg, 91%).^{1}H NMR (600 MHz, $CDCl_3$): 8.96 (2H, d, *J* = 4.5, 2,18-β-H), 8.94 (4H, s, 4,5,12,13- β-H), 8.90 (2H, d, *J* = 4.5, 3,17-β-H), 8.39 (4H, d, *J* = 7.1, o-Ph-H), 8.38 (2H, d, *J* = 7.4, o-Ph-H), 8.28 (3H, s, 5-triazole-H), 8.23 (2H, *J* = 6.8, o-Ph-H), 8.13 (2H, d, *J* = 7.4, m-Ph-H), 8.11 (4H, d, *J* = 7.1, m-Ph-H), 7.78 (3H, m, m,p-Ph-H), 5.30 (3H, t, *J* = 9.4, H-3), 5.18 (3H, t, *J* = 9.4, H-4), 5.15 (3H, dd, *J* = 9.4, 7.9, H-2), 5.14 (3H, d, *J* = 12.5, OCH_2), 5.03 (3H, d, *J* = 12.5, OCH_2), 4.84 (3H, d, *J* = 7.9, H-1), 4.35 (3H, dd, *J* = 4.7, *J* = 12.4, H-6), 4.24 (3H, dd, *J* = 2.2, *J* = 12.4, H-6'), 3.83 (3H, ddd, *J* = 9.4, 4.4, 2.2, H-5), 2.16 (9H, s, CH_3), 2.11 (9H, s, CH_3), 2.08 (9H, s, CH_3), 2.06 (9H, s, CH_3);^{13}C NMR (150 MHz, DEPT, $CDCl_3$): 136.2 (o-Ph-C), 134.4 (o-Ph-C), 132.2 (β-C), 131.6 (β-C), 131.3 (β-C), 127.3 (m-Ph-C), 126.3 (p-Ph-C), 121.6 (5-triazole-C), 118.9 (m-Ph-C), 100.7 (C-1), 72.8 (C-3), 72.1 (C-5), 71.4 (C-2), 68.2 (C-4), 62.9 (CH_2), 61.9 (C-6), 20.7 (CH_3), 20.5 (CH_3); IR: 2923 (m, C-H stretch), 2852 (m, C-H stretch), 1742 (s, C=O stretch), MS(MALDI-TOF): Calculated for $C_{95}H_{91}N_{13}O_{30}Zn$ M^+ 1957.5286; Found 1957.5430.

5,10,15,20-Tetra-((4-[(2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranosyl)oxymethyl]-1-*H*-1,2,3-triazol-1-yl])phenyl)-porphyrinatozinc(II) (16)

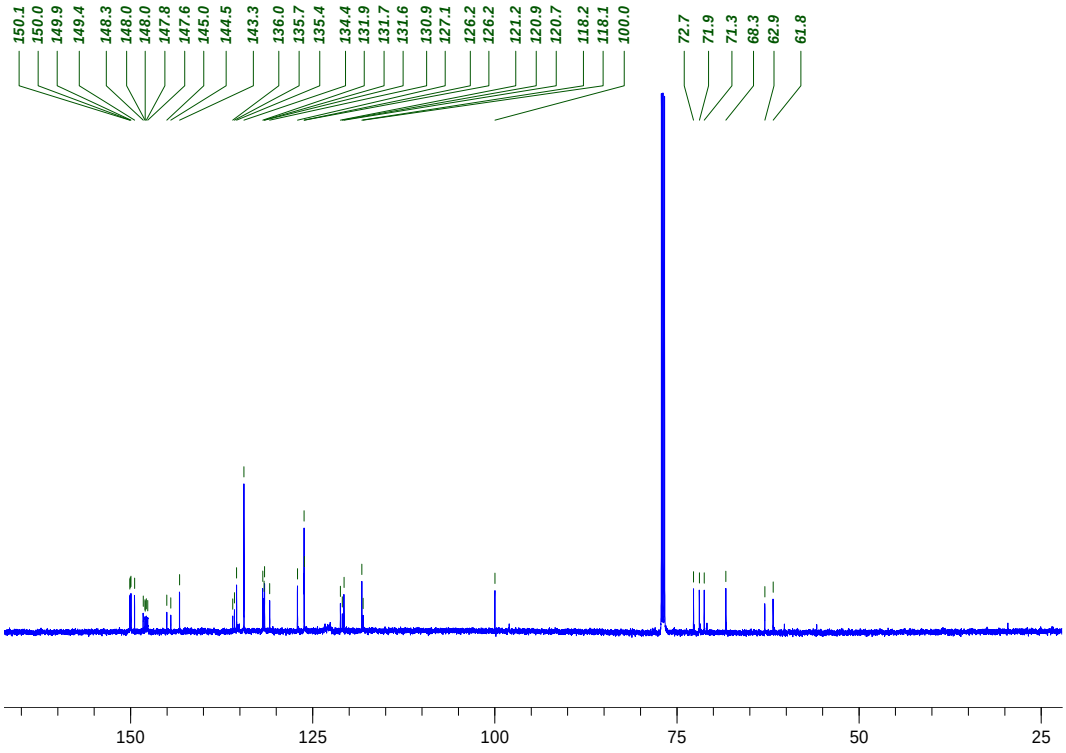


12 mg of porphyrin **12** (0.025 mmol), 1 mg CuCl (0.007 mmol), 48 mg of glucose **2** (0.120 mmol) were added to a 10 mL microwave tube and dissolved in 1.25 mL toluene/H_2O (4:1). The tube was then sealed and heated to 140°C in a microwave reactor for 20 mins. TLC analyses after the 20 minutes were performed in both CH_2Cl_2 and EtOAc/CH_2Cl_2 (2:1). The solvent was removed in vacuo and the residue was purified by column chromatography using CH_2Cl_2 as eluent initially to remove any porphyrin material other than product. The solvent was subsequently changed to EtOAc/CH_2Cl_2 (5:2) and the desired glycoporphyrin was isolated as a bright purple shiny solid (18 mg, 50%).^{1}H NMR (400MHz, $CDCl_3$): 8.96 (8H, s, 2,3,7,8,12,13, 17,18-β-H), 8.41 (8H, d, J = 8.2, o-Ph-H), 8.32 (4H, s, 5-triazole-H), 8.16 (8H, J = 8.2, m-Ph-H), 5.31 (4H, t, J = 9.5, H-3), 5.19 (4H, t, J = 9.5, H-4), 5.18 (4H, d, J = 12.5, OCH_2), 5.16 (4H, dd, J = 9.5, 7.9, H-2), 5.07 (4H, d, J = 12.5, OCH_2), 4.87 (4H, d, J = 7.9, H-1), 4.37 (4H, dd, J = 4.8, 2J 12.3, H-6), 4.27 (4H, dd, J = 2.3, J = 12.3, H-6'), 3.83 (4H, ddd, J = 10.0, 4.7, 2.3, H-5), 2.16 (12H, s, CH_3), 2.12 (12H, s, CH_3), 2.08 (12H, s, CH_3), 2.06 (12H, s, CH_3); ^{13}C NMR (100 MHz, $CDCl_3$): 170.5, 170.0, 169.3, 149.8, 145.1, 143.9, 136.2, 135.4 (o-Ph-C), 131.7 (β-C), 121.1 (5-triazole-C)), 119.1, 118.4 (m-Ph-C), 100.0 (C-1), 72.6 (C-3), 71.9 (C-5), 71.2 (C-2), 68.2 (C-4), 63.0 (CH_2), 61.8 (C-6), 20.6 (CH_3), 20.4 (CH_3); IR: 2921 (m, C-H stretch), 2852 (m, C-H stretch), 1744 (s, C=O stretch), MS (MALDI-TOF): Calculated for $C_{112}H_{112}N_{16}O_{40}Zn$ M^+ 2384.6513; Found 2384.6604.

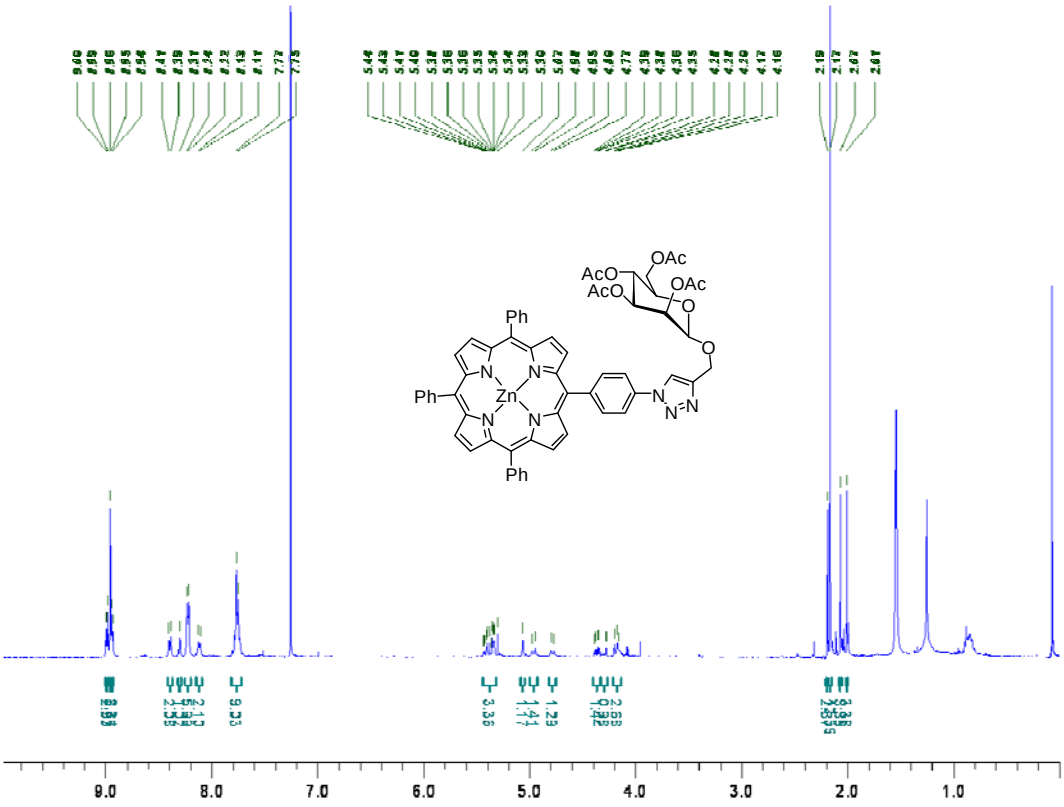
Porphyrin **5** ^{1}H Spectrum



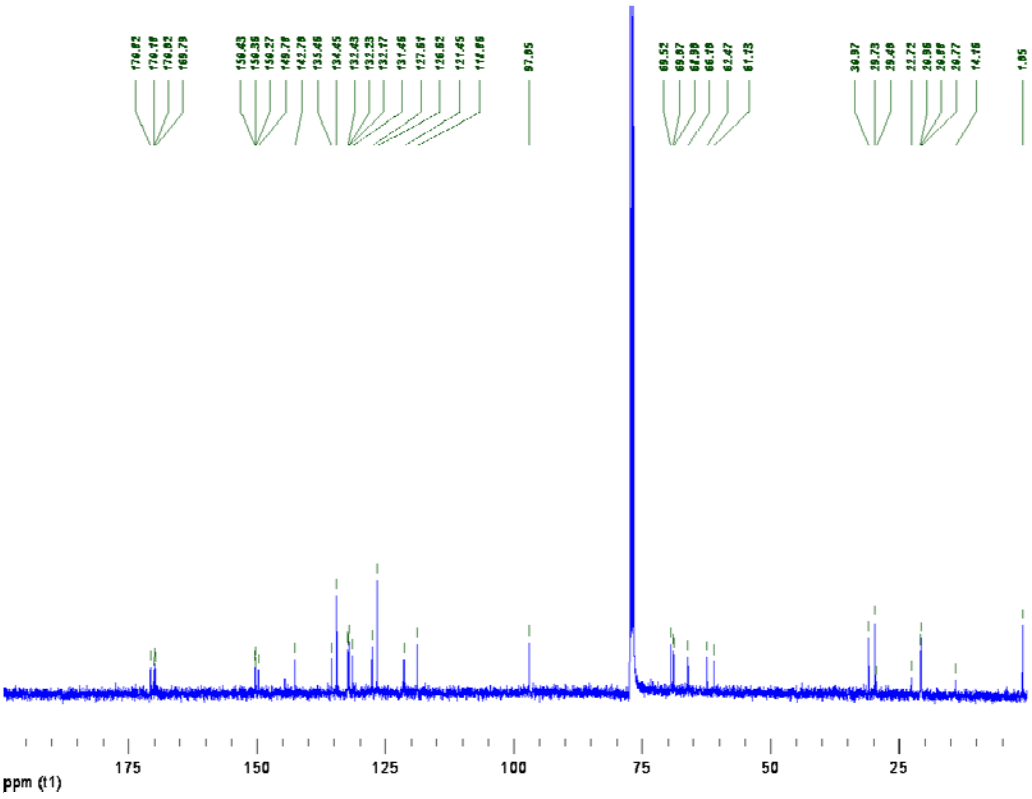
^{13}C Spectrum



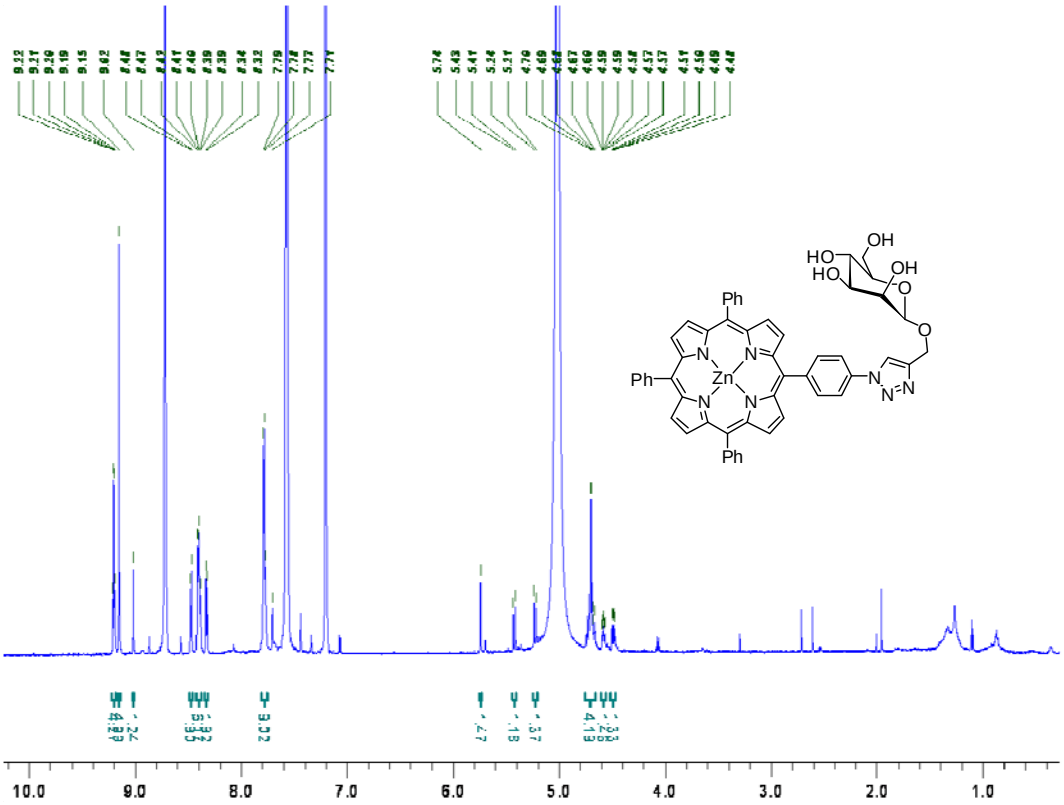
Porphyrin **6** ^{1}H Spectrum



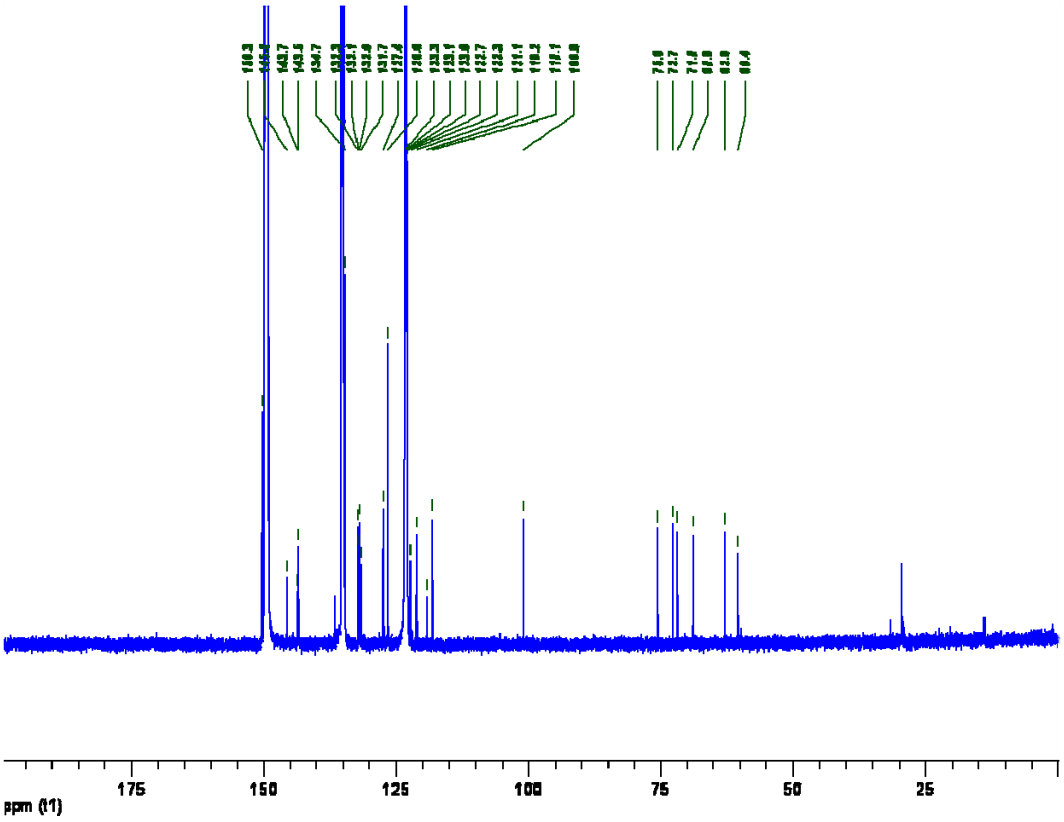
^{13}C Spectrum



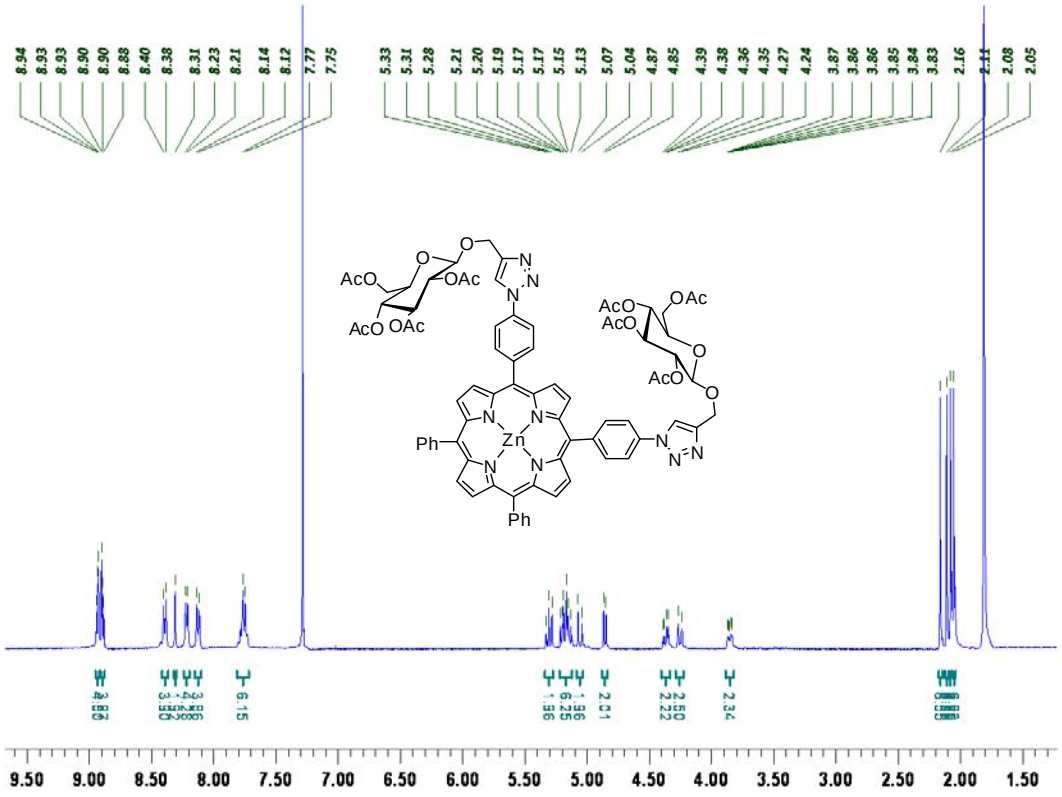
Porphyrin **7** ^{1}H Spectrum



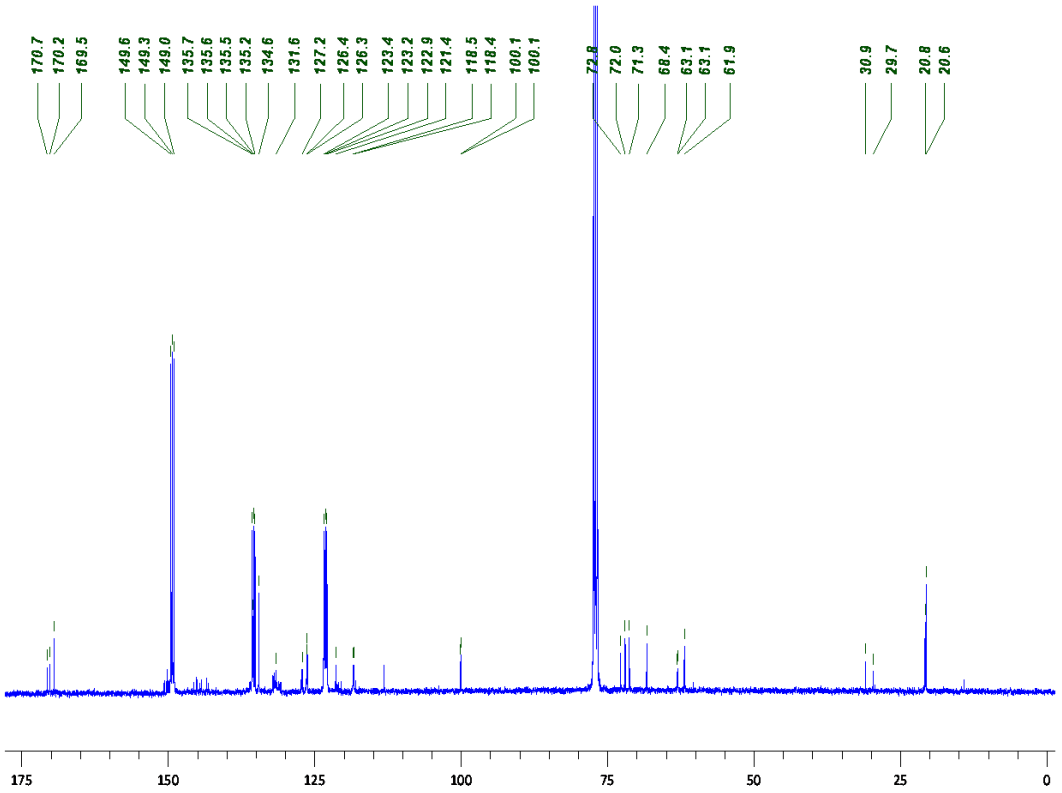
^{13}C Spectrum



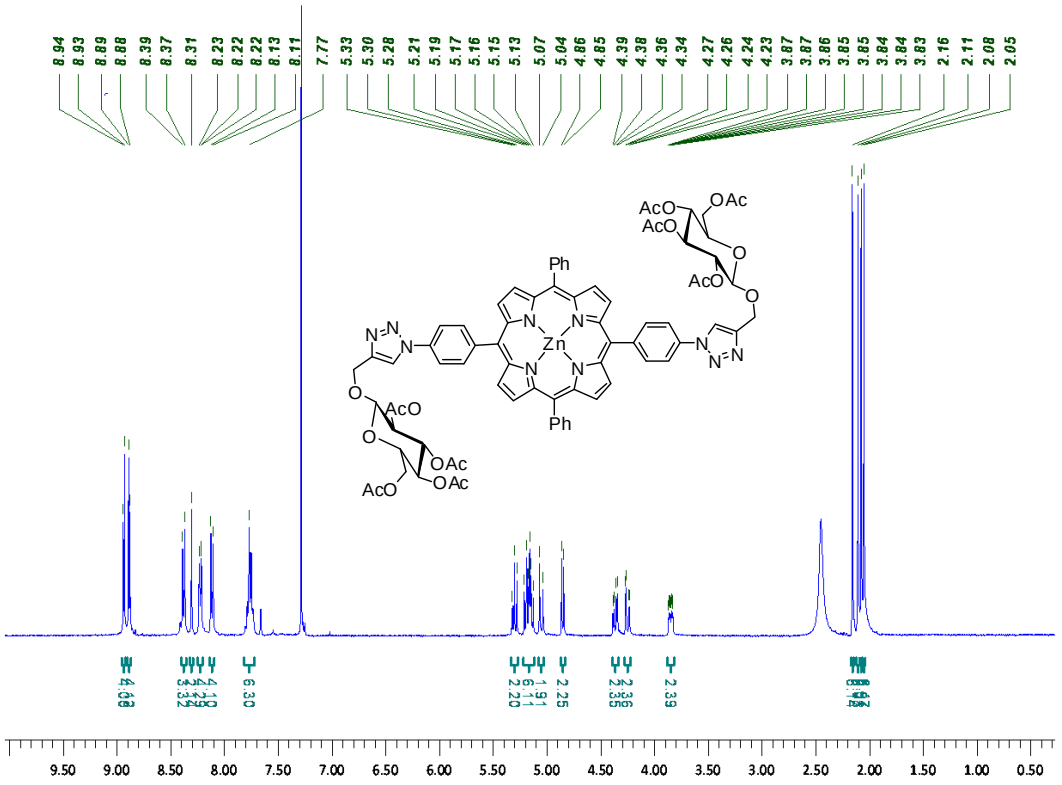
Porphyrin **8** ^{1}H Spectrum



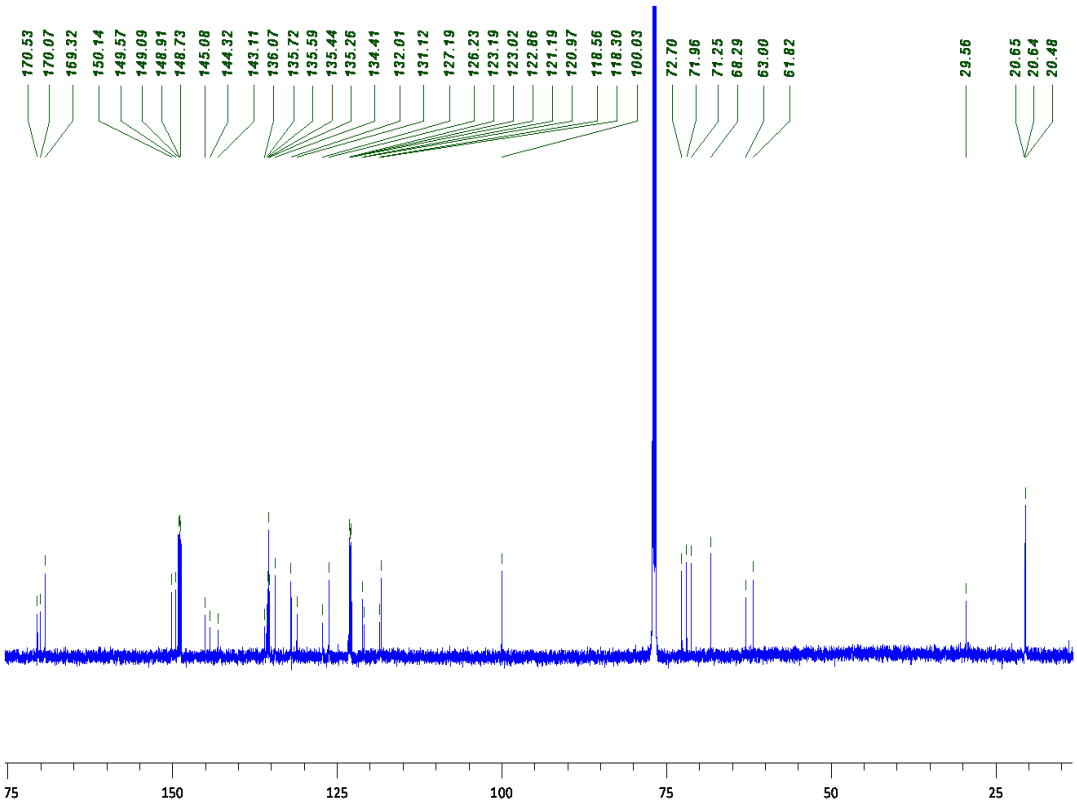
^{13}C Spectrum



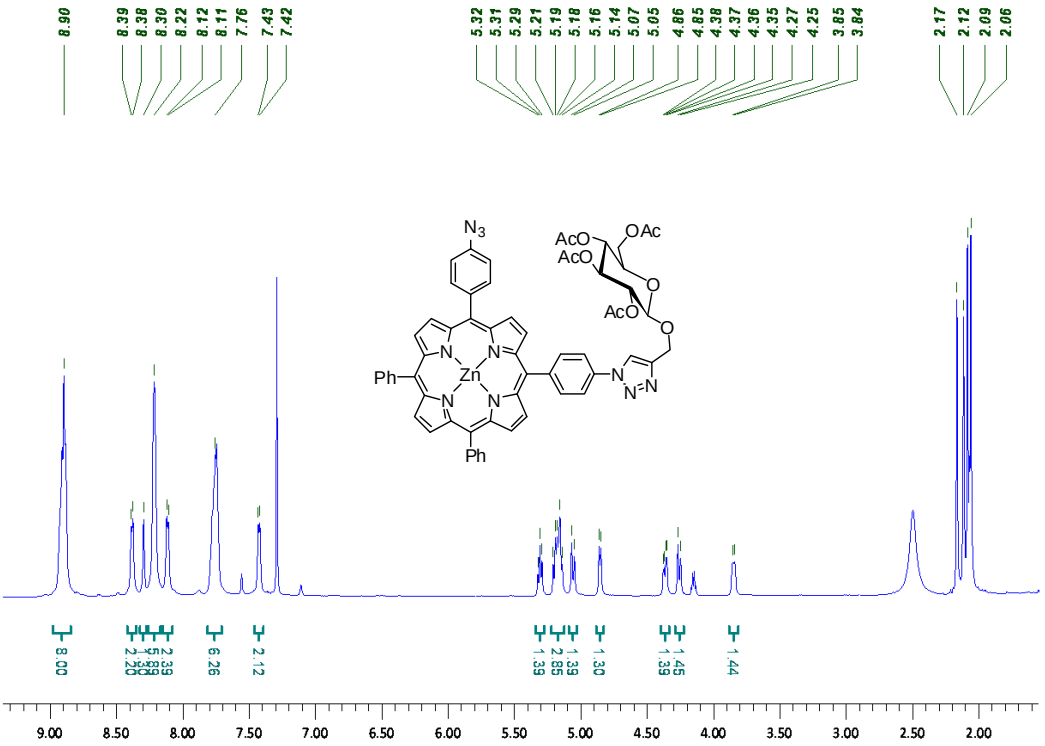
Porphyrin **9** ^{1}H Spectrum



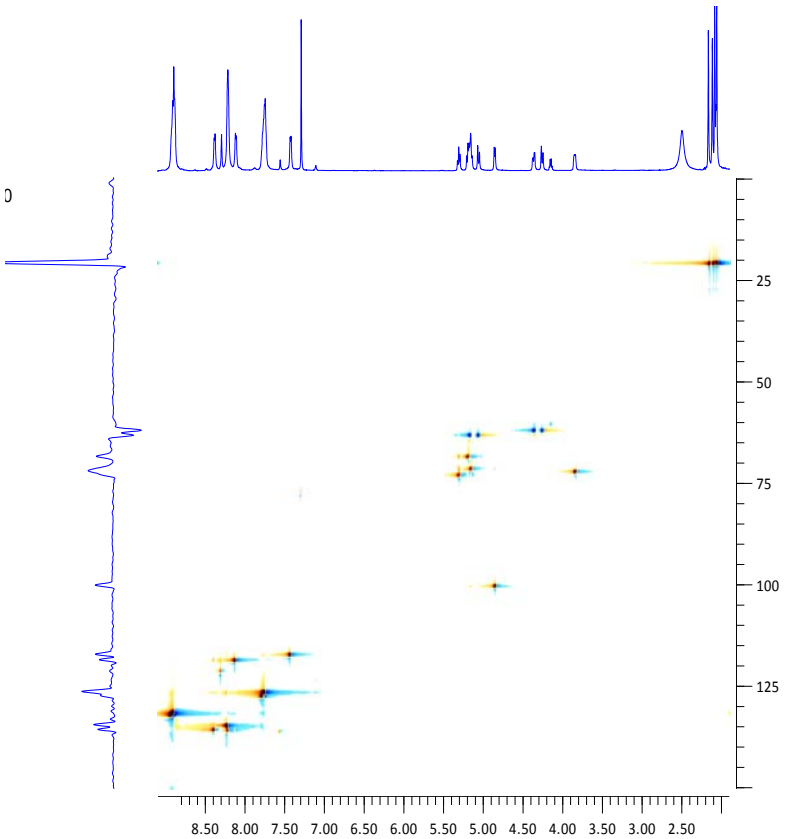
^{13}C Spectrum



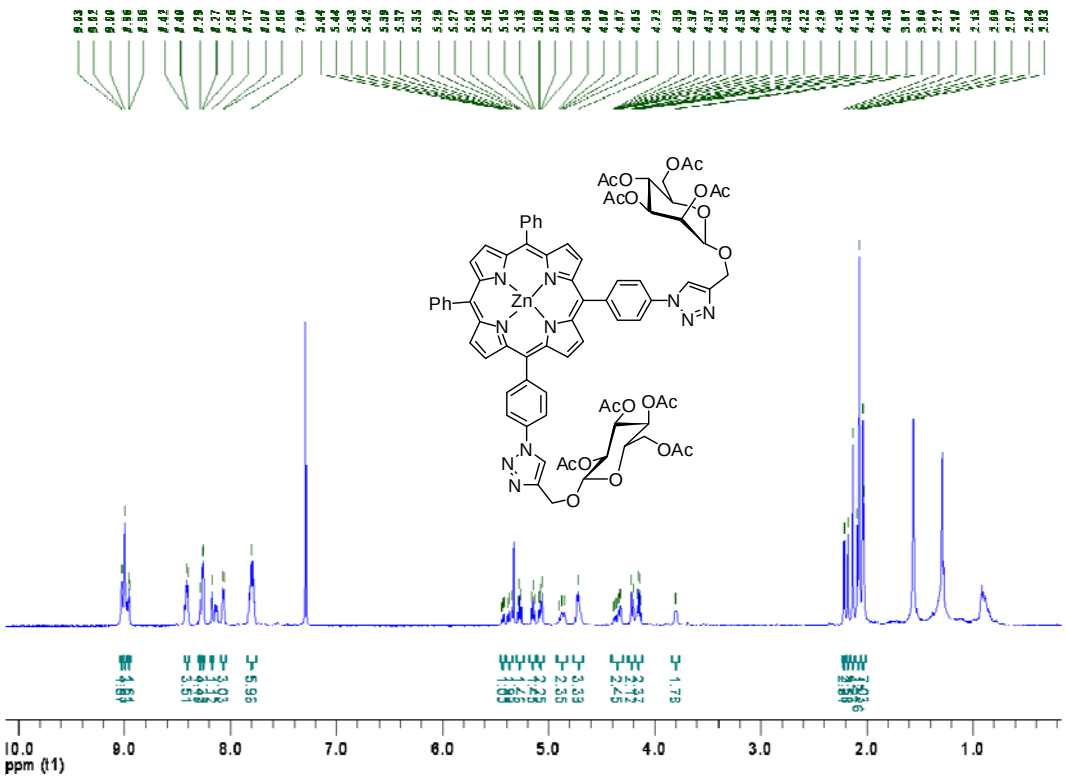
Porphyrin **11** ^{1}H Spectrum



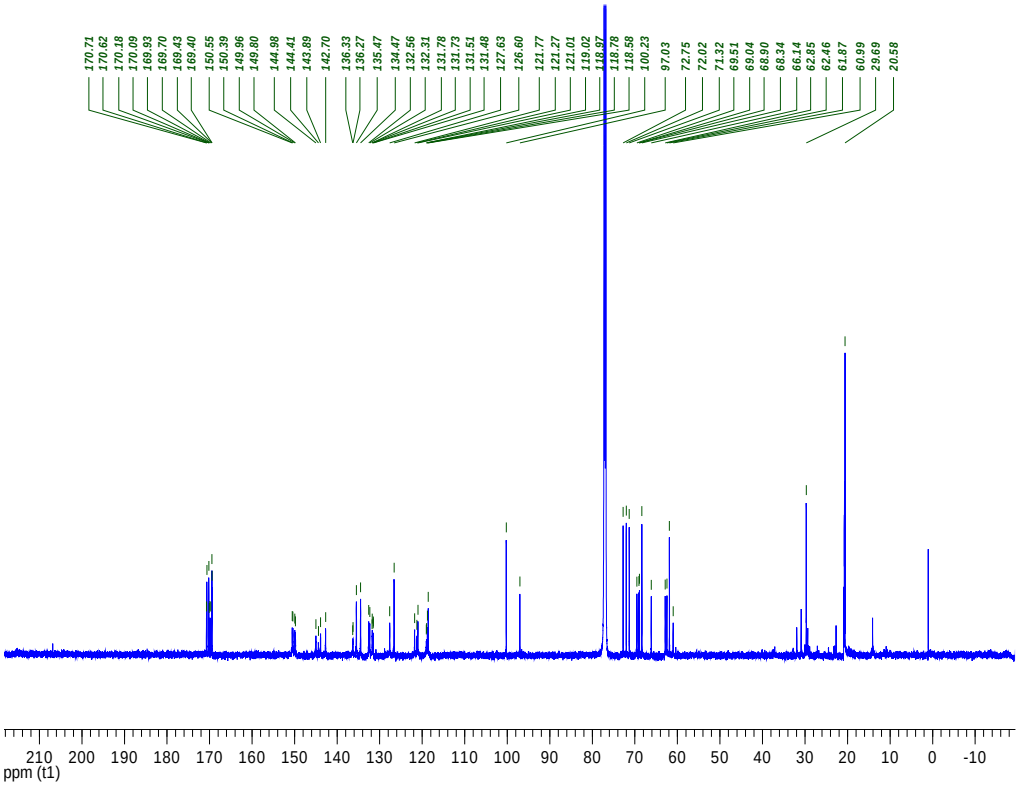
HSQC Spectrum



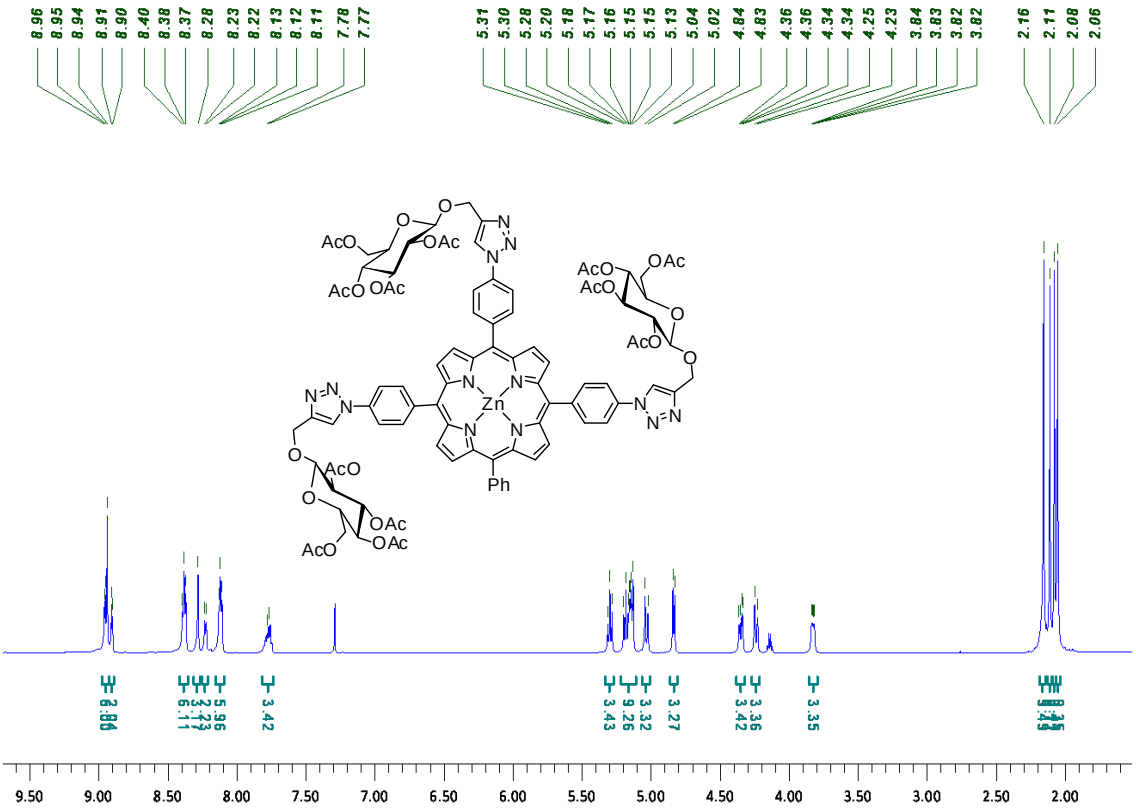
Porphyrin **12** ^{1}H Spectrum



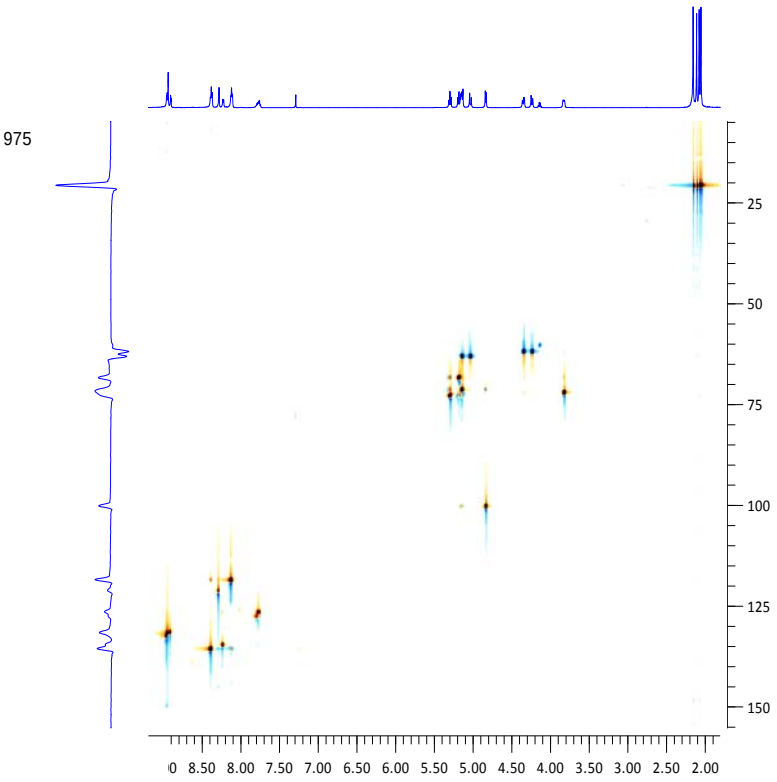
^{13}C Spectrum\



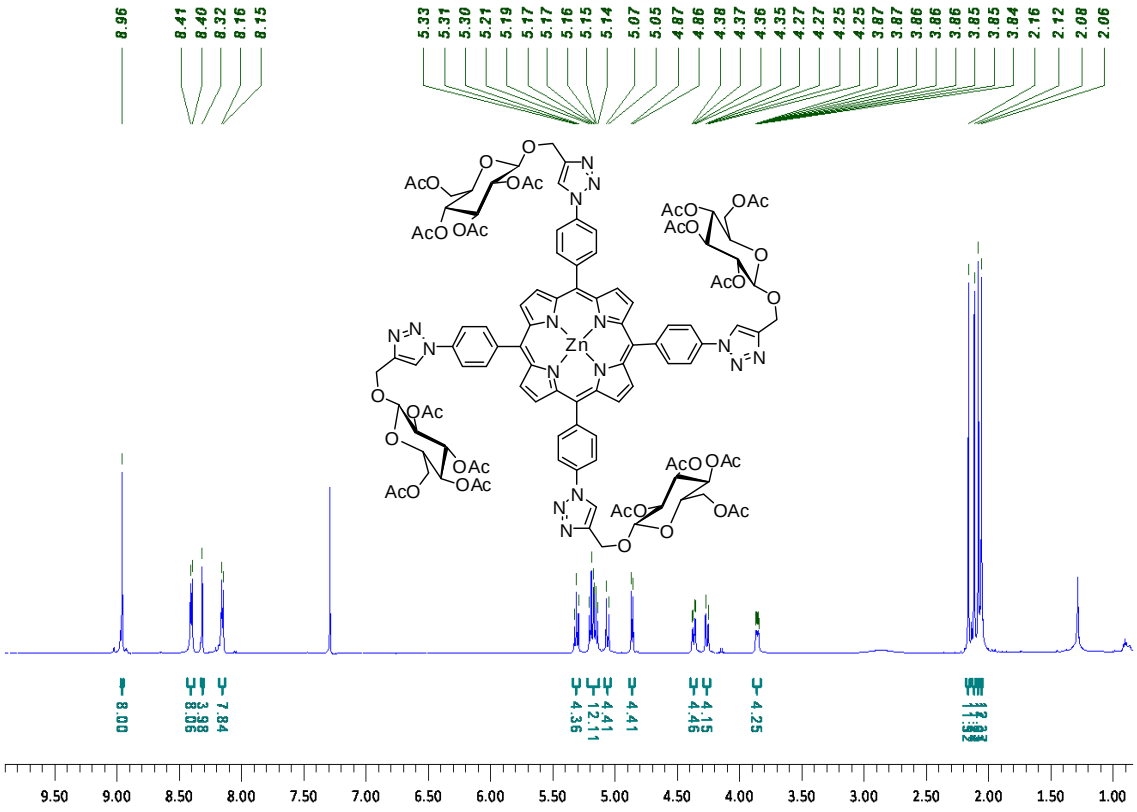
Porphyrin **15** ^{1}H Spectrum



HSQC Spectrum



Porphyrin **16** ^{1}H Spectrum



^{13}C Spectrum