Cite this: *Chem. Commun.*, 2014, 50, 10819

Received 26th May 2014,
Accepted 25th July 2014

DOI: 10.1039/c4cc04028g

www.rsc.org/chemcomm

Supramolecular pyridyl urea gels as soft matter with antibacterial properties against MRSA and/or *E. coli*†

Komala Pandurangan,^a Jonathan A. Kitchen,^{ab} Salvador Blasco,^a Francesca Paradisi^c and Thorfinnur Gunnlaugsson^{*a}

The synthesis and characterisation of novel aryl-pyridyl ureas are described, which form self-assembly structures via extended hydrogen bonding and π - π interactions in the solid state, and in selected cases, form supramolecular gels with antimicrobial properties against *Staphylococcus aureus* and/or *Escherichia coli*.

Hydrogen bonding is a powerful structural interaction that is commonly used in the synthetic self-assembly of supramolecular structures.^{1,2} Ureas and thioureas are particularly suited for such synthesis, providing two directional hydrogen bond donors and an acceptor.¹⁻⁴ Being easily accessible synthetically, they can adopt either the *syn* or the *anti* conformation^{5,6} and are known to have high affinity for anions.⁷⁻¹¹ Urea/thiourea structures have also been found to play a major role in human biology and physiology and they are also of industrial value; an example is polyurea, an elastomer that is often used for protective coating. It is also well known that hydrogen bonding structures¹² like ureas¹³ and polyurea derivatives¹⁴ can possess antibacterial properties against strains such as Gram(+) (e.g. *Bacillus subtilis* and *Staphylococcus aureus*) and Gram(−) (e.g. *Escherichia coli* and *Salmonella typhi*) bacteria.

Gels are highly versatile materials^{1,2} with properties suited for use in a variety of applications.^{15a} This includes their use in drug delivery,^{15b} but polymeric hydrogels have been shown to be suitable for coating implants with the view of preventing surface bacterial growth.^{15c} In an extensive body of work, Steed *et al.*^{11,15} have recently developed examples of pyridyl based ureas and studied their supramolecular properties in detail. This has included the study of their ability to form soft materials such as supramolecular gels^{1,2} and ligands for the formation of metal-organic frameworks (MOFs).¹⁶

Similarly, Wu *et al.*,¹⁷ Das *et al.*,¹⁸ Hay *et al.*,¹⁹ and Nangia *et al.*⁴ have also studied the use of pyridyl ureas in such supramolecular applications. Inspired by this work, we set out to develop new families of simple pyridyl urea and thiosemicarbazide²⁰ derivatives, with the view of exploring their gelation properties and use for sensing and biomedical applications: an area that is fast growing within supramolecular chemistry.^{21,22} Herein we present three families of novel aryl pyridyl ureas, some of which function as supramolecular gelators possessing antibacterial properties.

Compounds **1–12**, Fig. 1, were synthesised in a single step each from commercially available reagents, by reacting the corresponding 2-, 3- or 4-pyridyl amine with 4-fluorophenyl-, 2,4-difluorophenyl-, 2-nitrophenyl- and 4-trifluoromethylphenylisocyanates under microwave conditions for 50 minutes at 100 °C in CH₃CN. All the products were fully characterised and for all, the ¹H NMR spectrum in DMSO-*d*₆ clearly showed the urea proton resonances as broad signals occurring between 8 and 11 ppm (see ESI†).

Steed *et al.* have demonstrated that urea-pyridyl structures tend to crystallise to give urea NH...N_{pyridyl} hydrogen-bonding interactions.²³ Such intermolecular networks would also be expected to dominate in the case of the 3- and 4-pyridyl ureas **5–12**, developed herein, while in the case of **1–4**, the formation of hydrogen bonding dimers would be foreseen, consisting of both intermolecular and intramolecular hydrogen bonding interactions.^{2a,3}

Of the compounds developed herein, **1–3**, **5–7** and **10–12** gave crystals suitable for X-ray diffraction analysis upon crystallisation from CH₃CN, or CH₃CN/MeOH (**9** was reported by Wu *et al.*¹⁸). The crystal structures of compounds **2**, **6**, **12** and **7**, and

Fig. 1 The pyridyl ureas **1–12** developed in this study.

^a School of Chemistry and Trinity Biomedical Sciences Institute, Trinity College Dublin, Dublin 2, Ireland. E-mail: gunnlaut@tcd.ie; Tel: +353 1 896 3459

^b Chemistry, Faculty of Natural & Environmental Sciences, University of Southampton, Highfield, Southampton, SO17 1BJ, UK

^c School of Chemistry and Chemical Biology, University College Dublin, Dublin 4, Ireland

† Electronic supplementary information (ESI) available: Synthesis and characterisations; Fig. S1–S20. CCDC 1002407–1002415. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c4cc04028g

Fig. 2 The crystal structure of (a) centrosymmetric dimeric structure of **2**; (b) and (c): the Steed 'urea pyridyl synthon II' seen for **6** and **12**, respectively; (d) the modified Steed 'urea pyridyl synthon II' consisting of a single $\text{NH}\cdots\text{N}_{\text{pyridyl}}$ hydrogen bonding in **7**.

the intermolecular hydrogen bonding which takes place for these structures are shown in Fig. 2a–d, respectively (see ESI† for the remaining structures). All of the 2-pyridyl derivatives showed that the urea protons adopt an *anti* conformation and, as a result, a dimer is formed with both moieties oriented in opposite directions related by a centre of inversion. In the case of **1–3**, the expected dimer shows two intramolecular and two intermolecular hydrogen bonds. For **2**, Fig. 2a, the intermolecular hydrogen bonding arises between the NH urea proton adjacent to the pyridyl unit and oxygen atom of the urea carbonyl with a bond distance of 2.843(2) Å. These $\text{NH}\cdots\text{O}$ interactions result in the formation of aforementioned dimeric structures in the solid state.^{23a} The intramolecular hydrogen bonding arises from the additional urea NH to the nitrogen atom of the pyridine. This is possible because the nitrogen atom of the pyridyl group is in an ideal position to accept a hydrogen bond from the NH urea group thus stabilising the *anti* conformation.^{23a} Moreover, the molecules pack with multiple and extended face-to-face and edge-to-face π -stacking interactions giving rise to a supramolecular polymeric network.

The crystal structure of **6**, Fig. 2b, demonstrated that the molecules adopt a planar conformation in the solid state, where the urea protons are in a *cis* conformation. These interact in an intermolecular fashion with the $\text{N}_{\text{pyridyl}}$ atom of an adjacent molecule through hydrogen bonding, which has been referred to by Steed *et al.*^{23a} as a 'urea pyridyl synthon II' geometry. Here, the classical urea α -tape motif was not observed, and each molecule is offset to the next. As in the case of **2**, the aryl fluorine atoms also play a significant role in the long-range interactions of the packing of **6**,²⁴ the overall packing being supported by face-to-face π -stacking (see ESI†). Compound **7**, Fig. 2d, also adopts a modified 'urea pyridyl synthon II' configuration, but different from that seen in **6** due to the presence of the nitro group. Here, a supramolecular hydrogen bonding polymer is also formed, which is extended into three dimensions through face-to-face and edge-to-face π - π interactions. However, unlike that in **6** and **12**, the $\text{N}_{\text{pyridyl}}$ moiety interacts only with the NH urea proton of the adjacent molecule; the (distal) NH-aryl proton is hydrogen bonded to the 2-nitro group in a six-membered ring system.

Fig. 3 (a) SEM imaging (scale bar at 10 μm) of an organogel formed after 24 h using 1% by weight of **1** in THF. (b) **1** upon gelation in: (1) 1% by weight in MeOH; (2) 1% by weight in a mixture of toluene : THF : CHCl_3 solution. (c) SEM image (scale bar at 2 μm) of AgNO_3 containing the gel of **1** (1% by weight) in 8 : 2 THF : H_2O mixture.

The ability of these structures to function as low molecular weight gelators was investigated in various solvents. The results show that for structures **1**, **6** and **7** all formed robust gels under a variety of solvents in 1% weight (see ESI†) that were stable towards a tube inversion tests. The gel formation was reversible, as upon heating, the solution phase was obtained, which after sonication reformed the gel. Several others formed self-assembly structures such as aggregates or suspensions in a variety of solvents and solvent mixtures, including compounds **3**, **8** and **12**. These, at 1% weight, did not observe the tube inversion tests, and hence, cannot be classified as robust gel-like materials. Fig. 3a shows the scanning electron microscopy (SEM) images of the organogel formed upon sonication of **1** (1% by weight) in THF solution, demonstrating the formation of fibres or a 'splinter' like morphology, as has been reported by Steed *et al.*^{2b} Both **6** and **7** gave rise to organogel formation in THF, as well as gel formation in a mixture of 8 : 2 THF : H_2O solution. While **1** did not give rise to the formation of a gel in such THF : H_2O mixture, the addition of AgNO_3 to a solution of **1** (1% by weight) in THF : H_2O generated a gel with a similar morphology to that shown in Fig. 3a (see ESI†), except for possessing a more fibrous texture, Fig. 3c. The three hydrogels were also doped with cresol red, an acid sensitive pH indicator.²⁵ This resulted in the formation of red coloured hydrogels (see ESI†). Upon exposure of these gels to HCl gas, the intermolecular hydrogen bonding interactions were disrupted, resulting in de-gelation, while upon exposure to ammonia, the gel remained intact, demonstrating no such hydrogen bonding disruption. These results also demonstrate that these gels are highly versatile, and can host other molecules within their fibrous network, which is currently under investigation in our laboratory.

It is reasonable to speculate that compounds **1–12** might possess antibacterial properties as hydrogen bonding plays a major role in the function of many antibacterial agents, such as vancomycin, which can inhibit bacterial cell wall synthesis by forming hydrogen bond interactions with the terminal D-alanyl-D-alanine moieties of the NAM/NAG-peptides. With this in mind, compounds **1–12** (the gels formed) were all tested for their antibacterial activities towards Gram(+) MRSA and Gram(−) *E. coli* strains using, in the case of the free compounds, a qualitative

Fig. 4 (a) Kirby–Bauer disk diffusion susceptibility test results for compounds **12** (top) and **11** (bottom, showing no clearance) using MRSA (A) 0.72 μmol , (B) 1.8 μmol and (C) 3.6 μmol of **12**, respectively. (b) Liquid turbidity test studies for gels **1** and **6** using *E. coli* demonstrating that bacterial growth is inhibited over 18 hours.

Kirby–Bauer disk diffusion method or in the case of the gels, a liquid turbidity test. The areas of clearance were measured for compounds **1–12** (see ESI†) to compare relative activities. Of these, **9** and **12** gave the most interesting and promising results, where **9** showed good activity against both strains, while **12** showed more activity towards MRSA after 18 hours of incubation at 37 °C, Fig. 4, at three different concentrations (denoted as A to C in Fig. 4). The Kirby–Bauer disk diffusion results for the 2-nitro substituted analogue **11** are also shown in Fig. 4a.

Interestingly, of the three-pyridyl families (*i.e.* **1–4**; **5–8** and **9–12**), compounds **1–4** were seen to have the least activity (area of clearance of 7 mm, 6 mm and 10 mm when 4.3 μmol , 3.9 μmol , and 3.6 μmol , were applied to the disc for **1**, **3** and **4** using MRSA), displaying a moderate selectivity in most cases for MRSA, while the 3-pyridine analogues **7** and **8** showed moderate activity towards *E. coli* both resulting in an area of clearance of 8 mm and 7 mm. However, and as demonstrated for **12** in Fig. 4a, the 4-pyridyl family gave the most potent activity [*e.g.* **12** area of clearance of 13 mm (1.8 μmol) and 20 mm (3.6 μmol) for MRSA]. In addition to these experiments, we also carried out Alamar blue viability assays on these compounds using HeLa cells. These showed that the compounds were either not toxic or only slightly toxic. Hence, we were unable to determine EC50 values for these.

Having demonstrated that these structurally simple pyridyl ureas could function as antibacterial agents, we next investigated the abilities of the hydro- and organogels for such activity using the aforementioned liquid turbidity test. The results demonstrated that in the gel form, all were able to inhibit bacterial growth over 16–18 hours. Even compound **1**, which showed moderate activity (MIC = 4.3) prevented bacterial growth for both MRSA and *E. coli* as demonstrated in Fig. 4b, which also shows the bacterial growth in a solution (as cloudy solution) in a control experiment after 18 hours. These results clearly demonstrate that the supramolecular nature of the systems, *i.e.* the formation of a self-assembly gel, changes the antibacterial properties of the pyridyl urea structures. While this phenomenon is not well understood, it is possible that the overall supramolecular structure can give rise to multiple hydrogen bonding interactions that can interact with the bacterial cell wall and possibly disrupt it or prevent its formation. Such properties are highly desirable, particularly as the soft-matter is easily applicable and as such highly attractive

for use in coating implants to prevent the onset of bacterial infection.^{15,26} Self-assembled soft materials are good candidates for application in therapeutic delivery²⁶ as they can be easily moulded, shaped and made to be responsive to external stimuli, such as pH.²⁵ Since the above urea gels were reversibly formed^{1b} we anticipated that our systems could also ‘naturally’ degrade with time, for instance, upon interactions with biological anions, such as carboxylates,^{2b} which could competitively interrupt the hydrogen bonding networks within the gels. This we demonstrated by subjecting the gels formed from **6**, and the AgNO₃ based gel of **1**, to a solution of acetate. These showed that while the degradation was initially slow, the gels indeed ‘dissolved’ over a period of days. As these gels can host other organic substrates (*e.g.* cresol red), this result potentially also allows for their application as drug delivery systems; whereupon such degradation, substrates such as known antibacterial drugs can be released. We are currently investigating the formation of such dual functional gels and their antimicrobial properties in greater detail.

We thank the Science Foundation Ireland (SFI 2008 RFP, 2010 PI Awards and ISAC Indian Initiative), FEMS TCD Indian Scheme (KP), and Marie Curie IEF Postdoctoral Grant (SB) for financial support. We thank Prof. Uday Maitra (IISc Bangalore) and Prof. D. Clive Williams (TCD) for their support and encouragement, Dr Neal Leddy, TCD CMA, for assisting with SEM imaging, and Joseph Byrne (TCD) and Sam Bradberry (TCD) for their help.

Notes and references

- (a) L. Meazza, J. A. Foster, K. Fucke, P. Metrangolo, G. Resnati and J. W. Steed, *Nat. Chem.*, 2013, **5**, 42–47; (b) A. Nangia, *J. Chem. Sci.*, 2010, **122**, 295–310; (c) A. R. Hirst, B. Escuder, J. F. Miravet and D. K. Smith, *Angew. Chem., Int. Ed.*, 2008, **47**, 8002–8018.
- (a) M.-O. M. Piepenbrock, G. O. Lloyd, N. Clarke and J. W. Steed, *Chem. Rev.*, 2010, **110**, 1960–2004; (b) J. A. Foster, M.-O. M. Piepenbrock, G. O. Lloyd, N. Clarke, J. A. K. Howard and J. W. Steed, *Nat. Chem.*, 2010, **2**, 1037–1043.
- M. C. Etter, *Acc. Chem. Res.*, 1990, **23**, 120–126.
- (a) L. S. Reddy, S. K. Chandran, S. George, N. J. Babu and A. Nangia, *Cryst. Growth Des.*, 2007, **7**, 2675–2690; (b) L. S. Reddy, S. Basavoju, V. R. Vangala and A. Nangia, *Cryst. Growth Des.*, 2006, **6**, 161–173; (c) S. K. Chandran, R. Thakuria and A. Nangia, *CrystEngComm*, 2008, **10**, 1891–1898.
- S. Boileau, L. Bouteiller, F. Laupretre and F. Lortie, *New J. Chem.*, 2000, **24**, 845–848.
- (a) J. R. Hiscock, P. A. Gale, N. Lalaoui, M. E. Light and N. J. Wells, *Org. Biomol. Chem.*, 2012, **10**, 7780–7788; (b) P. A. Gale, J. R. Hiscock, N. Lalaoui, M. E. Light, N. J. Wells and M. Wenzel, *Org. Biomol. Chem.*, 2012, **10**, 5909–5915.
- (a) R. M. Duke, E. B. Veale, F. M. Pfeffer, P. E. Kruger and T. Gunnlaugsson, *Chem. Soc. Rev.*, 2010, **39**, 3936–3959; (b) P. A. Gale and T. Gunnlaugsson, *Chem. Soc. Rev.*, 2010, **39**, 3595–3596.
- T. Gunnlaugsson, P. E. Kruger, P. Jensen, J. Tierney, H. D. P. Ali and G. M. Hussey, *J. Org. Chem.*, 2005, **70**, 10875–10878.
- (a) E. M. Boyle, S. Comby, J. K. Molloy and T. Gunnlaugsson, *J. Org. Chem.*, 2013, **78**, 8312–8319; (b) R. M. Duke, T. McCabe, W. Schmitt and T. Gunnlaugsson, *J. Org. Chem.*, 2012, **77**, 3115–3126; (c) E. B. Veale, G. M. Tocci, F. M. Pfeffer, P. E. Kruger and T. Gunnlaugsson, *Org. Biomol. Chem.*, 2009, **7**, 3447–3454; (d) F. M. Pfeffer, P. E. Kruger and T. Gunnlaugsson, *Org. Biomol. Chem.*, 2007, **5**, 1894–1902; (e) C. M. G. dos Santos, T. McCabe and T. Gunnlaugsson, *Tetrahedron Lett.*, 2007, **48**, 3135–3139.
- (a) S. J. Moore, M. Wenzel, M. E. Light, R. Morley, S. J. Bradberry, P. Gómez-Iglesias, V. Soto-Cerrato, R. Pérez-Tomás and P. A. Gale, *Chem. Sci.*, 2012, **3**, 2501–2509; (b) N. Busschaert, S. J. Bradberry, M. Wenzel, C. J. E. Haynes, J. R. Hiscock, I. L. Kirby, L. E. Karagiannidis, S. J. Moore, N. J. Wells, J. Herniman, G. J. Langley, P. N. Horton, M. E. Light,

- I. Marques, P. J. Costa, V. Félix, J. G. Frey and P. A. Gale, *Chem. Sci.*, 2013, **4**, 3036–3045.
- 11 (a) J. W. Steed, *Chem. Soc. Rev.*, 2010, **39**, 3686–3699; (b) G. O. Lloyd and J. W. Steed, *Nat. Chem.*, 2009, **1**, 437–442; (c) L. Fischer and G. Guichard, *Org. Biomol. Chem.*, 2010, **8**, 3101–3117; (d) H. Maeda, *Chem. – Eur. J.*, 2008, **14**, 11274–11282.
- 12 L. C. Henderson, J. Li, R. L. Nation, T. Velkov and F. M. Pfeffer, *Chem. Commun.*, 2010, **46**, 3197–3199.
- 13 G. S. Basarab, J. I. Manchester, S. Bist, P. A. Boriack-Sjodin, B. Dangel, R. Illingworth, B. A. Sherer, S. Sriram, M. Uria-Nickelsen and A. E. Eakin, *J. Med. Chem.*, 2013, **56**, 8712–8735.
- 14 (a) E. R. Kenawy, S. D. Worley and R. Broughton, *Biomacromolecules*, 2007, **8**, 1359–1384; (b) N. Nishat, T. Ahamad, M. Zulfequar and S. Hasnain, *J. Appl. Polym. Sci.*, 2008, **110**, 3305–3312; (c) T. Ahamad, V. Kumar and N. Nishat, *J. Biomed. Mater. Res., Part A*, 2009, **88**, 288–294.
- 15 (a) *Functional Molecular Gels*, ed. B. Escuder and J. F. Miravet, Royal Society Chemistry, Cambridge, UK, 2014; (b) A. Friggeri, B. L. Feringa and J. van Esch, *J. Controlled Release*, 2004, **97**, 241–248; (c) C. P. McCoy, C. Brady, J. F. Cowley, S. M. McGlinchey, N. McGoldrick, D. J. Kinnear, G. P. Andrews and D. S. Jones, *Expert Opin. Drug Delivery*, 2010, **7**, 605–616.
- 16 (a) J. W. Steed, *Chem. Commun.*, 2011, **47**, 1379–1383; (b) J. T. Lenthall, J. A. Foster, K. M. Anderson, M. R. Probert, J. A. K. Howard and J. W. Steed, *CrystEngComm*, 2011, **13**, 3202–3212; (c) P. Byrne, G. O. Lloyd, L. Applegarth, K. M. Anderson, N. Clarke and J. W. Steed, *New J. Chem.*, 2010, **34**, 2261–2274.
- 17 B. Wu, X. Huang, Y. Xia, X.-J. Yang and C. Janiak, *CrystEngComm*, 2007, **9**, 676–685.
- 18 D. K. Kumar, D. A. Jose, A. Das and P. Dastidar, *Chem. Commun.*, 2005, 4059–4061.
- 19 R. Custelcean, B. A. Moyer, V. S. Bryantsev and B. P. Hay, *Cryst. Growth Des.*, 2006, **6**, 555–563.
- 20 K. Pandurangan, J. A. Kitchen, T. McCabe and T. Gunnlaugsson, *CrystEngComm*, 2013, **15**, 1421–1431.
- 21 (a) V. S. Sajisha and U. Maitra, *Chimia*, 2013, **67**, 44–50; (b) A. Chakrabarty and U. Maitra, *J. Phys. Chem. B*, 2013, **117**, 8039–8046; (c) S. Banerjee, R. Kandaneli, S. Bhowmik and U. Maitra, *Soft Matter*, 2012, **7**, 8207–8215; (d) S. Bhowmik, S. Banerjee and U. Maitra, *Chem. Commun.*, 2010, **46**, 8642–8644.
- 22 (a) R. Daly, O. Kotova, M. Boese, T. Gunnlaugsson and J. J. Boland, *ACS Nano*, 2013, **7**, 4838–4845; (b) O. Kotova, R. Daly, C. M. G. dos Santos, M. Boese, P. E. Kruger, J. J. Boland and T. Gunnlaugsson, *Angew. Chem., Int. Ed.*, 2012, **51**, 7208–7212.
- 23 (a) P. Byrne, D. R. Turner, G. O. Lloyd, N. Clarke and J. W. Steed, *Cryst. Growth Des.*, 2008, **8**, 3335–3344; (b) A. M. Todd, K. M. Anderson, P. Byrne, A. E. Goeta and J. W. Steed, *Cryst. Growth Des.*, 2006, **6**, 1750–1752.
- 24 A. Abad, C. Agulló, A. C. Cuñat, C. Vilanova and M. C. Ramírez de Arellano, *Cryst. Growth Des.*, 2006, **6**, 46–47.
- 25 U. Maitra and A. Chakrabarty, *Beilstein J. Org. Chem.*, 2011, **7**, 304–309.
- 26 (a) A. M. Klibanov, *J. Mater. Chem.*, 2007, **17**, 2479–2482; (b) P. Li, Y. F. Poon, W. Li, H.-Y. Zhu, S. H. Yeap, Y. Cao, X. Qi, C. Zhou, M. Lamrani, R. W. Beuerman, E.-T. Kang, Y. Mu, C. M. Li, M. W. Chang, S. S. J. Leong and M. B. Chan-Park, *Nat. Mater.*, 2011, **10**, 149–156.